

1 **Title:** Phylogenetic dependence of plant-soil feedback promotes rare species in a subtropical forest

2 **Running title:** Plant-soil feedback and species coexistence

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35 **Abstract**

36 The widespread observation that rare species have stronger conspecific plant-soil feedback (PSF) than
37 common species raises more questions than answers on how rare species can possibly win the dance
38 with abundant species. Here, we test soil feedback effect of phylogenetically related species on seedlings
39 of contrasting local abundance in a subtropical forest. The results showed that although rare species
40 suffered strong negative PSF in soils of conspecifics or phylogenetically close relatives, no such
41 feedback was found in the soils of distant relatives. In contrast, although common species had weak
42 conspecific PSF, they suffered consistently strong heterospecific soil feedback. These mechanisms
43 ensure that rare species would fare well in the neighborhood of phylogenetically distant heterospecifics
44 but do poorly under their close relatives, while common species perform relatively well in their own
45 neighborhood but poorly in others'. This phylogenetic conservatism in PSF facilitates the persistence of
46 rare species in a community.

47 **Introduction**

48 Negative plant-soil feedback (PSF) has been considered to play a major role in regulating plant
49 diversity worldwide, from tropical to temperate forests and grasslands (Packer & Clay 2000;
50 Klironomos 2002; Petermann *et al.* 2008; Mangan *et al.* 2010b; Bagchi *et al.* 2014; Liu *et al.* 2015; Levi
51 *et al.* 2019). The process accumulates host-specific soil enemies around plant adults that in turn reduce
52 the seedling performance of conspecifics relative to heterospecifics (Bever *et al.* 1997; Bever 2003) so
53 that to prevent dominant species from competitively excluding others (Connell *et al.* 1984; Bever 2003;
54 Chesson & Kuang 2008). However, over the past two decades, much empirical evidence has shown that
55 rare species in plant communities suffer stronger conspecific soil feedback than common species
56 (Klironomos 2002; Mangan *et al.* 2010b; MacDougall *et al.* 2011; Rutten *et al.* 2016), and other studies
57 that model long-term tree mortality data have also shown that their statistical results are consistent with
58 the prediction of stronger rare species PSF although those studies themselves are not a test of PSF
59 (Comita *et al.* 2010; Johnson *et al.* 2012). However, these results are not universally unequivocal
60 (Reinhart 2012; Liu *et al.* 2015; Maron *et al.* 2016; Kempel *et al.* 2018) and they have been questioned
61 to be biased due to study design and analytical approaches (Detto *et al.* 2019). It remains an open
62 question how plant-soil feedback checks the dominance of common species and prevents rare species
63 from competitive exclusion if the negative PSF is stronger for rare species than common species.

64 Critical to testing biodiversity maintenance is the quantification of the relative strength of intra- vs
65 interspecific density (or frequency) dependence (Chesson 2000; Adler *et al.* 2018). However, similar to
66 competition studies where most experiments were not designed to compare the per-capita strength of
67 intra- and interspecific competition needed to test species coexistence (Inouye 2001), few studies on the
68 PSF effect quantify the per capita effects of host-specific soil microbiota on conspecific and
69 heterospecific performance. For example, as a primary mechanism of PSF, the Janzen-Connell

70 hypothesis stipulates that host-specific natural enemies in the neighborhood of adult trees limit
71 conspecific regeneration (Janzen 1970; Connell 1971), thus releasing the vicinity of the adult trees for
72 other species to colonize. While the reduced conspecific spatial aggregation is necessary to maintaining
73 biodiversity, little is known how well other species would actually fare in the released space – is the
74 conspecific suppression by the natural enemies strong enough to allow heterospecifics to overwhelm the
75 focal species, or hopelessly weak so that the focal species continues to hold the space in quantity?

76 Plants interact with not only conspecifics but also heterospecifics as seed dispersal frequently leads
77 to seedlings growing near heterospecific adults (Levine & Murrell 2003). It is thus important to
78 understand the effect of soil feedback on the performance of heterospecific seedlings. Fig. 1
79 hypothesizes the possibilities of the relative performance of seedlings of rare and common species under
80 conspecific and heterospecific adults when rare species have stronger conspecific negative PSF. A
81 species could suffer strong conspecific negative PSF but escapes the effect under heterospecific adults
82 (e.g., the common species in Fig. 1a or the rare species in Fig. 1b) or be insensitive to neighbor identity
83 (e.g., the rare species in Fig. 1a or the common species in Fig. 1b). Species coexistence is not possible if
84 rare species suffer stronger suppression than common species, regardless under which adults (Fig. 1a
85 and b). Only in the case of Fig. 1c where common species experience stronger heterospecific negative
86 PSF than rare species can the two species coexist.

87 Much empirical evidence has shown that PSF is phylogenetically dependent, i.e., phylogenetically
88 related host species share common pathogens and the sharing decays phylogenetically (Gilbert & Webb
89 2007; Liu *et al.* 2012). This phylogenetic conservatism in plant-soil microbiota interactions favors plant
90 recruitment under heterospecific trees, which is especially important for species with high dispersal
91 ability, low density, or those having fewer close relatives (Levine & Murrell 2003; Parker *et al.* 2015).
92 Because of this phylogenetic conservatism, plants would suffer from stronger negative effects when

93 exposed to soil microbiota inoculated by phylogenetically closer heterospecifics (Brandt *et al.* 2009;
94 Anacker *et al.* 2014; Crawford *et al.* 2019). Phylogenetic conservatism in soil feedback effect also varies
95 across focal plant species (Brandt *et al.* 2009; Anacker *et al.* 2014; Sweet & Burns 2017; Wilschut *et al.*
96 2019), arising from the difference in the degree of specificity of soil microbiota in association with host
97 plants. For example, plants associated with more host-specialized enemies tend to exhibit stronger
98 phylogenetic conservatism in the strength of negative PSF, while those with more generalized enemies
99 do not (Liu *et al.* 2012; Crawford *et al.* 2019). This phylogenetic variation in PSF inevitably leads to
100 different relative performance of species under their own adults versus heterospecific adults, of different
101 phylogenetic relatedness, as illustrated in Fig. 1, and it is critical for explaining rare species maintenance
102 in facing stronger negative PSF than abundant species.

103 To address the question of how phylogenetic dependence in soil feedback effect may promote rare
104 species in natural communities, we conducted a shade-house sterilization experiment and a reciprocal
105 field experiment on seedling growth of seven species in a subtropical forest in southern China. The
106 selected species comprised paired sister species with contrasting local abundances. Conspecific and
107 heterospecific soil feedback effects on seedling growth were quantified and the phylogenetic signal in
108 plant-soil feedback was tested. Our results showed that rare species experienced stronger conspecific
109 negative soil feedback than common species. They also suffered stronger heterospecific negative soil
110 feedback under closely than distantly related trees and the feedback disappeared with further increase in
111 phylogenetic distance. In contrast, common species experienced consistently negative soil feedback
112 from heterospecifics, independent of phylogenetic distance. These results provide empirical evidence
113 showing that rare species are promoted by soil feedback effects that favor their recruitment under
114 phylogenetically distant heterospecific trees where the recruitment of common species is restricted, as
115 predicted by Fig. 1c.

116 **Materials and Methods**

117 **Study site, species selection and seed germination**

118 The study site is located in the Heishiding Nature Reserve (N 23°27', E 111°53'; altitude: 150-927
119 m asl), an evergreen broadleaf subtropical forest in southern China. Tree species, *Ormosia fordiana*, *O.*
120 *glaberrima*, *Quercus auricoma* and *Q. macrocalyx* were selected in both shade-house and field
121 experiments, constituting two pairs of sister species with distinctly low and high abundances in a 50-ha
122 stem-mapped plot (established in 2012) where our experiments were conducted (see Table S1 and Fig.
123 S1 in Supporting Information). We additionally used *Canarium album* and *Helicia reticulata* in the
124 shade-house (*ex situ*) experiment and *Q. pachyloma* in the field (*in situ*) experiment (Table S1). Species
125 sets were not identical in the two experiments because seeds for some species were not available and we
126 preferred species from the same genus in the field experiment to generate a phylogenetic distance
127 gradient of conspecific, congeneric and heterofamilial species. Seeds were collected from the study site
128 during the autumn and winter (September to December) of 2016, immediately surface-sterilized (1
129 minute in 70% ethanol, 3 minute 50% commercial bleach and 1 minute 70% ethanol, and 1 minute
130 distilled water), and then stored in a refrigerator at 4°C. In January 2017, seeds were sowed in sterilized
131 sandy soil and germinated about 8 weeks later. At the beginning of our experiments in May 2017, we
132 randomly selected 22 seedlings for each of the seven species. The stem height of each seedling was
133 measured, and the total leaf area was also measured using an LI-COR LI-3000C portable leaf area meter
134 to the nearest 0.01 cm². The aboveground biomass of each individual was then weighed after oven-dried
135 at 70°C for 48 h in laboratory. These data were used to estimate initial aboveground biomass for
136 seedlings that were used in shade-house and field experiments as described below.

137

138 **Shade-house sterilization experiment**

139 A shade-house experiment was designed to test whether the variation in soil feedback from
140 different phylogenetically related species between rare and common species is mediated by soil microbe.
141 The experiment considered three factors: focal species with contrast abundance, phylogenetic distance
142 of soil source species to focal species, and sterilization treatment (soil with gamma radiation versus
143 without), as described below. In March 2017, for each of the six species, *C. album*, *H. reticulata*, *O.*
144 *fordiana*, *O. glaberrima*, *Q. auricoma*, and *Q. macrocalyx*, we collected soils beneath three adults of
145 each conspecific, congeneric, heterogeneric (confamilial), and heterofamilial species (Table S2). For
146 each of the three replicate adults, topsoil (*ca.* 10 cm in depth) at three randomly set directions were
147 sampled and mixed to give a representative sample of soil of the adult. Soil samples were then sieved
148 using meshes (0.2 cm in diameter) to remove any stones, roots and seeds, and divided into two parts.
149 One was sterilized using gamma radiation with a dose of 25 Gy and the other was used as a microbe
150 inoculum source. The species from which soils were sampled and used for cross seedling planting is
151 called soil-source species (also see Fig. 2).

152 To eliminate the interference of nutrition differences, we randomly collected background soils from
153 the study area (not specifically associating with the study species) and sterilized with gamma radiation.
154 These background soils were then mixed with the sterilized or live soil sampled from the soil-source
155 species at a proportion of 93:7 volume/volume following Teste *et al.* (2017). The mixed soils were filled
156 into plastic pots (15 cm in diameter, 17 cm in height) and one seedling was then transplanted into each
157 pot. In total, 48 combinations of experimental treatments (6 seedling species \times 4 soil sources \times 2
158 sterilization treatments) were replicated eight times, resulting in 384 pots. The position of the pots was
159 randomly shifted every two months and seedlings were watered weekly. After transplanted in May 2017,
160 the stem height and the total leaf area of each seedling were measured. After one year growth, seedlings
161 were harvested, and the aboveground dry weight was determined in June 2018.

162 **Reciprocal field experiment**

163 A reciprocal seedling planting experiment was conducted to test how the strength of soil feedback
164 from different phylogenetically related species is related to the abundance of host species. In March
165 2017, for each of the five species, *O. fordiana*, *O. glaberrima*, *Q. auricoma*, *Q. macrocalyx*, *Q.*
166 *pachyloma*, three adults with diameter at breast height larger than 20 cm were selected in the field, and
167 these adults were required to have no neighboring adults belonging to any of the five species within 20
168 meters. 50 seedlings (10 for each species) were transplanted to a 1.5×2.5 m seedling quadrat beneath a
169 selected adult, with a random mixture of the seedlings in five rows (10 seedlings per row) and 20 cm
170 apart from each other. To minimize the effect of possible transplanting injuries, we replaced the dead or
171 dying seedlings within the first two weeks. In total, 750 seedlings (5 host species × 5 seedling species ×
172 3 host adult replicates × 10 seedling replicates) were planted in the field. Their initial stem height and
173 total leaf area were measured in May 2017. Seedlings were harvested and the aboveground dry weight
174 was determined in November 2018.

175

176 **Statistical analyses**

177 To estimate the initial aboveground biomass of each transplanted seedling, we used data from the
178 22 seedlings of each species harvested at the beginning of the experiment to model the aboveground dry
179 weight (B) as a function of the total leaf area (TLA) and stem height (H), with the linear form: $\log(B) =$
180 $\beta_0 + \beta_1 \cdot \log(TLA) + \beta_2 \cdot \log(H)$. The R^2 of these models was at least 0.78 for all the seven species
181 (Table S3), allowing reliable estimation of initial aboveground biomass using the stem height and total
182 leaf area of those transplanted seedlings. The relative growth rate (RGR) of seedlings was then
183 calculated using the equation: $RGR = (\log(\text{harvest biomass}) - \log(\text{initial biomass}))/\text{time interval}$.

184 For the shade-house experiment, plant-soil feedback was measured as a log-response ratio of RGR

185 between live and sterilized soils (Kempel *et al.* 2018), where the RGR in sterilized soils was averaged
186 on the eight replicates in each soil source treatment, and the RGR of each individual in live soils was
187 used, allowing evaluating the feedback strength for each seedling. By comparing the performance of
188 seedling growth between sterilized and live soils, we assessed the net effect of soil microbiota on
189 seedling performance, which was the overall effect of different soil microbiota including pathogenic and
190 mutualistic fungi. A negative value of plant-soil feedback indicates a lower growth rate in live than
191 sterile soils and reflects soil pathogenic effects dominate, while a positive value indicates a better growth
192 in live soil, reflecting mutualistic effects overwhelm.

193 For the field experiment, conspecific plant-soil feedback was defined as the difference of seedling
194 performance in ‘home vs. alien’ soils, which was calculated as a pairwise linear contrast of $RGR(A)_\alpha -$
195 $RGR(A)_\beta + RGR(B)_\beta - RGR(B)_\alpha$, where A refers to focal seedlings and B as the seedlings of another
196 species; α and β are the soils from adults A (home soil) and B (alien soil), respectively; and $RGR(A)_\alpha$,
197 for example, indicates the relative growth rate of A seedlings in conspecific (home) soils α (Bever 1994,
198 2003; Bever *et al.* 1997). The averaged RGR of the seedlings per species per seedling quadrat was
199 calculated to represent the growth of seedlings in that soil. There were three RGR values in home soil
200 and 12 in alien soil for each of the five species, resulting in a total of 180 pairwise feedbacks.

201 Similarly, the congeneric plant-soil feedback was defined as the relative seedling performance of
202 two species, e.g., focal species A and another species B , in soils from the congeneric species of A and
203 from B , respectively. The strength of congeneric PSF was calculated in the same way as that for
204 conspecific PSF shown in the above, where the home soil α was defined as soil from congenics. The
205 same procedure was also applied to calculate the strength of heterofamilial plant-soil feedback. In total,
206 288 congeneric and 432 heterofamilial soil feedbacks were calculated as each focal species in *Quercus*
207 had two congeneric and heterofamilial species, and each focal species in *Ormosia* had one congeneric

208 and three heterofamilial species in our study.

209 In both the shade-house and field experiments, linear mixed-effects models were used to test the
210 significance of phylogenetic relatedness between focal species and the soil-source species, the
211 abundance of focal species and their interaction term in determining the strength of PSF. The
212 phylogenetic relationships between focal species and the soil-source species (Fig. S2) were obtained by
213 pruning the GBOTB.extended mega-tree for our species set using the “phylo.maker” function of the
214 “V.PhyloMaker” package (Jin & Qian 2019). The mixed-effects models included phylogenetic distance,
215 abundance and their interaction as fixed effects while seedling species identity as a random effect using
216 “lme” function in “nlme” package of R software (Pinheiro & Bates 2000). To detect phylogenetic signal
217 in PSF, we used nonparametric Kruskal-Wallis rank-sum test and Dunnett modified Tukey-Kramer
218 multiple comparison test to test the variation in the effect of PSF from different phylogenetically related
219 host tree species on seedlings of focal species. We further used one-sided Wilcoxon signed-rank test to
220 test whether the strength of the soil feedback is significantly negative. All analyses were conducted in R
221 v.3.6.2 (<https://www.r-project.org/>).

222

223 **Results**

224 **Plant-soil feedback in the shade-house sterilization experiment**

225 The results from shade-house experiment showed that plant seedlings grew worse in live than in
226 sterilized soil (Fig. S3; two-sample Wilcoxon test, $W = 15206$, one-sided $P = 0.003$), suggesting an
227 overall negative effect of soil microbiota on seedling performance. The strength of this negative effect
228 decreased with the phylogenetic distance between the focal species and soil-source species for rare
229 species (Fig. 2; Kruskal-Wallis test, $\chi^2 = 30.53$, $df = 3$, $P < 0.001$), while this relationship disappeared
230 for common species (Fig. 2; Kruskal-Wallis test, $\chi^2 = 1.02$, $df = 3$, $P = 0.797$). Conspecific negative soil

231 feedback was found in both rare and common species (Fig. 2) and the strength of this effect was weaker
232 for common than rare species (two-sample Wilcoxon test, $W = 394$, two-sided $P = 0.011$). However,
233 common species were strongly affected by negative effects of heterofamilial soil feedback while rare
234 species were not (Fig. 2), and the strength of soil feedback from distantly related adults, e.g.
235 heterofamilials, was stronger for common than rare species (two-sample Wilcoxon test, $W = 146$, two-
236 sided $P = 0.003$).

237 The difference in PSF strength between common and rare species can be further compared by the
238 performance of the six species in conspecific versus heterospecific soils (Fig. 3). The rare species
239 experienced significant conspecific negative soil feedback, while the common species showed no or
240 weak negative conspecific PSF (Fig. 3a). In contrast, plant-soil feedback from heterofamilials showed
241 no effects of soil microbiota on rare species but strong negative feedback on common species (Fig. 3b).
242 These results were confirmed by a linear mixed-effects model where the strength of PSF was modeled
243 by abundance of focal species and the phylogenetic distance between the focal species and soil-source
244 species (Table 1). The model showed that the strength of PSF was positively associated with the
245 phylogenetic distance but negatively associated with the interaction term between the phylogenetic
246 distance and local abundance (Table 1; $P < 0.001$), suggesting that rare species had a stronger
247 relationship between the phylogenetic distance and the strength of soil feedback (Fig. 2). Overall, this
248 experiment showed significant phylogenetic conservatism in the effects of soil microbiota for rare
249 species but not for common species; and common species but not rare species suffered strong negative
250 soil feedbacks from phylogenetic-distant relatives, e.g., heterofamilials.

251

252 **Plant-soil feedback in the reciprocal field experiment**

253 In addition to the shade-house *ex situ* experiment, we also conducted an *in situ* seedling growth

254 experiment involving transplanting 750 seedlings into the forest (see Methods). 643 (86%) of the
255 seedlings still survived after 18 months when the experiment was ended (Table S4). We found that for
256 the rare species, the strength of plant-soil feedback from conspecifics was stronger than that from
257 heterofamilials but showed no difference between the conspecifics and congeners (Fig. 4 and Fig. S4a,
258 b; Kruskal-Wallis test, $\chi^2 = 25.55$, $df = 2$, $P < 0.001$). In contrast, seedlings of common species suffered
259 similar PSF strengths beneath the adults of conspecifics and heterospecifics (Fig. 4 and Fig. S4d, e;
260 Kruskal-Wallis test, $\chi^2 = 2.65$, $df = 2$, $P = 0.265$). Furthermore, for sister species pairs, the soils beneath
261 conspecific adults had a stronger negative effect on seedling growth of rare species but did not affect
262 that of common species (Fig. 5a), while the heterofamilial negative soil feedback was stronger for the
263 common than the rare species (Fig. 5b). The linear mixed-effects model for the strength of plant-soil
264 feedback showed consistent results with that of the shade-house experiment (Table 1).

265

266 **Discussion**

267 The observation of stronger conspecific PSF for locally rare than common species has been widely
268 reported (Klironomos 2002; Comita *et al.* 2010; Mangan *et al.* 2010b; MacDougall *et al.* 2011; Johnson
269 *et al.* 2012; McCarthy-Neumann & Ibáñez 2012; Rutten *et al.* 2016). This however raises more
270 questions than answers in explaining how rare species can possibly persist if they experience stronger
271 self-limitation (Metz *et al.* 2010; Zhu *et al.* 2015; Detto *et al.* 2019). Answers to this question are
272 ultimately important for understanding the maintenance of plant diversity. Our results from both shade-
273 house and field experiments consistently support the observation that rare species experience stronger
274 conspecific negative PSF than common species. In the meantime, we also found rare species had
275 stronger phylogenetic conservatism in plant-soil feedback than common species (Figs. 2 and 4; Table 1).
276 This conservatism in PSF shelters rare species from the effect of negative soil-feedback under distantly

277 related adults as hypothesized by Fig. 1c and provides them recruitment advantage under heterospecific
278 herd protection by lowering chance to encounter host-specific natural enemies (Peters 2003; Bachelot *et*
279 *al.* 2016). In contrast, although common species had weak negative conspecific PSF, they suffered
280 consistently strong heterospecific soil feedback, independent of phylogenetic distance (Figs. 3 and 5). In
281 other words, different from rare species, the recruitment of common species is less restricted under their
282 own adults but more restricted under heterospecifics. This contrast PSF between rare and common
283 species offers a promising answer to the puzzle of rare species-common species coexistence.

284 The variation in the strength of soil feedback from heterospecific adults in maintaining plant
285 diversity has been largely overlooked in the past. Our study highlights the importance of heterospecific
286 PSF in mediating seedling recruitment and species coexistence. Recruitment in the proximity of
287 heterospecifics is crucial for rare species whose seedlings are more likely surrounded by heterospecific
288 individuals than conspecific individuals. For example, the majority of seeds and seedlings of less
289 abundant species were found to be away from the conspecific adults in our study site (Fig. S5), and rare
290 tree species in most forests are more scattered in distribution than common species (He *et al.* 1997). The
291 variation in spatial distribution of recruitments suggested that heterospecific instead of conspecific soil
292 feedback is often the trouble that rare species have to deal with in nature, and thus weaker soil feedback
293 from heterospecific trees would facilitates recruitment of rare species and promote species coexistence.

294 Our finding that rare species had stronger conspecific but weaker heterospecific PSF than common
295 species suggests higher specificity in the effect of soil microbiota on rare species. This mechanism
296 would protect rare species from other adversaries, e.g., stochastic extinction. When facing disturbance,
297 the decrease in population density of rare species would reduce associated soil enemies and weaken the
298 strength of conspecific negative soil feedback. This, coupled with the fact that soil enemies maintained
299 by heterospecifics would have weak or no negative effects on rare species recruitment, could make rare

300 species rebound quickly to equilibrium abundance (Yenni *et al.* 2012, 2017; Liu *et al.* 2015). In contrast,
301 for common species, the decrease in population density does not necessarily promote recruitment as they
302 often experience strong negative feedback from the soil enemies of heterospecifics (McCarthy-Neumann
303 & Kobe 2010).

304 Our sterilized treatment that removed all soil microbiota including mutualists and antagonists
305 showed an overall positive effect on seedling performance (Fig. S3), implying the shared antagonists
306 (e.g., pathogens) among phylogenetically related plant species are responsible for the effect of negative
307 plant-soil feedback (Dyer *et al.* 2007; Gilbert & Webb 2007; Barrett & Heil 2012; Gilbert *et al.* 2012;
308 Parker *et al.* 2015; Kempel *et al.* 2018). A possible reason that phylogenetic signal in PSF was detected
309 for rare species but not for common species (Figs. 2 and 4) could be because rare species are associated
310 with more host-specific pathogens than common species. Pathogens of a focal tree, theoretically, can
311 spillover onto phylogenetically related species, and species with greater abundance are likely to produce
312 and receive more pathogen inocula which would dilute specialized pathogens in those abundant species
313 (Power & Mitchell 2004; Parker *et al.* 2015). An alternative possibility is that the specialized effect of
314 soil microbiota on rare species could result from the co-infection of (strong) pathogens and (weak)
315 mycorrhizal fungi (Corradi & Bonfante 2012). Common species are generally considered to have a
316 stronger association with mycorrhizal fungi and rely more on their mutualists compared to rare species
317 (Hart *et al.* 2003). The beneficial effects of mycorrhizal fungi could also be host specialized (Mangan *et*
318 *al.* 2010a), which counteracts negative pathogenic effects on seedling recruitment, and thus lead to the
319 host-general net effect of soil microbiota on common species (Bennett *et al.* 2017; García-Parisi &
320 Omacini 2017; Wang *et al.* 2019). Although we did not directly identify the specific pathogens and
321 mycorrhizal fungi, the net effects of these two fungal guilds have been suggested to be important in
322 plant-soil feedback (Bever *et al.* 2012). We suggest future studies to isolate these host-specific microbes

323 and also to analyze both fungal and bacterial community composition for better understanding the
324 effects of plant-soil feedback.

325 Our study quantified phylogenetic variation in plant-soil microbiota interactions between rare and
326 common species in a subtropical forest in China. We detected strong phylogenetic conservatism in the
327 strength of soil feedback for rare species but not for common species. Consequently, rare species
328 suffered stronger negative soil feedback from close relatives (e.g. conspecifics) but weaker or no
329 feedback from distant relatives (e.g. heterofamilials), while common species were less affected by
330 conspecific negative effects but more strongly affected by heterospecific negative effects. Our study
331 highlights the importance of quantifying asymmetric heterospecific plant-soil feedback for
332 understanding the role of plant-soil feedback in maintaining plant diversity, particularly rare species-
333 common species coexistence.

334

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468 **Tables and Figures**

469 **Table 1 Linear mixed-effect models for testing effects of phylogenetic relatedness, species relative**
 470 **abundance and their interaction term on the strength of plant-soil feedback.** The estimates of the
 471 effects are shown for the shade-house sterilization experiment and the reciprocal field experiment,
 472 respectively. Phylogenetic distance was measured as the distance between focal seedling species and
 473 soil-source species. Boldfaced values indicate significance at $P < 0.05$.

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	Effect	Estimate	Std.Error	$P(> t)$	
Shade-house experiment	Intercept	-1.331	0.402	0.002	
	Phylogenetic distance	0.006	0.001	< 0.001	
	Log(Abundance)	0.175	0.065	0.054	
	Phylogenetic distance	-0.001	0.0002	< 0.001	
	Random effect:		Std.Dev		
	Species (intercept)		0.120		
Field experiment	Intercept	-0.013	0.004	0.004	
	Phylogenetic distance	0.0001	0.00003	< 0.001	
	Log(Abundance)	0.001	0.0008	0.184	
	Phylogenetic distance	-0.00001	0.000005	0.006	
	Random effect:		Std.Dev		
	Species (intercept)		2.91e-07		

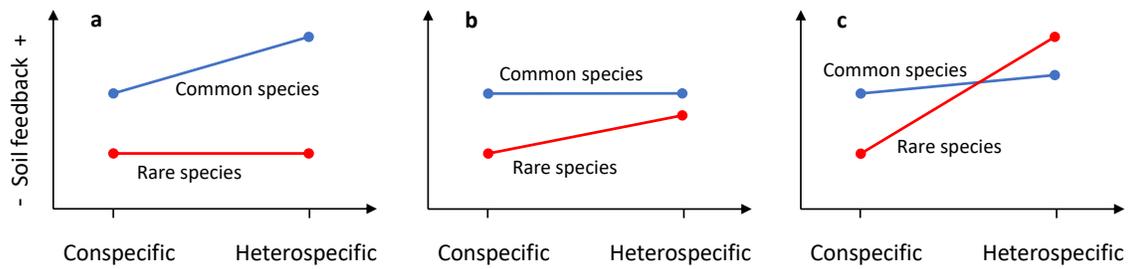
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481 **Figure 1 Variation in soil feedback from conspecifics and heterospecifics between rare and**

482 **common species.** The x axis indicates conspecific versus heterospecific soils and y axis indicates the

483 strength of soil feedback, ranging from negative to positive. **(a)** Rare species shows no change in

484 conspecific and heterospecific soil feedback strength, while common species has weaker heterospecific

485 effect (positive slope), leading to recruitment advantage in heterospecific soils, thus preventing

486 colonization of rare species. **(b)** Common species shows no change in soil feedback, while rare species

487 has weaker negative feedback in heterospecific soils, leading to recruitment advantage under

488 heterospecifics but the strength is not strong enough to overwhelm the common species. **(c)** Rare species

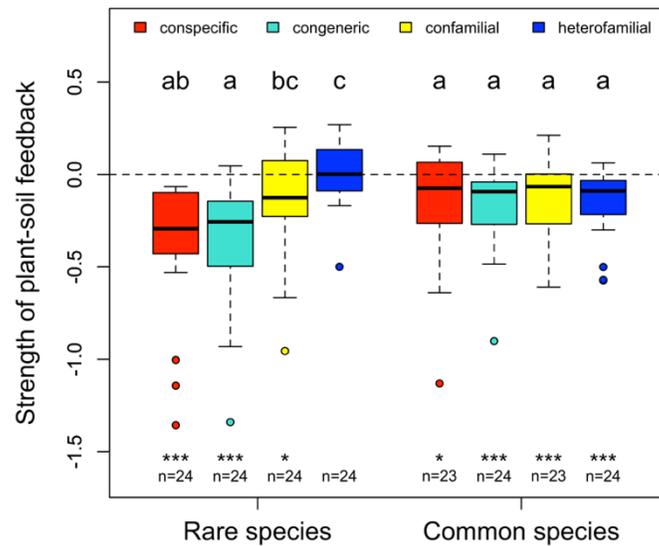
489 has stronger conspecific soil feedback but weaker heterospecific feedback than common species, leading

490 to species coexistence. Note negative slopes are unlikely to occur as predicted by the natural enemy

491 hypothesis of invasive species and thus are not considered here.

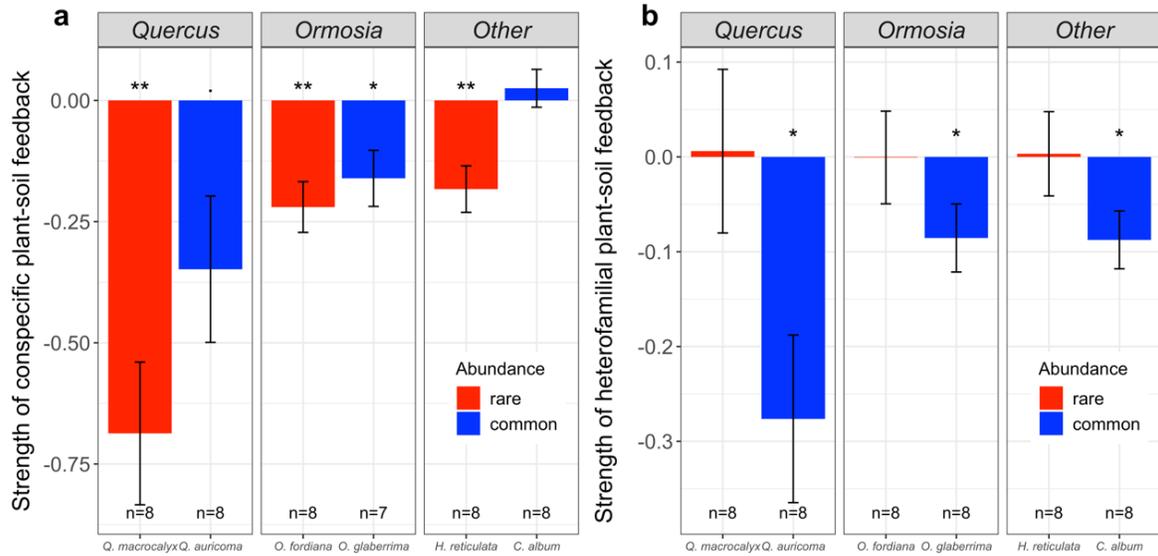
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495 **Figure 2** Boxplots showing the change of plant-soil feedback strength in relation to phylogenetic
 496 **distance between seedlings of focal species and soil-source species for rare and common species, as**
 497 **measured in shade-house experiment.** Rare species: *Helicia reticulata*; *Ormosia fordiana*; *Quercus*
 498 *macrocalyx*; Common species: *Canarium album*; *O. glaberrima*; *Q. auricoma*. “conspecific” means
 499 conspecific seedlings grew in soils taken under conspecific adults, “congeneric” means seedlings grew
 500 in soils taken from adult trees of congeneric sister species, etc. The species that donated soils is called
 501 “soil-source species”. Letter labels indicate the significant difference in the strength of plant-soil
 502 feedback across phylogenetic relatedness at $P < 0.05$ by the Dunnett's Modified Tukey-Kramer Pairwise
 503 Multiple Comparison Test. Asterisks indicate whether the strength of plant-soil feedback is significantly
 504 negative using one-sided Wilcoxon signed-rank test (* $P < 0.05$; ** $P < 0.01$; *** $P < 0.001$). The
 505 sample size for each box is given by n .



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508 **Figure 3 Variation in the strength of plant-soil feedback from (a) conspecifics and (b)**

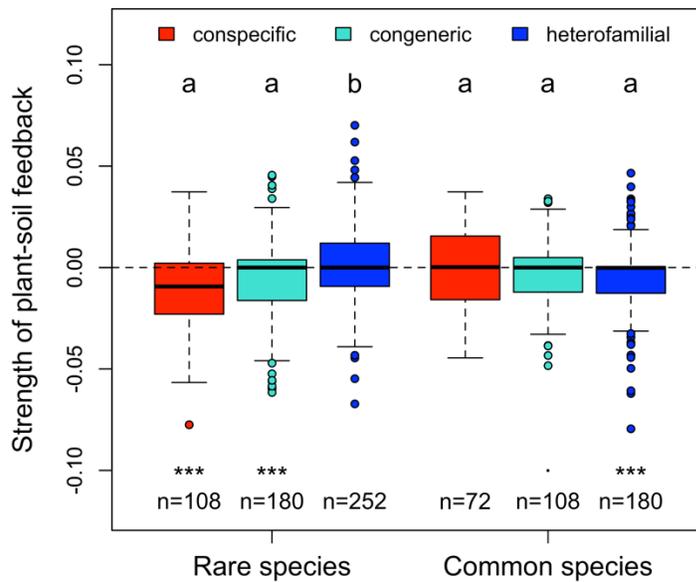
509 **heterofamilials between the rare and common sister species pairs in the shade-house experiment.**

510 Bars indicate \pm one standard error, and asterisks indicate whether the mean in strength is significantly

511 negative ($\cdot P < 0.1$; $* P < 0.05$; $** P < 0.01$; $*** P < 0.001$). The sample size for each bar is given by n .

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516 **Figure 4** Boxplots showing the strength of plant-soil feedback in relation to phylogenetic distance

517 between focal seedling species and soil-source species for rare and common species, in the *in situ*

518 field experiment. Rare species: *Ormosia fordiana*; *Quercus macrocalyx*; *Q. pachyloma*; Common

519 species: *O. glaberrima*; *Q. auricoma*. Letter labels indicate the significant difference in plant-soil

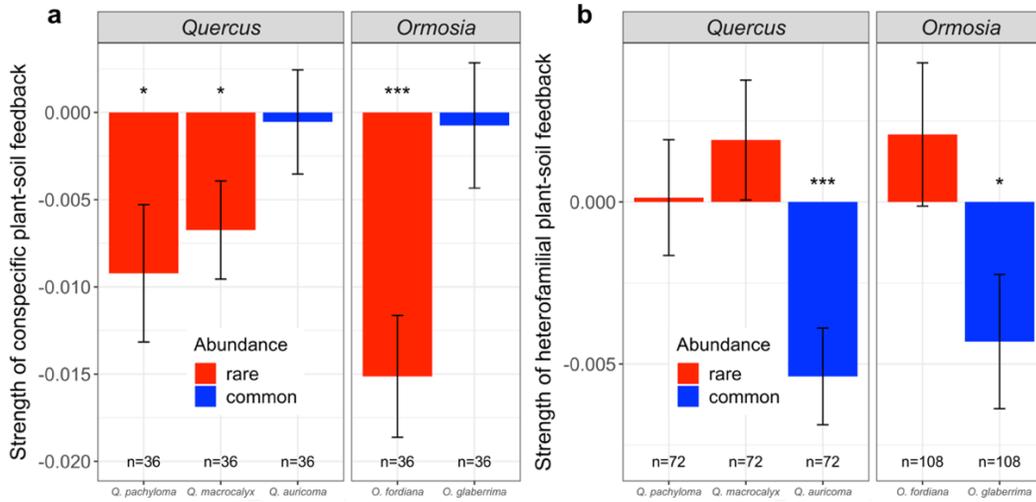
520 feedback across phylogenetic relatedness at $P < 0.05$ by the Dunnett's Modified Tukey-Kramer Pairwise

521 Multiple Comparison Test. Asterisks indicate whether the strength of plant-soil feedback is significantly

522 negative ($\cdot P < 0.1$; $* P < 0.05$; $** P < 0.01$; $*** P < 0.001$). The sample size for each box is given by n .

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527 **Figure 5 Variation in the strength of conspecific plant-soil feedback (a) and heterofamilial**

528 **feedback (b) between the rare and common sister species pairs in the field experiment. Bars**

529 indicate \pm one standard error, and asterisks indicate significantly negative plant-soil feedback ($* P <$

530 0.05 ; $** P < 0.01$; $*** P < 0.001$). The sample size for each bar is given by n .

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