

Rethinking the external globus pallidus and information flow in cortico-basal ganglia-thalamic circuits

Cristina Giossi¹ | Jonathan E. Rubin Ph.D.² | Aryn Gittis Ph.D.³ | Timothy Verstynen Ph.D.^{4*} | Catalina Vich Ph.D.^{1*}

¹Dept. de Matemàtiques i Informàtica and Institute of Applied Computing and Community Code, Universitat de les Illes Balears, Palma, Illes Balears, Spain.

²Dept. of Mathematics and Center for the Neural Basis of Cognition, University of Pittsburgh, Pittsburgh, PA, USA.

³Dept. of Biology, Carnegie Mellon University, and Center for the Neural Basis of Cognition, University of Pittsburgh, Pittsburgh, PA, USA.

⁴Dept. of Psychology, Carnegie Mellon University, and Center for the Neural Basis of Cognition, University of Pittsburgh, Pittsburgh, PA, USA.

Correspondence

T. Verstynen and C. Vich

Email: timothyv@andrew.cmu.edu;

catalina.vich@uib.es

Funding information

For decades the external globus pallidus (GPe) has been viewed as a passive way-station in the indirect pathway of the cortico-basal ganglia-thalamic (CBGT) circuit, sandwiched between striatal inputs and basal ganglia outputs. According to this model, one-way descending striatal signals in the indirect pathway amplify the suppression of downstream thalamic nuclei by inhibiting GPe activity. Here we revisit this assumption, in light of new and emerging work on the cellular complexity, connectivity, and functional role of the GPe in behavior. We show how, according to this new circuit-level logic, the GPe is ideally positioned for relaying ascending and descending control signals within the basal ganglia. Focusing on the problem of inhibitory control, we illustrate how this bidirectional flow of information allows for the integration of reactive and proactive control mechanisms during action selection. Taken together, this new evidence points to the GPe as being a central hub in the CBGT circuit, participating in bidirectional information

Abbreviations: CBGT, cortico-basal ganglia-thalamic; dSPNs, direct pathway spiny projection neurons; GPe, external globus pallidus; GPe_A, GPe arkypallidal neurons; GPe_P, GPe prototypical neurons; iSPNs, indirect pathway spiny projection neurons; STN, subthalamic nucleus; Th, thalamus.

* Equally contributing authors.

flow and linking multifaceted control signals to regulate behavior.

KEYWORDS

arkypallidal, prototypical, inhibitory control, stop task.

1 | INTRODUCTION

Imagine walking up to a busy intersection, intending to cross the street. You look at the crosswalk sign and it indicates that it is not safe for you to enter the intersection, so you stop at the corner. The light turns green and the crosswalk sign indicates that it is now safe to proceed. You begin to step off the curb and into the intersection, but you sense something approaching quickly from your side. You stop and step back to the curb before you are even consciously aware of why you have stopped. Then a car shoots in front of you, running the red light and narrowly missing you.

This scenario highlights some of the multifaceted control processes that regulate our actions (Dunovan et al., 2015; Meyer and Bucci, 2016; Aron, 2011; Braver, 2012). The first stop on the approach to the corner reflects what is known as *proactive* inhibitory control. This form of control represents the cessation of action as an internally generated choice (Dunovan et al., 2015) and is often referred to as a “no go” response. The second stop, reflecting an unconscious reaction to the approaching car, illustrates *reactive* inhibitory control (Mallet et al., 2016; Wessel, 2018), sometimes referred to as “braking” or “stopping”. This is a faster, and less deliberative, form of inhibition that involves the termination of an ongoing or almost initiated motor plan in response to an external stimulus.

Both proactive and reactive inhibitory control appear to rely on common neural substrates known as the cortico-basal ganglia-thalamic (CBGT) pathways (Nambu et al., 2002). These distributed circuits are thought to function as loops that relay information from the cortex to subcortical pathways and back up to the same cortical areas, regulating the tone of cortical activity (Mink, 1996; Haber, 2003). Yet despite this reliance on a common circuit, the means of control for proactive and reactive inhibition have been thought to be largely independent within the CBGT network. In this view, proactive control relies primarily on the *indirect pathway*, which regulates inhibition of the thalamus via striatopallidal connections. Essentially the indirect pathway, which results in reduced CBGT feedback to cortex, has to “win” a competition against the *direct pathway* that runs as a parallel loop and pushes to increase thalamocortical excitatory drive (Dunovan and Verstynen, 2016). In contrast, reactive control has been thought to rely on the so-called *hyperdirect pathway*, bypassing the striatum and regulating inhibition of the thalamus via cortical input to the subthalamic nucleus and its excitatory impact on pallidal nuclei (Nambu et al., 2002). This model of control via the CBGT pathways relies on two critical assumptions: 1) information flows one-way through the CBGT circuits, and 2) the major pathways in these circuits do not interact with each other before the output stage of processing.

Recent discoveries over the past 15 years have called these two assumptions into question, however. In particular, new discoveries concerning one critical nucleus in the CBGT circuit, the external segment of the globus pallidus (GPe), reveal a rich intricacy of cell types and connections that are forcing us to reconsider how information flows through the CBGT network (Mallet et al., 2012; Dodson et al., 2015; Abecassis et al. 2020; Ketzef and Silberberg, 2021). Here we review the old and new literature on the GPe and its role in behavior, focusing on the process of inhibitory control. We begin by reviewing the classical stop signal task and models of reactive inhibition via the hyperdirect pathway. We then review recent discoveries about the complexity of the GPe, including novel cell types and connectivity patterns, as well as functional observations, both physiological and behavioral. From a circuit-level logic perspective, we show how these new discoveries point to the GPe as a central hub that relays ascending and descending control signals through

the CBGT circuit, linking proactive and reactive mechanisms together. We finish by highlighting future directions on which the field can focus, to flesh out the nature of the interaction of these pathways and their consequences for behavior and cognition.

2 | REACTIVE STOPPING AND THE CLASSICAL MODEL

2.1 | Reactive inhibition and the stop signal task

To illustrate the nature of information flow in the CBGT circuit, we will focus on the process of inhibitory control, particularly reactive inhibition. We make this choice for two reasons. First, reactive inhibition is one of the most well-studied paradigms in the context of CBGT circuits and cognition. Second, the classical model of how CBGT circuits implement reactive stopping relies on the two fundamental assumptions of CBGT circuit computation mentioned in Section 1: unidirectional information flow and independent control pathways.

As illustrated in our opening example by the ability to quickly withdraw from a crosswalk as a car approaches, reactive inhibition involves terminating an action or planned action in response to an external stimulus. The most popular paradigm for studying reactive inhibitory control is the classic stop signal reaction time task (Vince, 1948; Lappin and Eriksen, 1966). A systematic review of the task and its limitations is beyond this review's scope (we suggest (Verbruggen et al. 2019) for any interested readers). Here we provide a short overview of the general stop signal task paradigm in order to walk the reader through the CBGT computations involved.

In a typical stop signal task (Figure 1A), participants are asked to quickly respond to a primary stimulus (the "Go" cue; e.g., pressing a button when they see a green circle) but to withhold that response when they encounter a secondary "stop" signal (the "Stop" cue; e.g., a brief tone) that is presented a short time, usually a few hundred milliseconds, after the primary stimulus. The timeline between the "Go" cue and the "Stop" signal, known as the stop signal delay (SSD), is usually varied - either dynamically adapted based on the participant's performance or sampled at specific intervals. The participant's ability to stop when experiencing the different SSDs is used to estimate their stop signal reaction time (SSRT). The SSRT reflects the median time it takes to "react" to the stop signal and successfully inhibit the action.

The stop signal task has become a popular tool due to its sensitivity at detecting individual differences across a range of subject groups. In clinical populations, elevated SSRTs (indicating poorer inhibitory control) have been associated with various pathologies such as attention-deficit/hyperactivity disorder (Alderson et al., 2007), substance abuse disorders (Fillmore and Rush, 2002) and obsessive-compulsive disorder (Chamberlain et al., 2006). Such findings suggest that compromised reactive inhibitory control may play a role in the etiology or maintenance of these disorders. In non-clinical populations, SSRTs have been linked to individual differences in personality traits like impulsivity (Logan et al., 1997) and have been used to investigate cognitive changes across the lifespan (Williams et al., 1999), with SSRTs appearing to initially shorten during early development and then lengthen with aging after adulthood. The popularity of the stop signal task, as well as its sensitivity at spotting individual differences in both clinical and non-clinical populations, makes it an ideal paradigm for describing the process of reactive inhibitory control. We now move on to consider traditional models for how the stop signal task is implemented in CBGT pathways.

2.2 | Classical model: Unidirectional flow

The classical model of CBGT inhibitory control in the stop signal task posits a one-way flow of information "downward" in the CBGT circuit, with proactive (striatal-initiated) and reactive (STN-initiated) control arising from independent

sources. According to this model (Aron and Poldrack, 2006), the “Go” stimulus triggers engagement of the *direct pathway*, starting with glutamatergic, excitatory signals from cortical regions to D1-expressing spiny projection neurons (SPNs) in the striatum. The direct pathway SPNs (dSPNs) then send GABAergic inhibition into the main output nucleus of the basal ganglia, which in primates is the internal globus pallidus (GPi). The GPi has tonically active neurons that send GABAergic, inhibitory projections into the matrix of the thalamus (Kita et al., 2005; Nambu et al., 2000). Thus, inhibiting the GPi increases the activity of the thalamus through disinhibition, potentially amplifying cortical activity via thalamocortical pathways and increasing the likelihood of a response.

Running parallel to the direct pathway is the *indirect pathway*. This pathway is also driven by excitatory signals from the cortex, but in this case, they terminate on D2-expressing SPNs. These indirect pathway SPNs (iSPNs) send GABAergic projections to the GPe. The GPe cells, in turn, inhibit both the subthalamic nucleus (STN) and the GPi, forming the so-called long and short indirect pathways, respectively. The net result of the engagement of both branches of the indirect pathway is the increased activity of GPi cells, which strengthens the suppression of their thalamic targets and reduces activity in the thalamocortical loops. These effects decrease the likelihood of a subsequent response.

A third canonical control pathway, known as the *hyperdirect pathway* (Nambu et al., 2002), runs through the CBGT circuit. This more recently discovered circuit bypasses the striatum altogether, by sending glutamatergic projections to the STN itself. Because the STN directly projects to GPi, this architecture provides a rapid, two synapses link from the cortex to the outputs of the basal ganglia, proposed as a faster control mechanism than the indirect pathway that can implement an urgent command to interrupt evolving action plans.

In the classical model of reactive stopping, only the hyperdirect pathway is thought to regulate reactive control (Nambu et al., 2002; Aron and Poldrack, 2006). Following the timeline of the stop signal task (Figure 1A), this model proposes that the “Go” cue initiates activation of the direct pathway (Figure 1B, left panel), thereby starting a drive process that, if left unchecked, will eventually lead to the triggering of a response via disinhibition of the thalamus. The presentation of the “Stop” cue activates the hyperdirect pathway (Figure 1B, right panel), which quickly boosts GPi activity and stop the action initiation process. If the hyperdirect pathway can sufficiently rapidly achieve and maintain the suppression of the thalamus, then a successful reactive stop occurs. Otherwise, the direct pathway succeeds, and an erroneous go response is produced.

The classical model of reactive control arose mainly from a combination of inferring function from the logic of the canonical CBGT circuit as understood at the time (see next section for more on this) (Aron et al., 2016), as well as correlational evidence of fast STN activation in response to a stop signal (e.g., (Aron and Poldrack, 2006; Chen et al., 2020; Sano et al., 2013; Wadsley et al., 2022)). Baked into this model are some critical assumptions. First, it assumed that the diffuse excitatory projections from STN to GPi are strong enough to prevent a significant reduction of GPi activity due to direct pathway inhibition. This assumption leads to the second assumption: information only flows down to the GPi within CBGT circuits. This unidirectional model of information flow means that reactive control, implemented via hyperdirect pathway signals, does not interact with more proactive control mechanisms, which involve a competition between the direct and indirect pathways. As we will soon show, these two assumptions have come into question as our understanding of the CBGT circuit, and particularly our knowledge about the GPe nucleus, has expanded in recent years. New discoveries have forced the field to rethink the circuit-level architecture of the CBGT pathways and how information flows through this circuit to contribute to behavioral control.

3 | EMERGING VIEWS OF THE GPE

While the subject of a recent surge in attention, aspects of the cellular complexity of the GPe, including the identification of prototypical and arky pallidal neuron subtypes, have been known for over 50 years (DeLong, 1971). Yet the functions of the various cell types in the GPe and their roles in guiding behavior were, until recently, largely overlooked. The GPe was essentially treated as a homogenous node in the CBGT circuit, with primarily a descending influence on information flow from the STN to the GPi. This simplified view changed with the advent of new methods that allowed researchers to study how GPe cells can be classified from a molecular perspective, as well as in terms of their electrophysiological properties, axonal projections, or dendritic morphology. Here we summarize the current understanding of the cellular composition, connectivity, and functional properties of this nucleus. This rundown allows us to lay out the foundation for rethinking the role of the GPe in CBGT circuit computation and behavioral control. For a more complete summary of the anatomy and physiology of the GPe we point the interested reader to (Dong et al., 2021) and (Courtney et al., 2023).

3.1 | Cellular composition

The GPe is mostly comprised of GABAergic neurons, with only about 5% of its cells being cholinergic (Abdi et al. 2015; Hernández et al., 2015; Abecassis et al. 2020; Mastro et al., 2014) (Figure 2A). Typically researchers focus on the non-cholinergic cells, characterizing the GPe in terms of two principal classes of inhibitory neurons: almost 70% of its neurons are labeled as *prototypical* (GPe_P in Figure 2), whereas *arkypallidal* neurons (GPe_A in Figure 2) represent approximately 20% of the neurons in the GPe. Prototypical cells themselves form a heterogeneous class of neurons. In some studies, they are labeled based on the expression of a specific calcium-binding protein, parvalbumin (PV), as a molecular marker (Mallet et al., 2012; Abdi et al. 2015). Others prefer to cluster prototypical cells based on the expression of transcription factors such as Nkx2.1 (NK2 homeobox 1) (Dodson et al., 2015) and Lhx6 (LIM homeobox 6) (Abdi et al. 2015). On the other hand, arky pallidal neurons present a unique molecular signature, expressing the opioid precursor preproenkephalin (PPE) and the forkhead box protein P2 (FoxP2) (Mallet et al., 2012; Abdi et al. 2015; Dodson et al., 2015). In some studies, Npas1 (neuronal PAS domain protein 1), another protein-coding gene, is used to label arky pallidal neurons, since it largely overlaps with FoxP2-expressing neurons (Pamukcu et al. 2020).

One key property that has been used to distinguish between prototypical and arky pallidal neurons is their firing rates. This distinction was first observed in DeLong's original analysis of cells in non-human primates (DeLong, 1971). He found that in dopamine-intact *in vivo* conditions, prototypical neurons had reliable spontaneous firing rates ranging from 1 to 100 spikes/s, with an average of 50 – 55 spike/s overall (see also (Dodson et al., 2015)). On the other hand, arky pallidal neurons had more irregular and sporadic activity in both *in vivo* and *ex vivo* conditions, with overall lower firing rates ranging from 1 to 30 spike/s (10 spikes/s average). This firing dropped during sleep, unlike prototypical neurons (see also (Mallet et al., 2012; Gittis et al., 2014; Abdi et al. 2015; Dodson et al., 2015; Mallet et al., 2016; Pamukcu et al. 2020; Aristieta et al. 2021; Ketzef and Silberberg, 2021)). These observations highlight how these two cell populations have clearly distinct spike rate characteristics that distinguish them, along with their different underlying molecular signatures.

There is also some evidence to suggest that the cells in these two GPe neuron classes have somewhat different morphologies (Mallet et al., 2012). Arky pallidal neurons' axons appear to be characterized by lengths far exceeding those of prototypical neurons and dendritic spines with significantly higher density. On the other hand, prototypical cells have been shown to feature significantly longer local axon collaterals and a larger number of synaptic boutons compared to arky pallidals. More work is still needed to confirm these morphological distinctions between the two

major cell types, however. Despite these morphological distinctions, prototypical and arky pallidal neurons appear to be intermingled throughout the GPe (DeLong, 1971; Dodson et al., 2015), and this homogeneous distribution appears to be relatively consistent across the rostral, central, and caudal segments of the GPe (Abdi et al. 2015).

Collectively, the prototypical and arky pallidal cells account for about 95% of all GABAergic GPe neurons (Abdi et al. 2015). Given their molecular and electrophysiological distinctions, it seems reasonable to suspect that they contribute differently to GPe's role in regulating the information flow through the CBGT circuits, which raises the possibility of a more sophisticated role for the GPe than the classical theories suggest.

3.2 | Connectivity

Another way to categorize GPe neurons is by the nature of their connections. In this view, two different groups of neurons emerge: one relays information downward within the basal ganglia, while the other participates in the flow of information upwards through the basal ganglia. It is generally accepted that the former group aligns with the prototypical neurons, while the latter are mostly arky pallidal neurons. The efferent and afferent pathways from these two cell types are shown in Figure 2B and 2C, respectively. Here we provide a brief overview of the established afferent, efferent, and collateral projections of the GPe.

We begin with the outward, efferent projections from the GPe. The only cell class known to make synapses with the STN and the GPi (or substantia nigra pars reticulata, SNr) are the prototypical neurons (Abdi et al. 2015; Mallet et al., 2012; Mastro et al., 2014; Saunders et al., 2016; Glajch et al. 2016; Aristieta et al. 2021) (Figure 2B). Arky pallidal neurons instead provide GABAergic innervation - around a thousand axonal boutons per arky pallidal neuron (Mallet et al., 2012; Fujiyama et al., 2016; Dong et al., 2021) - across the striatum, projecting to direct and indirect pathway SPNs as well as to striatal fast spiking interneurons (FSIs) (Mallet et al., 2012; Gittis et al., 2014; Abdi et al. 2015; Hernández et al., 2015; Saunders et al., 2015; Fujiyama et al., 2016; Corbit et al., 2016; Pamukcu et al. 2020; Aristieta et al. 2021; Ketzef and Silberberg, 2021). Still little is known about the relative strength of arky pallidal projections to striatum. Glajch et al (Glajch et al. 2016), however, assessed that arky pallidal neurons project more strongly to iSPN neurons than to dSPNs, with a 2:1 ratio. Prototypical neurons also have been determined to project to striatum, although experimental findings do not agree on the target populations, how dense these projections are, or to what extent they are functionally relevant. According to some studies, prototypical neurons have been shown to have dense projections into the striatum (Fujiyama et al., 2016), while other studies suggest that the density of striatal connections from prototypical cells may be more modest (Mallet et al., 2012). These projections appear to mainly target the FSI cells in the striatum (Gittis et al., 2014; Abdi et al. 2015; Saunders et al., 2016; Glajch et al. 2016), and with more strength than that with which arky pallidal neurons signal to FSIs (Corbit et al., 2016). Interestingly, findings have revealed the existence of GPe projections to the neocortex itself (Saunders et al., 2015; Chen et al., 2015), particularly to cortical motor areas from a specific subset of GPe neurons co-expressing Npas1 and Nkx2.1 and from GPe cholinergic neurons (Abecassis et al. 2020), suggesting that the GPe could be involved in the regulation of premotor and motor activity through a cortico-pallidal-cortical loop (Chen et al., 2015; Courtney et al., 2023). It remains to be determined whether there are other pallidal cell subtypes that also project to the neocortex.

As for the afferent projections, most of the existing evidence points to distinct input connections to prototypical and arky pallidal neurons (Figure 2C). Indeed, both the STN and striatum have been shown to differentially innervate GPe cells. STN and iSPNs provide more robust inputs to prototypicals than to arky pallidal neurons (Pamukcu et al. 2020; Aristieta et al. 2021; Gast et al., 2021; Ketzef and Silberberg, 2021). In particular, iSPN projections to arky pallidal neurons have been estimated to be 85% weaker than those targeting prototypical neurons and also less numerous (Aristieta et al. 2021), while STN inputs have been measured to be 74% weaker to arky pallidal than prototypical cells

(Aristieta et al. 2021). In addition, emerging studies have reported the existence of other GABAergic projections coming from the striatum, specifically from dSPN neurons and preferentially targeting arky pallidal neurons (Ketzel and Silberberg, 2021; Cui et al. 2021) or, more broadly, Npas1 neurons (Labouesse et al. 2023). For consistency's sake, we will refer to these as arky pallidal neurons, even though not all Npas1 neurons are of this type. Given the nature of the suppressive, outward efferent projections from GPe_A neurons (Figure 2B), these inhibitory inputs from dSPNs may promote action initiation via inhibition of arky pallidal cells, constituting a so-called non-canonical striatopallidal "Go" pathway (Ketzel and Silberberg, 2021; Aristieta et al. 2021; Labouesse et al. 2023). Little, however, is known about the specifics of this connection. It could be possible, indeed, that the inhibition promoted by dSPNs onto arky pallidal neurons is topographically organized into functional units that encode specific motor patterns and outcome behaviors (Labouesse et al. 2023). Moreover, Aristieta et al. (2021) showed that *in vivo* opto-stimulation of iSPN neurons disinhibits arky pallidal neurons through a disynaptic circuit, suggesting that dSPN-pallidal projections could balance arky pallidal outputs through an indirect local competition with iSPN-GPe inputs (Labouesse et al. 2023). These ideas about dSPN impact on GPe_A, however, contrast with other findings that dSPNs exclusively affect prototypical neurons (Mizutani et al., 2017). More work is required to resolve this discrepancy. Lastly, cortical projections have been identified as the source of almost 10% of the total input into GPe, preferentially targeting arky pallidal neurons (Abecassis et al. 2020; Karube et al., 2019). Indeed, experiments suggest that only one third to half of prototypical neurons receive cortical inputs (Abecassis et al. 2020).

Finally, we turn to the internal connectivity of the GPe. Within the GPe, strong collateral GABAergic projections from prototypical to arky pallidal neurons have been observed and are thought to play a fundamental role in switching the activity of arky pallidal neurons on and off, according to whether prototypical neurons are inhibited or not, respectively (Mallet et al., 2012; Dodson et al., 2015; Fujiyama et al., 2016; Aristieta et al. 2021; Ketzel and Silberberg, 2021). What is less certain is whether these collateral connections are reciprocal. Optogenetic excitation of arky pallidal neurons has not been shown to produce inhibitory responses in prototypical neurons, in either *ex vivo* or *in vivo* (Aristieta et al. 2021) conditions, suggesting either that synapses from arky pallidal onto prototypical neurons do not exist or that their influence is fairly weak (Mallet et al., 2012; Ketzel and Silberberg, 2021; Gast et al., 2021). Lastly, some studies suggest that prototypical neurons exhibit a stronger degree of intra-population inhibition than do arky pallidal neurons (Ketzel and Silberberg, 2021; Gast et al., 2021). In contrast, others have assessed that the strengths of connections between arky pallidal neurons seem modest, but connections between prototypical cells are even weaker (Nevado-Holgado et al., 2014).

3.3 | Functional roles of GPe neuron subtypes in behavior

As was pointed out in Section 2.2, the GPe was traditionally considered to be a node of the "motor-suppressing" indirect pathway, conveying descending signals to the output nuclei of the basal ganglia circuit (Calabresi et al., 2014). Indeed, according to the classical model, striatal iSPN neurons send the majority of their projections to the GPe, which then exerts an inhibitory influence on the STN and the GPi (Kita, 2007). Taking into account the excitatory connections from STN to GPe immediately complicates this picture. As a specific example, if GPe activity increases and results in strengthened inhibition to the STN, then the GPe neurons lose some of their excitatory input and hence may reduce their activity back towards baseline levels. This feedback loop has long been recognized as a possible source of parkinsonian oscillations when dopaminergic effects become compromised (Plenz and Kital, 1999; Terman et al., 2002). More recently, the emerging work of the past decade has cast doubt on the simple, one-dimensional view of the functional role of the GPe (Mallet et al., 2012; Abdi et al. 2015; Dodson et al., 2015; Abecassis et al. 2020). With the discovery of new cell types within the GPe and new connections (e.g., arky pallidal projections to striatum;

Section 3.2), a renewed interest has emerged in understanding the functional properties of this nucleus in light of its complex connectivity and cellular composition.

It is now known that the GPe contributes to both motor and non-motor functions and communicates with other basal ganglia nuclei and other brain regions through both upstream and downstream projections (Figure 2B-C). Amongst the non-motor roles of GPe cells, a small pool of PV-expressing GPe neurons seems to be associated with reversal learning and processing of sensory and reward cues (Courtney et al., 2023; Lilascharoen et al. 2021; Farries et al., 2023). Consistent with this, a recent study by Isett et al. (2023) showed that inhibition of PV cells in the GPe drove transient punishment of behavior, not motor suppression (Isett et al., 2023), highlighting the role of GPe in learning processes, as well as movement control. Evidence suggests that other GPe cells, likely Npas1-Nkx2.1-expressing, could be involved in regulating sleep (Qiu et al., 2016; Vetrivelan et al., 2010; Lazarus et al., 2013) and limbic functions (Stephenson-Jones et al., 2016; Wallace et al., 2017). Dysfunctions of GPe contribute to several clinical conditions such as Parkinson's disease (Mallet et al., 2008; Gittis et al., 2014; Crompe et al., 2020; Courtney et al., 2023), Huntington's disease (Courtney et al., 2023; Starr et al., 2008; Beste et al., 2015; Deng et al., 2021) and dystonia (Baron et al., 2011; Nambu et al., 2011; Starr et al., 2005; Chiken et al., 2008). Analysis of the various functional roles of the GPe would be worthy of an entire review in and of itself (see also (Courtney et al., 2023; Dong et al., 2021; Hegeman et al., 2016)). For simplicity's sake, we will focus here on those roles related to action control.

There is now a consistent body of evidence linking variations in firing rate of GPe neurons with the dynamics of movement (Yoshida and Tanaka, 2016; Gu et al., 2020; Arkadir et al., 2004), as well as a variety of more direct causal tests of their role in motor control (Glajch et al. 2016; Pamukcu et al. 2020; Aristieta et al. 2021; Cui et al. 2021; Mastro et al., 2017; Lilascharoen et al. 2021). Indeed, GPe neuron firing patterns are not only correlated with movement specifics, such as amplitude, velocity, and direction (Georgopoulos et al., 1983; Mitchell et al., 1987; Gage et al., 2010), but have also been found to tune for the body region involved, and the nature of the movement itself (e.g., whether is passive, active, or externally cued) (Georgopoulos et al., 1983; Turner and Anderson, 1997; 2005; Gage et al., 2010). In this vein, it is important to recognize that even though GPe output is inhibitory, it is nonetheless possible that it passes along informative signals to its synaptic targets via deviations from its baseline, pacemaker-like firing (see e.g. (Corbit et al., 2016) for analogous effects related to inhibitory outputs of FSIs).

Within the context of the various cell types in the GPe, it has been proposed that there is a cell-type-specific encoding of spontaneous movement in the GPe (Dodson et al., 2015). According to this model, the prototypical neurons show heterogeneous firing responses, while arkypallidal neurons present robust increases in their firing profile during spontaneous movements. Here, the decrease in firing of GPe cells, particularly prototypical neurons, from activation of striatal neurons during movement (Cui et al., 2013) would reflect the traditional role of the GPe as an arm of the indirect pathway (Dodson et al., 2015; Kravitz et al., 2010; Sano et al., 2013). In contrast, since arkypallidal neurons show little firing at rest (Mallet et al., 2012; Dodson et al., 2015; DeLong, 1971) and robustly increase activity around movement onset (Dodson et al., 2015), they could be engaged in action facilitation. Inhibiting large striatal regions could prevent competing actions from being expressed in order to promote the selection of a desired action (Mallet et al., 2012; Aristieta et al. 2021; Ketzef and Silberberg, 2021; Glajch et al. 2016; Hegeman et al., 2016). For the remainder of this section, we will focus on this distinction in the roles of prototypical and arkypallidal neurons in the action selection (or inhibition) process.

Much of the recent work examining the role of the GPe cell types in motor control and action selection has focused primarily on the process of inhibitory control, with most of the emphasis placed on the contribution of arkypallidal neurons (Mallet et al., 2016; Schmidt and Berke, 2017; Dodson et al., 2015). Mallet et al. (2016) (Mallet et al., 2016) examined the activity of the two GPe subpopulations in a stop signal task scenario (Section 2.1) and found that the time courses of both prototypical and arkypallidal firing exhibit a clear increase following the presentation of a

stop signal. Arkypallidal neurons produce a significantly stronger response to the stop cue than prototypical neurons do, suggesting that arkypallidals have a greater influence on the production of a stop response. This enhancement of arkypallidal activity occurs just before the surge of movement-related striatal activity, as would be expected if the arkypallidal neurons play an important role in canceling imminent actions. A more recent study by Aristieta et al. (2021) (Aristieta et al. 2021) supported this hypothesis by using *in vivo* optogenetic stimulation of arkypallidal FoxP2-expressing neurons during the same stop signal task (Figure 3A). This manipulation produced a strong inhibition of ongoing locomotion, providing causal evidence that activation of arkypallidal neurons is sufficient to induce this effect, likely through a global suppression of go-related striatal activity. Similarly, Pamucku et al. (2020) (Pamucku et al. 2020) showed that optogenetic stimulation of Npas1-expressing neurons, a subset of cells within the GPe mostly consisting of arkypallidal neurons but also including a small fraction of prototypical neurons (Abdi et al. 2015), induces a decrease in the vigor of motor output, both in terms of duration and speed of movement (Figure 3B). To confirm that the movement-suppression effect enhanced by Npas1-expressing neurons upon optogenetic stimulation is mediated through the inhibition of the dorsal striatum, they also optogenetically stimulated axon terminals of Npas1-expressing neurons in the dorsal striatum directly. This stimulation produced a decrease in locomotion, which provides further support for the hypothesis that the arkypallidal pathways play a pivotal role in movement inhibition.

On the other hand, there is no consensus on the involvement of prototypical cells in reactive inhibition. Some lines of evidence have shown that prototypical neurons also become mildly excited during the presentation of a stop signal (Mallet et al., 2016). This evidence, however, must be contrasted with evidence suggesting that prototypical neurons do not play any role in the context of reactive inhibition (Aristieta et al. 2021). One way to reconcile these disparate observations comes from Aristieta et al. (2021) (Aristieta et al. 2021), who showed that inhibitory inputs from axon collaterals from prototypical neurons control the activity of arkypallidal neurons. Specifically, the authors found that optogenetic excitation of prototypical neurons produced a strong inhibition of the activity of recorded arkypallidal cells. The influence of prototypical neurons on movement suppression then makes sense given the circuit-level logic of the striato-pallidal-striatal loop. Specifically, activation of iSPNs inhibits prototypical neurons that, in turn, inhibit arkypallidal units that inhibit dSPNs. In this way, there is a secondary arm of the canonical indirect pathway that further suppresses the motor-promoting signals from dSPNs through disinhibition of arkypallidal cells. These findings provide evidence for the existence of a wiring architecture between prototypical and arkypallidal cells that is capable of regulating the activity of arkypallidal neurons, suggesting that the prototypical neurons could play an indirect role in the control of reactive inhibition via modulation of arkypallidal activity. Moreover, Gage et al. (2010) suggest that a sharp decrease of GPe prototypical neurons plays a role in information processing during behavioral choice tasks (Gage et al., 2010). Indeed, the disinhibition provoked by prototypical neurons onto FSIs, causes a coordinated pulse of increasing activity in FSIs as chosen actions are initiated while suppressing unwanted alternatives.

4 | RETHINKING THE CLASSICAL MODEL: GPE AS A CENTRAL HUB

These new insights into the organization, connectivity, and functional roles of the GPe fundamentally shift our understanding of this nucleus and its role in regulating the flow of information through CBGT circuits. While the classical model posits the GPe as a nucleus with a homogenous neuronal composition, we now recognize a rich complexity of cell types in the GPe with a qualitative distinction into two major classes: prototypical and arkypallidal neurons. Also in the classical model, the GPe only projects to the STN and GPi/SNr, thus relaying only descending signals to the output of the basal ganglia. We now know, however, that the GPe also sends ascending signals to the striatum, directly onto striatal SPNs and FSIs. Thus, the influence of the GPe goes both up and down relative to the traditional

basal ganglia pathways. Finally, the classical model interpreted the GPe as a simple way station along the movement-inhibiting pathway originating from the iSPNs (i.e., indirect pathway). New experimental evidence paints a much more complex picture of the GPe's functional role in the motor domain, with seemingly mixed results about its influence on subsequent behavior. These fundamental changes in our understanding of the GPe suggest that we need to re-think the role that this nucleus plays in CBGT computations. The terms descending and ascending information still make sense, because they are defined in terms of the dominant basal ganglia output nuclei, the GPi and SNr, being downstream. In this new view, however, the GPe is centrally located to regulate bidirectional information flow. The path of descending information (purple connections in Figure 4A) is an extension of the classical model, with striatal SPNs sending signals to the GPe that are then relayed down to the STN and GPi via prototypical cells. In addition, ascending information (orange connections, Figure 4A), originating either from hyperdirect pathway drive to STN or possibly from direct cortical projections to the GPe (Karube et al., 2019), propagates up to the striatum via arkypallidal GPe neurons and regulates SPN firing through GABAergic signaling. Moreover, the two GPe populations interact, at least through prototypical inhibition of arkypallidal GPe neurons, setting up a possible mechanism for dominance to switch between the two directions. In this way, information flow through the CBGT circuit is no longer unidirectional and the control of ascending and descending information is centrally regulated by the GPe.

We can appreciate this central role of the GPe in regulating bidirectional information flow most clearly in the process of reactive inhibition. In particular, the recently proposed *Pause-then-Cancel* model (Mallet et al., 2016; Schmidt and Berke, 2017) highlights the complementary roles of ascending and descending signals in reactive stopping (Figure 4B). Framed in the context of the typical stop signal task (Section 2.1), the *Pause-then-Cancel* model still separates two competing control signals from cortex: the imperative "Go" signal from cortex to the striatum, particularly the direct pathway, and the reactive "Stop" signal from the hyperdirect pathway. When a primary stimulus is presented (the "Go" cue), cortical inputs drive the activity of dSPNs (direct pathway; green path in Figure 4B). These then inhibit the GPi, reducing its inhibition on the thalamus and increasing the likelihood of an action. The *Pause-then-Cancel* model of reactive stopping begins with the same preliminary step as the classical model of reactive inhibition (Section 2.2), in which the hyperdirect pathway quickly activates the STN, sending a surge of excitation to the GPi and thus increasing its inhibition of the thalamus (yellow path in Figure 4B). In this model, however, this drive signal is not sufficient to fully cancel the planned action (Mallet et al., 2016; Schmidt and Berke, 2017), but appears only to delay the progression of the ramping up of thalamic excitation that would promote motor behavior. It is at this stage of the process that the *Pause-then-Cancel* model deviates from the classical model.

Specifically, the "cancel" stage of this model (red path in Figure 4B) relies heavily on the bidirectional influence of the GPe in order to fully implement the cancellation of the planned response. Starting with the ascending flow of information, arkypallidal neurons become engaged either via drive signals from the STN or direct cortical input signals (Karube et al., 2019). Since the sole known efferents of the arkypallidal neurons are inhibitory connections to the striatum, we can imagine that this activity reduces responses in the direct and indirect pathways. This reduction helps to maintain the suppression of thalamic ramping until cortical drive to iSPNs can take over. This begins a stage where descending information flow becomes the dominant factor in the stopping process, which happens in two parts. First, because of the inhibition of dSPNs, GPi is released from inhibition. Second, because iSPNs are being engaged, the balance of power shifts to the traditional indirect pathway. The iSPNs predominantly inhibit prototypical cells in the GPe, which are the only pallidal cells to project downward to the STN (long indirect pathway) and GPi (short indirect pathway). Thus, GPi output can increase, providing an enhanced suppression of the thalamic response and leading to the cessation of the planned response.

The *Pause-then-Cancel* model articulated within the context of our expanded knowledge of the GPe clearly highlights two critical aspects of CBGT computation. First, the GPe links proactive (direct/indirect pathways) and reactive

(hyperdirect pathway) inhibitory mechanisms together. Second, the GPe is ideally situated to regulate the bidirectional flow of information through the CBGT circuit. Indeed, the second feature is a mechanism for the first. Specifically, the GPe plays a key role in relaying ascending reactive control signals to the striatal pathways that implement proactive control through descending projections. Thus, the GPe acts as a central hub within the CBGT circuit, rather than being an isolated component of the indirect pathway alone.

5 | LOOKING FORWARD

As we have shown, the emerging evidence on the cellular composition, connectivity, and functional characteristics of the GPe fundamentally changes our understanding of the role that this nucleus plays in regulating information flow through the CBGT circuit. While the classical model considers the GPe as a homogeneous nucleus, we now know that it is composed of a variety of cell types that can be divided into at least two general categories: prototypical and arky pallidal neurons. The rediscovery of a heterogeneous collection of GPe cell types has led to renewed interest in the anatomical and functional characteristics of each subpopulation. Prototypical neurons continue to be seen as an integral part of the classical indirect pathway through which striatal iSPN signals impact the activity downstream in the STN and GPi. Interestingly, a new twist on this idea is that prototypical GPe neuron firing rates may tune the chloride load and hence GABA reversal potential for GPi (or SNr) neurons, thereby impacting how strongly dSPN inputs affect GPi firing rates and yielding a new form of interaction between the direct and indirect pathways (Phillips et al., 2020). On the other hand, arky pallidal neurons regulate ascending information through striatum-targeting projections. In light of this new evidence, we now see that the GPe assumes a central role in basal ganglia computations, becoming not just a relay station of the indirect pathway, but a pivotal hub of the full CBGT circuit, regulating both ascending and descending information streams. Using the Pause-then-Cancel model as an example, we have highlighted how the GPe is ideally situated to integrate signals from different cortical sources in order to implement behavioral control. In particular, this integration arises in stopping, because this process combines both reactive and proactive control signals to cancel planned actions. This example supports the idea that as a field, we should shift our conceptualization of the GPe from being an incidental node along the indirect pathway to a central hub that integrates signals from all three canonical CBGT pathways.

It is worth noting that this view of the central role of the GPe in basal ganglia function has been recently proposed by Courtney et al. (2023) (Courtney et al., 2023). In this review, the authors go over the same cellular and circuit-level discoveries as we review here, arriving at a similar conclusion that the GPe acts to regulate more distributed aspects of basal ganglia network function than previously thought. Unlike the current perspective piece, however, their work primarily focuses on how the GPe contributes to the etiology of disease states. For example, the loss of dopamine that characterizes Parkinson's disease correlates with alterations in GPe neuronal activity. Indeed, *Npas1* neurons show hypoactivity, while the STN input strength to GPe PV neurons is reduced (Pamukcu et al. 2020). Moreover, a late stage in the progression of Huntington's disease causes the loss of arky pallidal neurons (Deng et al., 2021), while dystonia symptomatology seems to rely on reduced activity of GPe PV neurons due to compromised hyperpolarization and cyclic nucleotide-gated (HCN) channels (Chiken et al., 2008). Here we complement this perspective on pathologies of the CBGT circuit by focusing more explicitly on the computational role that the GPe plays in normative function. Putting the two together, the disease states described by Courtney et al. (2023) can be understood in terms of alterations in the bidirectional information processing that occurs in normal basal ganglia dynamics. Nonetheless, this similarity in conclusions highlights the converging new view of the field on the role of the GPe in CBGT circuit dynamics.

Theoretically, this shift in perspective forces a fundamental change in how we think about information flow in CBGT circuits. Traditional models of these pathways describe a one-way architecture, where information conveyed by cortex propagates "down" towards the output nuclei, the GPI and SNr (Mink, 1996; Alexander et al., 1986). This has been the dominant view of CBGT information flow for over a half-century, reflecting the "independent" aspect of the "parallel and independent" pathways framework (Alexander et al., 1986). However, if the GPe regulates ascending information flow, as well as descending information, allowing signals originating at the STN (or GPe itself) to influence striatal computation, then our collective understanding of the circuit needs to be updated. The CBGT pathways in this new view comprise more a complex, recurrent network architecture that allows for the integration of signals from multiple cortical (and possibly subcortical) sources. No pathway in the CBGT circuit can be considered as fully "independent" anymore, and our models of CBGT computation need to be revised to reflect the interactions involved.

With this new understanding, many new questions arise. As we point out in Section 3.1, a consensus about the cellular composition of the GPe has yet to be reached. The pool of existing evidence characterizing the GPe cell populations remains small and incomplete. The heterogeneity of the GPe, in fact, goes beyond the simple dichotomous organization of GABAergic neurons into prototypical and arky pallidal cells (see Figure 2A). Indeed, this represents only a conveniently simplified reduction of the true underlying cellular complexity of the GPe. If different neuron types have different anatomical and functional properties, then the nature of GPe computations is likely still more complex than what we have discussed. Even within the dichotomous classification, unknowns remain. For example, are arky pallidal neurons homogenous in terms of their involvement in inhibitory control or is there functional variability across arky pallidal subpopulations? In addition, arky pallidal cells receive afferents from multiple sources, including the striatum, STN, and recently discovered inputs from cortex (Abecassis et al. 2020; Karube et al., 2019). How do arky pallidals integrate these disparate inputs and are there yet unknown afferents that remain to be discovered? Moreover, recent data suggest the possibility of heterogeneous neural subtypes within target nuclei of the GPe, setting up the possibility of a more complex collection of parallel pathways than those included in the classical basal ganglia model (Delgado-Zabalza et al., 2023). We are at the tip of the proverbial iceberg in terms of our understanding of this nucleus, and CBGT circuits more broadly.

Nevertheless, this new perspective on the CBGT circuit already raises a critical question: what advantages do a bidirectional flow of information, and the increased resource demands that come along with it, provide in terms of CBGT computation? Building off of our focus on inhibitory control, we can ask how ascending projections from arky pallidal neurons influence striatal computation. Do arky pallidal neurons serve to amplify inhibition promoted by the indirect pathway alone or does their role rely more on shifting the balance of power between iSPNs and dSPNs (Dunovan and Verstynen, 2016)? The answer to this question has critical implications for the nature of behavioral control mediated by these circuits.

Acknowledgements

The authors would like to thank Jyotika Bahuguna and Nico Mallet for helpful discussions on the GPe that informed the structure of this review.

Conflict of interest

The authors have no conflict of interest to report.

Funding Information

CG and CV are supported by the PCI2020-112026 project, and CV is also supported by the PCI2023-145982-2, both funded by MCIN/AEI/10.13039/501100011033 and by the European Union "NextGenerationEU"/PRTR as part of the CRCNS program. TV and JER are partly supported by NIH awards R01DA053014 and R01DA059993 as part of the CRCNS program. AG and JER are partly supported by NIH award R01NS125814, also part of the CRCNS program.

Author contributions

CG and TV conceived the idea of this manuscript, reviewed the literature, and wrote the manuscript. JER, CV, and AG provided many useful suggestions and helped to edit this manuscript. All authors read and approved the final manuscript.

References

- Abdi, A., N. Mallet, F. Y Mohamed, A. Sharott, P. D Dodson, K. C Nakamura, S. Suri, S. V Avery, J. T Larvin, F. N Garas, et al. 2015. *Prototypic and arkypallidal neurons in the dopamine-intact external globus pallidus*, *Journal of Neuroscience* **35**, no. 17, 6667–6688.
- Abecassis, Z. A, B. L Berceau, P. H Win, D. Garcia, H. S Xenias, Q. Cui, A. Pamukcu, S. Cherian, V. M Hernández, U. Chon, et al. 2020. *Npas1+*-*nkx2. 1+ neurons are an integral part of the cortico-pallido-cortical loop*, *Journal of Neuroscience* **40**, no. 4, 743–768.
- Alderson, R M., M. D Rapport, and M. J Kofler. 2007. *Attention-deficit/hyperactivity disorder and behavioral inhibition: a meta-analytic review of the stop-signal paradigm*, *Journal of abnormal child psychology* **35**, 745–758.
- Alexander, G. E, M. R DeLong, and P. L Strick. 1986. *Parallel organization of functionally segregated circuits linking basal ganglia and cortex*, *Annual review of neuroscience* **9**, no. 1, 357–381.
- Aristieta, A., M. Barresi, S. A. Lindi, G. Barriere, G. Courtand, B. de La Crompe, L. Guilhemsang, S. Gauthier, S. Fioramonti, J. Baufreton, et al. 2021. *A disinaptic circuit in the globus pallidus controls locomotion inhibition*, *Current Biology* **31**, no. 4, 707–721.
- Arkadir, D., G. Morris, E. Vaadia, and H. Bergman. 2004. *Independent coding of movement direction and reward prediction by single pallidal neurons*, *Journal of Neuroscience* **24**, no. 45, 10047–10056.
- Aron, A. R. 2011. *From reactive to proactive and selective control: developing a richer model for stopping inappropriate responses*, *Biological psychiatry* **69**, no. 12, e55–e68.
- Aron, A. R, D. M Herz, P. Brown, B. U Forstmann, and K. Zaghoul. 2016. *Frontosubthalamic circuits for control of action and cognition*, *Journal of Neuroscience* **36**, no. 45, 11489–11495.
- Aron, A. R and R. A Poldrack. 2006. *Cortical and subcortical contributions to stop signal response inhibition: role of the subthalamic nucleus*, *Journal of Neuroscience* **26**, no. 9, 2424–2433.
- Baron, M. S, K. D Chaniary, A. C Rice, and S. M Shapiro. 2011. *Multi-neuronal recordings in the basal ganglia in normal and dystonic rats*, *Frontiers in systems neuroscience* **5**, 67.
- Beste, C., M. Mückschel, S. Elben, C. J Hartmann, C. C McIntyre, C. Saft, J. Vesper, A. Schnitzler, and L. Wojtecki. 2015. *Behavioral and neurophysiological evidence for the enhancement of cognitive control under dorsal pallidal deep brain stimulation in huntington's disease*, *Brain Structure and Function* **220**, 2441–2448.

- Braver, T. S. 2012. *The variable nature of cognitive control: a dual mechanisms framework*, Trends in cognitive sciences **16**, no. 2, 106–113.
- Calabresi, P., B. Picconi, A. Tozzi, V. Ghiglieri, and M. Di Filippo. 2014. *Direct and indirect pathways of basal ganglia: a critical reappraisal*, Nature neuroscience **17**, no. 8, 1022–1030.
- Chamberlain, S. R., N. A. Fineberg, A. D. Blackwell, T. W. Robbins, and B. J. Sahakian. 2006. *Motor inhibition and cognitive flexibility in obsessive-compulsive disorder and trichotillomania*, American journal of psychiatry **163**, no. 7, 1282–1284.
- Chen, M. C., L. Ferrari, M. D. Sacchet, L. C. Foland-Ross, M.-H. Qiu, I. H. Gotlib, P. M. Fuller, E. Arrigoni, and J. Lu. 2015. *Identification of a direct gabaergic pallidocortical pathway in rodents*, European Journal of Neuroscience **41**, no. 6, 748–759.
- Chen, W., C. de Hemptinne, A. M. Miller, M. Leibbrand, S. J. Little, D. A. Lim, P. S. Larson, and P. A. Starr. 2020. *Prefrontal-subthalamic hyperdirect pathway modulates movement inhibition in humans*, Neuron **106**, no. 4, 579–588.
- Chiken, S., P. Shashidharan, and A. Nambu. 2008. *Cortically evoked long-lasting inhibition of pallidal neurons in a transgenic mouse model of dystonia*, Journal of Neuroscience **28**, no. 51, 13967–13977.
- Corbit, V. L., T. C. Whalen, K. T. Zitelli, S. Y. Crilly, J. E. Rubin, and A. H. Gittis. 2016. *Pallidostriatal projections promote β oscillations in a dopamine-depleted biophysical network model*, Journal of Neuroscience **36**, no. 20, 5556–5571.
- Courtney, C. D., A. Pamukcu, and C. S. Chan. 2023. *Cell and circuit complexity of the external globus pallidus*, Nature Neuroscience, 1–13.
- Crompe, B. d. l., A. Aristieta, A. Leblois, S. Elsherbiny, T. Boraud, and N. P. Mallet. 2020. *The globus pallidus orchestrates abnormal network dynamics in a model of parkinsonism*, Nature communications **11**, no. 1, 1570.
- Cui, G., S. B. Jun, X. Jin, M. D. Pham, S. S. Vogel, D. M. Lovinger, and R. M. Costa. 2013. *Concurrent activation of striatal direct and indirect pathways during action initiation*, Nature **494**, no. 7436, 238–242.
- Cui, Q., X. Du, I. Y. Chang, A. Pamukcu, V. Lilascharoen, B. L. Berceau, D. García, D. Hong, U. Chon, A. Narayanan, et al. 2021. *Striatal direct pathway targets npas1+ pallidal neurons*, Journal of Neuroscience **41**, no. 18, 3966–3987.
- Cui, Q., A. Pamukcu, S. Cherian, I. Y. Chang, B. L. Berceau, H. S. Xenias, M. H. Higgs, S. Rajamanickam, Y. Chen, X. Du, et al. 2021. *Dissociable roles of pallidal neuron subtypes in regulating motor patterns*, Journal of Neuroscience **41**, no. 18, 4036–4059.
- Delgado-Zabalza, L., N. P. Mallet, C. Glangetas, G. Dabee, M. Garret, C. Miguelez, and J. Baufreton. 2023. *Targeting parvalbumin-expressing neurons in the substantia nigra pars reticulata restores motor function in parkinsonian mice*, Cell Reports **42**, no. 10.
- DeLong, M. R. 1971. *Activity of pallidal neurons during movement.*, Journal of neurophysiology **34**, no. 3, 414–427.
- Deng, Y., H. Wang, M. Joni, R. Sekhri, and A. Reiner. 2021. *Progression of basal ganglia pathology in heterozygous q175 knock-in huntington's disease mice*, Journal of Comparative Neurology **529**, no. 7, 1327–1371.
- Dodson, P. D., J. T. Larvin, J. M. Duffell, F. N. Garas, N. M. Doig, N. Kessar, I. C. Duguid, R. Bogacz, S. J. Butt, and P. J. Magill. 2015. *Distinct developmental origins manifest in the specialized encoding of movement by adult neurons of the external globus pallidus*, Neuron **86**, no. 2, 501–513.
- Dong, J., S. Hawes, J. Wu, W. Le, and H. Cai. 2021. *Connectivity and functionality of the globus pallidus externa under normal conditions and parkinson's disease*, Frontiers in neural circuits **15**, 645287.
- Dunovan, K., B. Lynch, T. Molesworth, and T. Verstynen. 2015. *Competing basal ganglia pathways determine the difference between stopping and deciding not to go*, Elife **4**, e08723.
- Dunovan, K. and T. Verstynen. 2016. *Believer-skeptic meets actor-critic: rethinking the role of basal ganglia pathways during decision-making and reinforcement learning*, Frontiers in neuroscience **10**, 106.

- Farries, M. A, T. W Faust, A. Mohebi, and J. D Berke. 2023. *Selective encoding of reward predictions and prediction errors by globus pallidus subpopulations*, *Current Biology* **33**, no. 19, 4124–4135.
- Fillmore, M. T and C. R Rush. 2002. *Impaired inhibitory control of behavior in chronic cocaine users*, *Drug and alcohol dependence* **66**, no. 3, 265–273.
- Fujiyama, F., T. Nakano, W. Matsuda, T. Furuta, J. Udagawa, and T. Kaneko. 2016. *A single-neuron tracing study of arkypallidal and prototypic neurons in healthy rats*, *Brain Structure and Function* **221**, 4733–4740.
- Gage, G. J, C. R Stoetznner, A. B Wiltschko, and J. D Berke. 2010. *Selective activation of striatal fast-spiking interneurons during choice execution*, *Neuron* **67**, no. 3, 466–479.
- Gast, R., R. Gong, H. Schmidt, H. G. Meijer, and T. R Knösche. 2021. *On the role of arkypallidal and prototypical neurons for phase transitions in the external pallidum*, *Journal of neuroscience* **41**, no. 31, 6673–6683.
- Georgopoulos, A. P, M. R DeLong, and M. D Crutcher. 1983. *Relations between parameters of step-tracking movements and single cell discharge in the globus pallidus and subthalamic nucleus of the behaving monkey*, *Journal of Neuroscience* **3**, no. 8, 1586–1598.
- Gittis, A. H, J. D Berke, M. D Bevan, C S. Chan, N. Mallet, M. M Morrow, and R. Schmidt. 2014. *New roles for the external globus pallidus in basal ganglia circuits and behavior*, *Journal of Neuroscience* **34**, no. 46, 15178–15183.
- Glajch, K. E, D. A Kelter, D. J Hegeman, Q. Cui, H. S Xenias, E. C Augustine, V. M Hernández, N. Verma, T. Y Huang, M. Luo, et al. 2016. *Npas1+ pallidal neurons target striatal projection neurons*, *Journal of Neuroscience* **36**, no. 20, 5472–5488.
- Gu, B.-M., R. Schmidt, and J. D Berke. 2020. *Globus pallidus dynamics reveal covert strategies for behavioral inhibition*, *Elife* **9**, e57215.
- Haber, S. N. 2003. *The primate basal ganglia: parallel and integrative networks*, *Journal of Chemical Neuroanatomy* **26**, no. 4, 317–330.
- Hegeman, D. J, E. S Hong, V. M Hernández, and C S. Chan. 2016. *The external globus pallidus: progress and perspectives*, *European Journal of Neuroscience* **43**, no. 10, 1239–1265.
- Hernández, V. M, D. J Hegeman, Q. Cui, D. A Kelter, M. P Fiske, K. E Glajch, J. E Pitt, T. Y Huang, N. J Justice, and C S. Chan. 2015. *Parvalbumin+ neurons and npas1+ neurons are distinct neuron classes in the mouse external globus pallidus*, *Journal of Neuroscience* **35**, no. 34, 11830–11847.
- Isett, B. R, K. P Nguyen, J. C Schwenk, J. R Yurek, C. N Snyder, M. V Vounatsos, K. A Adegbesan, U. Ziausyte, and A. H Gittis. 2023. *The indirect pathway of the basal ganglia promotes transient punishment but not motor suppression*, *Neuron*.
- Karube, F., S. Takahashi, K. Kobayashi, and F. Fujiyama. 2019. *Motor cortex can directly drive the globus pallidus neurons in a projection neuron type-dependent manner in the rat*, *Elife* **8**, e49511.
- Ketzel, M. and G. Silberberg. 2021. *Differential synaptic input to external globus pallidus neuronal subpopulations in vivo*, *Neuron* **109**, no. 3, 516–529.
- Kita, H. 2007. *Globus pallidus external segment*, *Progress in brain research* **160**, 111–133.
- Kita, H., Y. Tachibana, A. Nambu, and S. Chiken. 2005. *Balance of monosynaptic excitatory and disynaptic inhibitory responses of the globus pallidus induced after stimulation of the subthalamic nucleus in the monkey*, *Journal of Neuroscience* **25**, no. 38, 8611–8619.
- Kravitz, A. V, B. S Freeze, P. R. Parker, K. Kay, M. T Thwin, K. Deisseroth, and A. C Kreitzer. 2010. *Regulation of parkinsonian motor behaviours by optogenetic control of basal ganglia circuitry*, *Nature* **466**, no. 7306, 622–626.

- Labouesse, M., A. Torres-Herraez, M. Chohan, J. Villarin, J. Greenwald, X. Sun, M. Zahran, A. Tang, S. Lam, J. Veenstra-VanderWeele, et al. 2023. *A non-canonical striatopallidal "go" pathway that supports motor control*.
- Lappin, J. S and C. W Eriksen. 1966. *Use of a delayed signal to stop a visual reaction-time response.*, Journal of Experimental Psychology **72**, no. 6, 805.
- Lazarus, M., J.-F. Chen, Y. Urade, and Z.-L. Huang. 2013. *Role of the basal ganglia in the control of sleep and wakefulness*, Current opinion in neurobiology **23**, no. 5, 780–785.
- Lilascharoen, V., E. H.-J. Wang, N. Do, S. C. Pate, A. N. Tran, C. D. Yoon, J.-H. Choi, X.-Y. Wang, H. Pribiag, Y.-G. Park, et al. 2021. *Divergent pallidal pathways underlying distinct parkinsonian behavioral deficits*, Nature neuroscience **24**, no. 4, 504–515.
- Logan, G. D, R. J Schachar, and R. Tannock. 1997. *Impulsivity and inhibitory control*, Psychological science **8**, no. 1, 60–64.
- Mallet, N., B. R Micklem, P. Henny, M. T Brown, C. Williams, J P. Bolam, K. C Nakamura, and P. J Magill. 2012. *Dichotomous organization of the external globus pallidus*, Neuron **74**, no. 6, 1075–1086.
- Mallet, N., A. Pogosyan, L. F Márton, J P. Bolam, P. Brown, and P. J Magill. 2008. *Parkinsonian beta oscillations in the external globus pallidus and their relationship with subthalamic nucleus activity*, Journal of neuroscience **28**, no. 52, 14245–14258.
- Mallet, N., R. Schmidt, D. Leventhal, F. Chen, N. Amer, T. Boraud, and J. D Berke. 2016. *Arkypallidal cells send a stop signal to striatum*, Neuron **89**, no. 2, 308–316.
- Mastro, K. J, R. S Bouchard, H. A. Holt, and A. H Gittis. 2014. *Transgenic mouse lines subdivide external segment of the globus pallidus (gpe) neurons and reveal distinct gpe output pathways*, Journal of Neuroscience **34**, no. 6, 2087–2099.
- Mastro, K. J, K. T Zitelli, A. M Willard, K. H Leblanc, A. V Kravitz, and A. H Gittis. 2017. *Cell-specific pallidal intervention induces long-lasting motor recovery in dopamine-depleted mice*, Nature neuroscience **20**, no. 6, 815–823.
- Meyer, H. C and D. J Bucci. 2016. *Neural and behavioral mechanisms of proactive and reactive inhibition*, Learning & Memory **23**, no. 10, 504–514.
- Mink, J. W. 1996. *The basal ganglia: focused selection and inhibition of competing motor programs*, Progress in neurobiology **50**, no. 4, 381–425.
- Mitchell, S., R. Richardson, F. Baker, and M. DeLong. 1987. *The primate globus pallidus: neuronal activity related to direction of movement*, Experimental Brain Research **68**, no. 3, 491–505.
- Mizutani, K., S. Takahashi, S. Okamoto, F. Karube, and F. Fujiyama. 2017. *Substance p effects exclusively on prototypic neurons in mouse globus pallidus*, Brain Structure and Function **222**, no. 9, 4089–4110.
- Nambu, A., S. Chiken, P. Shashidharan, H. Nishibayashi, M. Ogura, K. Kakishita, S. Tanaka, Y. Tachibana, H. Kita, and T. Itakura. 2011. *Reduced pallidal output causes dystonia*, Frontiers in Systems Neuroscience **5**, 89.
- Nambu, A., H. Tokuno, I. Hamada, H. Kita, M. Imanishi, T. Akazawa, Y. Ikeuchi, and N. Hasegawa. 2000. *Excitatory cortical inputs to pallidal neurons via the subthalamic nucleus in the monkey*, Journal of neurophysiology **84**, no. 1, 289–300.
- Nambu, A., H. Tokuno, and M. Takada. 2002. *Functional significance of the cortico-subthalamo-pallidal 'hyperdirect' pathway*, Neuroscience research **43**, no. 2, 111–117.
- Nevado-Holgado, A. J, N. Mallet, P. J Magill, and R. Bogacz. 2014. *Effective connectivity of the subthalamic nucleus-globus pallidus network during parkinsonian oscillations*, The Journal of physiology **592**, no. 7, 1429–1455.
- Pamukcu, A., Q. Cui, H. S Xenias, B. L Berceau, E. C Augustine, I. Fan, S. Chalasan, A. W Hantman, T. N Lerner, S. M Boca, et al. 2020. *Parvalbumin+ and npas1+ pallidal neurons have distinct circuit topology and function*, Journal of Neuroscience **40**, no. 41, 7855–7876.

- Phillips, R. S., I. Rosner, A. H. Gittis, and J. E. Rubin. 2020. *The effects of chloride dynamics on substantia nigra pars reticulata responses to pallidal and striatal inputs*, *Elife* **9**, e55592.
- Plenz, D. and S. T. Kital. 1999. *A basal ganglia pacemaker formed by the subthalamic nucleus and external globus pallidus*, *Nature* **400**, no. 6745, 677–682.
- Qiu, M., M. C. Chen, J. Wu, D. Nelson, and J. Lu. 2016. *Deep brain stimulation in the globus pallidus externa promotes sleep*, *Neuroscience* **322**, 115–120.
- Sano, H., S. Chiken, T. Hikida, K. Kobayashi, and A. Nambu. 2013. *Signals through the striatopallidal indirect pathway stop movements by phasic excitation in the substantia nigra*, *Journal of Neuroscience* **33**, no. 17, 7583–7594.
- Saunders, A., K. W. Huang, and B. L. Sabatini. 2016. *Globus pallidus externus neurons expressing parvalbumin interconnect the subthalamic nucleus and striatal interneurons*, *PloS one* **11**, no. 2, e0149798.
- Saunders, A., I. A. Oldenburg, V. K. Berezovskii, C. A. Johnson, N. D. Kingery, H. L. Elliott, T. Xie, C. R. Gerfen, and B. L. Sabatini. 2015. *A direct gabaergic output from the basal ganglia to frontal cortex*, *Nature* **521**, no. 7550, 85–89.
- Schmidt, R. and J. D. Berke. 2017. *A pause-then-cancel model of stopping: evidence from basal ganglia neurophysiology*, *Philosophical Transactions of the Royal Society B: Biological Sciences* **372**, no. 1718, 20160202.
- Starr, P. A., G. A. Kang, S. Heath, S. Shimamoto, and R. S. Turner. 2008. *Pallidal neuronal discharge in huntington's disease: support for selective loss of striatal cells originating the indirect pathway*, *Experimental neurology* **211**, no. 1, 227–233.
- Starr, P. A., G. M. Rau, V. Davis, W. J. Marks Jr, J. L. Ostrem, D. Simmons, N. Lindsey, and R. S. Turner. 2005. *Spontaneous pallidal neuronal activity in human dystonia: comparison with parkinson's disease and normal macaque*, *Journal of neurophysiology* **93**, no. 6, 3165–3176.
- Stephenson-Jones, M., K. Yu, S. Ahrens, J. M. Tucciarone, A. N. van Huijstee, L. A. Mejia, M. A. Penzo, L.-H. Tai, L. Wilbrecht, and B. Li. 2016. *A basal ganglia circuit for evaluating action outcomes*, *Nature* **539**, no. 7628, 289–293.
- Terman, D., J. E. Rubin, A. Yew, and C. Wilson. 2002. *Activity patterns in a model for the subthalamopallidal network of the basal ganglia*, *Journal of Neuroscience* **22**, no. 7, 2963–2976.
- Turner, R. S. and M. E. Anderson. 1997. *Pallidal discharge related to the kinematics of reaching movements in two dimensions*, *Journal of neurophysiology* **77**, no. 3, 1051–1074.
- Turner, R. S. and M. E. Anderson. 2005. *Context-dependent modulation of movement-related discharge in the primate globus pallidus*, *Journal of Neuroscience* **25**, no. 11, 2965–2976.
- Verbruggen, F., A. R. Aron, G. P. Band, C. Beste, P. G. Bissett, A. T. Brockett, J. W. Brown, S. R. Chamberlain, C. D. Chambers, H. Colonius, et al. 2019. *A consensus guide to capturing the ability to inhibit actions and impulsive behaviors in the stop-signal task*, *elife* **8**, e46323.
- Vetrivelan, R., M.-H. Qiu, C. Chang, and J. Lu. 2010. *Role of basal ganglia in sleep–wake regulation: neural circuitry and clinical significance*, *Frontiers in neuroanatomy* **4**, 145.
- Vince, M. A. 1948. *The intermittency of control movements and the psychological refractory period*, *British Journal of Psychology* **38**, no. 3, 149.
- Wadsley, C. G., J. Cirillo, A. Nieuwenhuys, and W. D. Byblow. 2022. *Stopping interference in response inhibition: behavioral and neural signatures of selective stopping*, *Journal of Neuroscience* **42**, no. 2, 156–165.
- Wallace, M. L., A. Saunders, K. W. Huang, A. C. Philson, M. Goldman, E. Z. Macosko, S. A. McCarroll, and B. L. Sabatini. 2017. *Genetically distinct parallel pathways in the entopeduncular nucleus for limbic and sensorimotor output of the basal ganglia*, *Neuron* **94**, no. 1, 138–152.

- Wessel, J. R. 2018. *Surprise: A more realistic framework for studying action stopping?*, Trends in cognitive sciences **22**, no. 9, 741–744.
- Williams, B. R., J. S Ponesse, R. J Schachar, G. D Logan, and R. Tannock. 1999. *Development of inhibitory control across the life span.*, Developmental psychology **35**, no. 1, 205.
- Yoshida, A. and M. Tanaka. 2016. *Two types of neurons in the primate globus pallidus external segment play distinct roles in antisaccade generation*, Cerebral Cortex **26**, no. 3, 1187–1199.

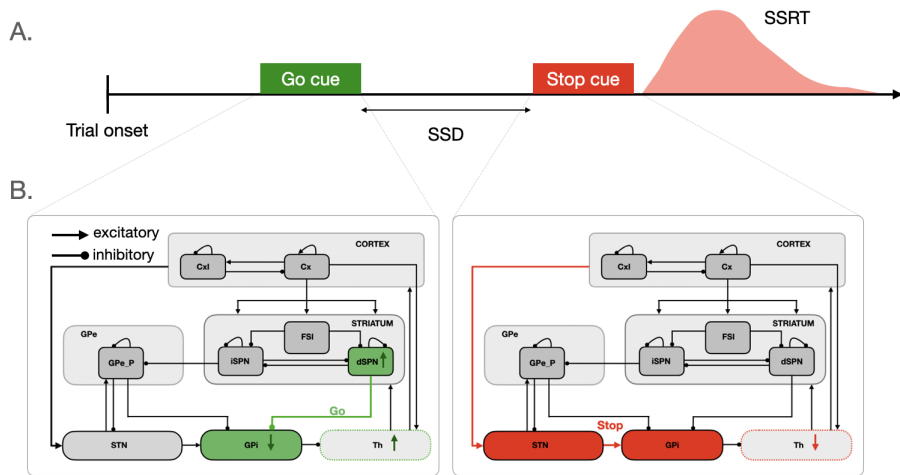


FIGURE 1 Classical model of reactive inhibition. **A.** Schematics of the general stop signal task paradigm. After the trial onset, a primary stimulus (the “Go” cue, in green) is presented to participants, signaling that they should quickly respond. When a secondary stop stimulus follows (“Stop” cue, in red), participants are expected to withhold their response. The delay between the presentation of the two stimuli is known as stop signal delay (SSD). A useful measure that reflects the time participants take to “react” to the “Stop” cue is the stop signal reaction time (SSRT). **B.** Network schematics showing the dynamics that take place within the CBGT circuit when “Go” and “Stop” cues are presented, respectively, according to the classical model. The presentation of the “Go” cue triggers the activation of the direct pathway (green regions), causing the disinhibition of the thalamus. In the classical view, the “Stop” cue triggers the engagement of the hyperdirect pathway (red regions), increasing the suppression of thalamic activity.

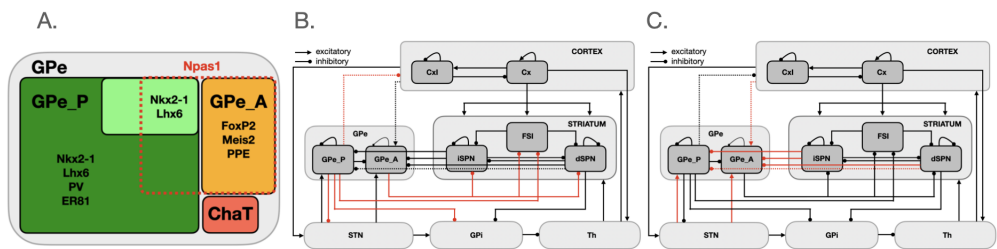


FIGURE 2 Cellular composition and connectivity of the GPe. **A.** Schematic diagram illustrating the cellular composition of the GPe. The areas of the rectangles represent the approximate sizes of the corresponding neuron classes. Prototypical neurons, which constitute ~70% of GPe inhibitory cells, can be subdivided according to whether or not they are PV-expressing. Arkypallidal neurons represent ~20% of GPe GABAergic neurons; all of them and a small pool of prototypical neurons are Npas1-expressing. ChaT-expressing neurons constitute ~5% of the total GPe neuron population, showing no overlap with other known clusters of GPe cells. **B.** Network diagram of the CBGT circuit, highlighting (in red) the efferent projections of GPe. **C.** Network diagram of the CBGT circuit, highlighting (in red) the afferent projections to GPe.

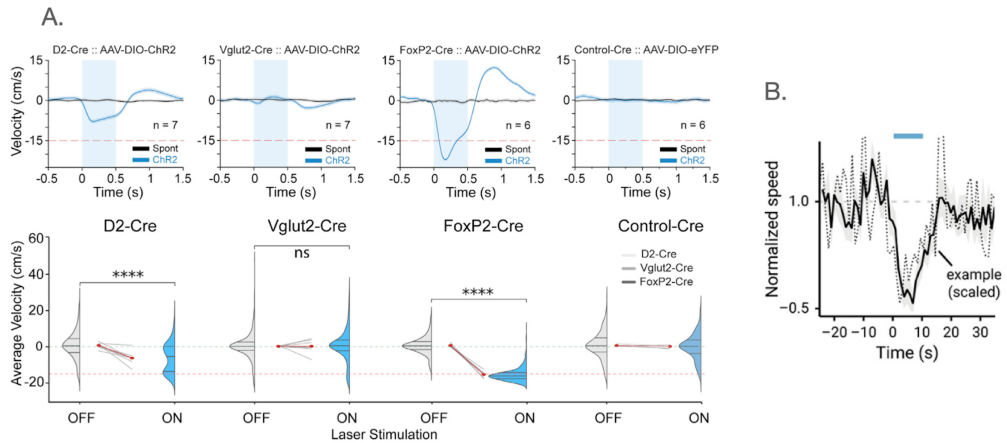


FIGURE 3 Optogenetic stimulation of arkyallid neurons induces locomotion inhibition. A. Activation of arkyallid neurons is sufficient to inhibit locomotion (reprinted with permission from (Aristieta et al. 2021)). Above, average velocity induced by optogenetic stimulation (in blue) or during spontaneous locomotion (in black) in D2-Cre::AAV-ChR2 (i.e. iSPNs optogenetic stimulation), Vglut2-Cre::AAV-ChR2 (i.e. STN optogenetic stimulation), FoxP2-Cre::AAV-ChR2 (i.e. GPe_A optogenetic stimulation), and control-Cre::AAV-eYFP mice, which were trained to walk continuously for five minutes on a motorized treadmill belt (velocity of 15cm/s) for at least one week before the beginning of optogenetic experiments. Below, velocity population graphs showing the velocity distributions during laser ON/OFF periods in the various animal groups. The gray lines represent individual animals while the red lines represent the mean between animals. The dashed red line shows the velocity of the treadmill. B. Optogenetic stimulation of GPe Npas1-expressing neurons suppresses locomotion (reprinted with permission from (Pamukcu et al. 2020)). Plot showing the relationship between normalized speed and time when stimulating GPe Npas1-expressing neurons. Blue horizontal bar indicates the duration (10 s) of light delivery. The dotted horizontal line indicates the baseline locomotor activity level. The black solid trace is the population mean calculated from all mice; shading indicates the SEM. The black dotted trace is a representative example from a single mouse; data were scaled to facilitate comparison.

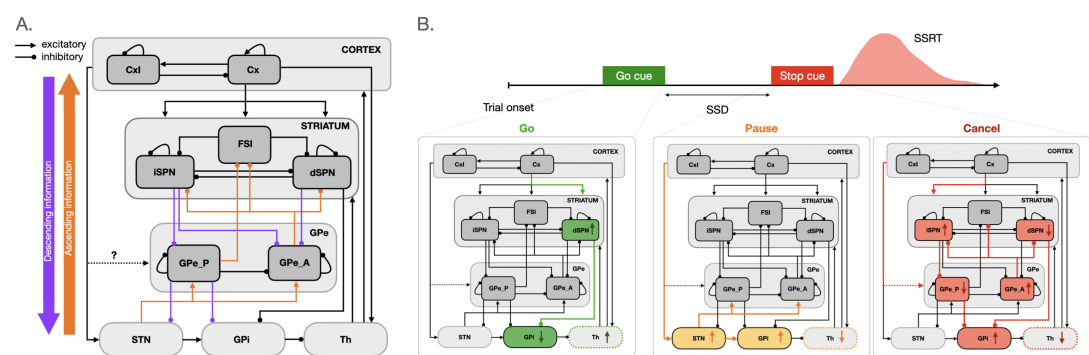
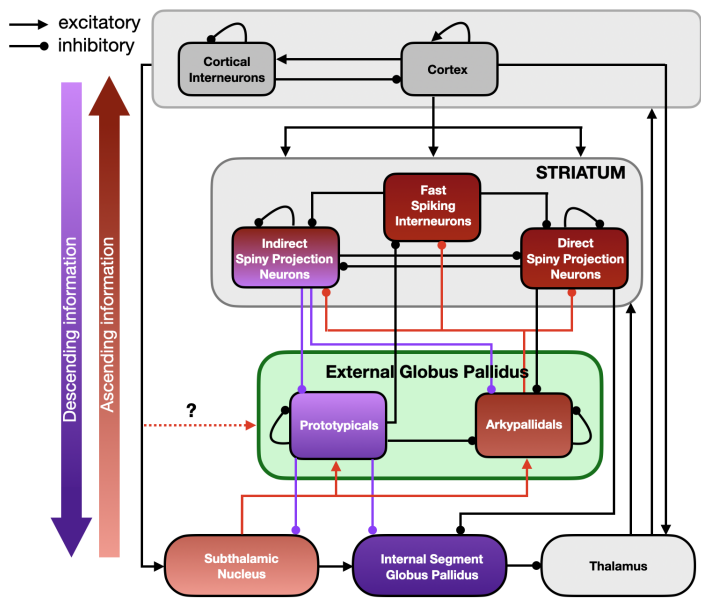


FIGURE 4 Casting the GPe as a central hub of the basal ganglia with a key role in regulating information flow in cortico-basal ganglia-thalamic circuits. A. CBGT network schematic. Here, the GPe is placed in a central position, regulating the bidirectional information flow across the circuit (descending pathways in purple, ascending pathways in orange). The cortical projection to GPe is indicated as a dotted line with a question mark because there is still little known about its nature. B. Network dynamics for action suppression according to the Pause-then-Cancel model. As in the classical view, the “Go” cue triggers the activation of the direct pathway, leading to thalamic disinhibition. In this model, the “Stop” signal triggers two subprocesses: the first fast process activated is a “pause” process, induced by the engagement of the hyperdirect pathway (in yellow), which transiently increases the inhibition of the thalamus. The second subprocess corresponds to the “cancel” stage of the model when arky pallidal neurons become engaged via stop-related signals from the STN or through direct cortical inputs and effectively relay this instruction upwards to the striatum.

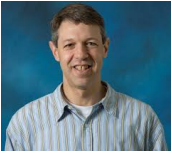


GRAPHICAL ABSTRACT The GPe regulates information flow in cortico-basal ganglia-thalamic circuits, relaying ascending and descending signals and serving as a central hub within the CBGT network.



dark fate...

Cristina Giossi Spaghetti, again?? No longer able to bear her gustatory ennui, Cristina Giossi hopped on the first available flight out of Italy and found herself on a strange island in an obscure body of water apparently called the 'Balearic Sea'. Realizing that she had no practical marketable skills, she naturally retreated to academia, enrolling in the first available Ph.D. program that she could find. If only her arkipallidal neurons had been more active, she might have avoided this



with the typical ivory tower abode, he boldly chose to reside in a goose house, where the honks of his feathered roommates add an unconventional soundtrack to his mathematical musings.

Jonathan Rubin, Ph.D. Jonathan Rubin ascended to the role of math maestro extraordinaire within the realm of Applied Mathematics in 1996, diving headfirst into the wild, wacky world of numbers and equations. Several years later, in 2000, he expanded his scholarly domain to the University of Pittsburgh, where he found his academic home and halted his program of conquest, opting instead for a settled and focused approach to his pursuits. Not content



Aryn Gittis, Ph.D. Aryn Gittis was found wondering the halls of the University of California, San Francisco shouting "Lasers! We'll put lasers in their brains!" That is precisely what she did. She now commands an army of laser controlled mammals in the Department of Biology at Carnegie Mellon University, where many suspect she that is hatching a plan to dominate the world, but Dr. Gittis assures everyone she is simply trying to understand the brain. Time will tell.



This reveals why Dr. Verstynen now finds himself endowed with an insatiable craving for late-night brain snacks – both literally and academically.

Timothy Verstynen, Ph.D. Timothy Verstynen lived a curious double life: a daytime neuroscientist delving into neural networks mysteries, and by moonlight, a zombie expert dissecting undead brains to reveal the secrets of their decision-making. But fate intervened when he found a hidden lab in the CMU campus, known as CoAx lab, right under the roof of the Psychology Department, and accidentally spilled a serum meant to save humanity from zombies, only to join their ranks himself.



who mistakenly wandered into her classroom thinking it was an introduction to biology class and stayed for the semester for the wonder of pure math itself.

Catalina Vich, Ph.D. It is unclear from whence Catalina Vich originated, but records indicate her presence on the island of Mallorca dating back several centuries. Legend has it that Leibniz walked away with the idea of calculus after spending a winter raising sheep on Catalina's farm at Puig de Randa. Catalina has spent her time hiding from wayward tourists (and occasional invaders) by teaching mathematics at the Universitat de les Illes Balears, mainly to students