

**1Humic fractions as indicators of soil organic matter responses to clear-cutting in the
2mountain and lowland conditions of south-western Poland**

3

4Effect of forest removing on soil organic matter in the lowland and mountain conditions

5

6Elzbieta Jamroz¹ and Maria Jerzykiewicz²

7

8¹*Wroclaw University of Environmental and Life Sciences,*

9*Institute of Soil Science and Environmental Protection, ul. Grunwaldzka 53,*

10 *50-357 Wroclaw, Poland*

11² *University of Wroclaw, Faculty of Chemistry, Biomaterials Chemistry Group,*

12*ul. F. Joliot-Curie 14, 50-383 Wroclaw, Poland*

13

14*Corresponding author: Elzbieta Jamroz, e-mail: elzbieta.jamroz@upwr.edu.pl*

15*Phone number +48 71 3205632; Fax: +48 71 3205604*

16

17Abstract

18 Clear-cutting means forest removing (stem only) and is the most common type of
19forest harvesting but undoubtedly has a negative impact on the C budget in soils. This work
20aimed to describe responses of soil organic matter in the forest soils to forest removing under
21temperate climate conditions of lowland and mountain regions in south-western Poland.
22Using advanced instrumental analysis, like EPR, ¹H NMR and FT-IR spectroscopy it has been
23found that clear-cutting, alters C cycling and accelerates decomposition in the forest floor
24leading to loss of humic fractions in the investigated soils. In the mountain forests the more
25labile, low-molecular fulvic fraction decreased as the effect of harvesting practice. The

26transformation of organic matter after clear-cutting resulted in the loss of less humified
27organic matter containing humic substances of less polymerised molecules. Analysis of the
28semiquinone radical structures and concentrations showed a decrease in radical concentration
29observed for HA from mountain clear-cut areas compare to the undisturbed forest. Results
30presented in this paper have proved less aliphatic character of humic acid molecules from the
31lowlands, compared to the mountain forest as the effect of clear-cutting. Harvesting practices
32in mountain regions should be approached with particular care due to the risk of erosion of
33exposed surfaces and soils containing less humified and less stable organic matter than in the
34lowlands. Humic fractions of higher solubility, less stability and tendency to migrate through
35the soil profile may favour the leaching of nutrients and consequently cause the eutrophication
36of waters.

37

38**Key words:** humic fraction, soil organic matter, humic acids, EPR, ¹H NMR spectroscopy,
39forest ecosystems

40

411. Introduction

42 Soils are a huge source of terrestrial carbon, storing several times more C than there is
43in the atmosphere (Mayer et al., 2020). Particularly important in this matter are forest soils,
44due to their storing over 40% of the global total organic carbon (Cerli et al., 2008; Falsone et
45al., 2012; Mayer et al., 2020). Most of the soil organic carbon, about 70%, occurs as humic
46substances (De Nobili et al., 2020; Loffredo and Senesi, 2006), whose properties can vary
47depending on not only environmental, natural factors such as soil properties, plant species,
48climate but also anthropogenic factors, such as type of management. Humic fractions, both
49humic and fulvic acids, have been used to study the properties and function of soil organic
50matter (SOM) for hundreds of years (Abbt-Braun and Frimmel, 2002; Olk et al., 2019).

51 According to many authors, changes in forest cover can alter microbial community and
52 accelerate the degree of organic matter decomposition, leading to C losses (Ishikawa et al.,
53 2007; Jamroz et al., 2014; Lan et al., 2020; Prescott, 2005; Ussiri and Johnson, 2007). Clear-
54 cutting, forest removing (stem only), is the most common type of forest harvesting worldwide
55 but undoubtedly has a negative impact on the C budget in soils (Mayer et al. 2020).
56 Understanding the effect of practices like clear-cutting on the soil environment is critical for
57 the postharvest management of sites.

58 Forests in Poland occur mainly in areas with the weakest soils; they are a significant
59 part of the geographic space, 30.4% of the country's area, and they are mainly publicly owned
60 (Jamroz 2014). Mountain habitats occupy 8.7% of the forest area. In 2018 the clear-cutting
61 constituted about 23% of the total timber production. The total area of clear-cutting in Poland
62 was 40.6 thousand ha. The increase in this area in the last two years was mainly due to the
63 necessity from hurricane (2017) liquidation (FinP 2019).

64 Forest harvesting, especially practices like clear-cutting, is described in the literature
65 for both deciduous and coniferous forests (Bergholm et al., 2015; Falsone et al., 2012;
66 Fuzukawa et al., 2006; Ishikawa et al., 2007; Jamroz et al., 2014; Jussy et al., 2004; Piirainen
67 et al., 2002; Ussiri and Johnson 2007). The articles, however, mainly characterise the effect of
68 clear-cutting on nutrient releases, nitrogen and total organic carbon fluxes. There is in the
69 literature still a lack of information describing the effect of clear-cutting on the soil organic
70 matter properties and direction of soil organic matter transformation.

71 Clear-cutting is removing all tree stems in a stand. It causes a strong disturbance of the
72 forest ecosystem by disruption of the biogeochemical cycle of elements (Mayer et al., 2020;
73 Jussy et al., 2004). The disadvantages of clear-cutting for the environment are, among others,
74 unfavourable growth conditions for species requiring shelter when they are young, the threat
75 of wind and water erosion to the surface, especially in the mountain areas, the risk of

76secondary swamping in wetlands, drying of the topsoil layers and a strong expansion of
77herbaceous vegetation (Borelli et al., 2017; Ussiri and Johnson 2007). This type of practice
78induces wide biogeochemical changes from the change in soil moisture and temperature,
79which affects the microbiological activity through rapid accelerated organic matter
80decomposition, leading to high C losses and the reduction of the organic horizon thickness
81although the results from the literature are not consistent across forest sites (Prescott et al.,
822000; Smolander et al., 2019; Valenzuela and Cervantes, 2021). Efforts should be made to
83long-term protection of soil carbon from losses. The C stabilisation in soils requires not only
84practices that can lead to the diminishing of organic matter decomposition but equally
85importantly enhancing the transformation of organic matter into more stable humic
86substances, which can form complexes with a lower turnover rate (Barancikova et al., 2018;
87De Nobili et al., 2020; Jamroz 2012; Jandl et al., 2007; Ojeda et al., 2015; Weber et al., 2018).

88 Decomposition of litter is a process that is very important for determining the
89sustainability of managed forest ecosystems (Rocha et al., 2016). Forest litter consists mainly
90of a mixture of polysaccharides, lignin, aliphatic biopolymers and tannins (Kögel-Knabner,
912002). Humic fractions, as a significant part of soil organic matter, are universally recognised
92as the most reactive soil components maintaining the soil fertility and productivity status and
93are involved in most physical, chemical and biological processes within the soil environment
94(Jerzykiewicz et al., 2018; Senesi et al., 2003, Stevenson 1994, Weber et al., 2018).

95 Ussiri and Johnson (2007) found that accelerated organic matter decomposition after
96clear-cutting resulted firstly in the decomposition of O-alkyl C compounds and the decrease in
97the humic substances in the soil. In some experiments in mountain conditions, it has been
98found that complete removal of trees causes increased humic acid internal oxidation states,
99and the increase of aromatic structures and carboxylic compounds share in the molecules as

100well (Jamroz 2009). The processes of organic matter transformation in the area after clear-
101cutting also reduces the share of aliphatic compounds in humic acid structures.

102 Using advanced instrumental analysis, like EPR, ¹H NMR and FTIR spectroscopy, we
103can describe not only the detailed chemical properties of humic substances and follow the
104direction of humification processes but also predict a change in the impact of organic matter
105after serious disturbance in the plant cover on the soil environment and the entire ecosystem.
106Thus, knowing how to apply modifications to the species composition of new plantings on the
107post-harvested areas to improve the quality of the organic matter will be much easier.

108 This work aimed to describe the direction of humification processes and properties of
109humic substances in the forest soils after clear-cutting in the lowland and mountain regions in
110south-western Poland.

111 The hypotheses were that (1) the direction of soil organic matter transformation after
112clear-cutting does not differ in the lowland and mountain soils and (2) the organic matter from
113the soils after clear-cutting, both in the lowland and mountain regions, transform into humic
114substances of simple molecules with low-molecular-weight and more aliphatic structure.

115

1162. Materials and methods

1172.1. Site description

118 The objects of the investigations were areas two years after clear-cutting (CC), both in
119the lowland and in the mountain region. The type of humus was mor.

1202.1.1. Lowland study site

121 The study area was located in the Oborniki Slaskie Forest District (51° 17.98' N, 19°
12249.059' E), south-western part of Poland. The main site parameters are presented in Table 1.
123Soils from the study area are Brunic Arenosols derived from sand (FAO, 2015). The forest

124habitat before forest harvesting was a mature mixed coniferous forest with dominant mature
125trees of *Pinus silvestris*, *Larix decidua*, *Quercus petraea* and *Acer pseudoplatanus*. At the
126same time, investigations were carried out on the sites after clear-cutting (LF CC) and the
127sites where trees were not cut (LF F). Types of forest habitat soils were the same.

128

1292.1.2. Mountain study site

130 The mountain study area (50° 14.669' N, 16° 50.071'E) was located at an altitude of
131970 m a.s.l. in the Zmijowiec Range, East Sudety Mountains, south-west Poland. The main
132characteristics are presented in Table 1. Soils were described as Dystric Cambisols (FAO,
1332015). The forest habitat was mountain mixed coniferous forest with dominant mature trees:
134*Picea abies*, *Abies alba*, *Acer pseudoplatanus* and *Fagus sylvatica*. The study area was
135located on the site after clear-cutting (MF CC) and at the same altitude under forest cover
136without any harvesting practice (MF F).

137

1382.2. Soil sampling

139 Two replicated soil profiles were located on each site, both after clear-cutting and
140without any harvesting practice, on the same soil type in the lowland – Brunic Arenosols and
141in the mountain – Dystric Cambisols. Two samples from each profile and each horizon were
142prepared for chemical analysis. The texture was determined only in mineral horizons. Soil
143samples from Oa and AE soil horizons in the lowland and Oa and AB soil horizons in the
144mountain area were taken for detailed structural analysis of humic acids.

145 Mineral horizons of lowland soils had a sandy texture, with a very low content (0–4%)
146of particle sizes > 2 mm and a very low content of clay (3–4%). Soils from the mountain area
147had a sandy texture as well but with a much higher (15–23%) content of particles > 2 mm and

148a higher content of silt (27–34%) and a low content of clay (5–7%). Very low pH values
149throughout the whole profile were found for soils investigated in both regions (Table 1).

150 Humic substances were extracted from organic and mineral horizons and detailed
151analyses were performed on humic acids from Oa and AE (AB) horizons.

152

1532.3. *Extraction and purification of humic and fulvic acids*

154 Humic fractions were extracted from the genetic horizons using the procedure
155described by Swift (1996), recommended by the International Humic Substances Society.
156Humic acids (HA) gel was purified with a 0.1 M HCl/0.3 M HF solution, left overnight and
157centrifuged; this procedure was repeated three times. The precipitate was then transferred to a
158Visking dialysis tube (Spectra/Por 7 MWCO 10,000, Spectrum Europe B.V., Breda, The
159Netherlands) and dialyzed against distilled water until a Cl⁻ test was negative. Then, the HA
160were freeze-dried.

161 Fulvic acid (FA) extracts were passed through the XAD-8 column and H⁺-saturated
162cation exchange resin. The eluate was freeze-dried.

163 Elemental analysis of HA and FA was performed with a Perkin-Elmer 2000
164instrument. The O was calculated from the mass balance.

165

1662.4. *Spectroscopic analysis*

1672.4.1 *EPR*

168 Semiquinone radicals, which are stabilised in humic substances, are highly sensitive to
169various factors, both physical (temperature and humidity) and chemical (redox and/or acid-
170base reactions, etc.) (Senesi and Schnitzer, 1977). The concentration of radicals and their g-
171parameters (related to the magnetic moment of the unpaired electron of radicals) depends on

172the type and origin of the substances in which these radicals are formed (Swift 1996; Senesi
173and Loffredo, 2001). Thus, the studies of the structure and concentration of these radicals
174could be indicative of the whole structure of the organic fraction (Jerzykiewicz et al., 2019).
175The electron paramagnetic resonance EPR method, as the best-known method for studies of
176paramagnetic substances, is a good choice for using for the investigation of structural changes
177occurring in radicals within soil organic matter. EPR analysis is known as an important
178method for determining humic substance characteristics of various origins (Senesi 1990;
179Jerzykiewicz et al., 1999).

180 All EPR spectra of humic acids were recorded at room temperature using a Bruker
181Elexsys E500 spectrometer equipped with an NMR Teslameter (ER 036TM) and standard
182Bruker frequency counter. X-band spectra were measured using a double rectangular cavity
183resonator ER 4105DR operating in the TD104 mode at a microwave power of 20 mW and
184modulation amplitude of 1G. For quantitative measurements, the radical species concentration
185standards were placed in the double resonator's first cavity, while the analysed sample was
186placed in its second cavity. After tuning, the spectra were recorded separately for each of the
187two cavities without changing any of the measurement parameters. As the references of
188radical concentration, the standards distributed by IHSS (peat and Leonardite HA; 2.0×10^{17}
189and 4.4×10^{17} spins per gram, respectively) and Bruker (alanine pill, 1.7×10^{17} spins per pill)
190were used. Double integrations were conducted using the *WinEPR 2.22 rev.12* programs
191(developed by Bruker).

192

1932.4.2 ¹H – NMR

194 ¹³C-NMR and/or ¹H-NMR spectroscopy is one of the most useful methods for the
195characterisation of soil organic matter (Schnitzer 1994; Kögel-Knabner 1997; Conte et al.,
1962004). This spectroscopy is commonly used in the study of humic substances to describe the

197concentration of functional groups (Jamroz et al., 2014). ¹H-NMR measurements were
198performed on a Bruker Avance III at 500 MHz. Samples of humic acids were dissolved in a
199solution of NaOH in D₂O and transferred to standard glass NMR tubes.

200

2012.4.3 FT-IR

202 Fourier transformed infrared (FTIR) spectroscopy provides information about nature
203and functional groups as well as structural information indicated the presence of both aliphatic
204and aromatic components. FTIR spectroscopy gives us also information about what
205degradation products are derived from, e.g. proteins and polysaccharides (Senesi et al., 1991;
206Traversa et al., 2008). Fourier transformed infrared spectra of humic and fulvic acids were
207recorded with a Bruker Vertex 70 FT-IR spectrometer on KBr pellets (an approximately 1 mg
208sample in 400 mg of KBr). The spectra were integrated into the oscillations ranges of the
209characteristic group.

210

2112.5 ICP-OES

212 Humic and fulvic acid digestion was carried out on the reactor's Ertec Magnum and
213using HNO₃. The thus prepared samples were examined for the study using an ARL Model
2143410 ICP (Fisons Instruments) spectrometer to determine the metal ion contents: Fe(III),
215Mn(II) and Cu(II).

216

2172.5. TOC and Nt analysis

218 The content of total organic carbon was analysed using a CS-mat 5500 instrument
219(Strohlein GmbH & Co., Kaarst, Germany, currently Bruker AXS Inc., Madison, WI, USA).
220Total nitrogen was analysed by the Kjeldahl method using a Buchi [Labortechnik GmbH](#) N
221analysis.

2222.6. *Total acidity of humic acids*

223 The total acidity of humic acids was analysed with Schnitzer and Gupta's method as
224described by Swift (1996).

2252.7. *Statistical analysis*

226 Results were verified using Statistica for Windows 13. Means were compared by the
227t-test, at a confidence level of $p < 0.05$

228

2293. **Results**

2303.1. *TOC and humic fractions content*

231 In our studies content of TOC in the Oa horizon decreased by 26% in the lowland and
232by 34% in the mountain soils, but the quantitative change was more evident in mineral
233horizons (Table 1). In the A horizons, TOC decreased by 22% in the lowlands and by 45% in
234the mountain areas. Surprisingly conversely in B horizons, soil organic carbon decreased by
23567% in the lowland and only by 8% in the mountain soils. That is why in mountain soils, even
236in natural sites and despite the soil formation process, there is a much higher content of TOC
237in B horizons (48.50 g kg^{-1}) in comparison to the lowland ones (6.17 g kg^{-1}). Content of total
238nitrogen was lower in the clear-cut sites nearly by 31% in organic horizons both in the
239lowlands and in the mountain areas (Table 1). In mineral soil horizons, the content of N total
240was adequate to the TOC concentration. The concentration of humic fraction decreased for
241both humic and fulvic acids after clear-cutting in whole soil horizons (Table 1). Humic acids
242in Oa horizons decreased after clear-cutting similarly in the lowland and the mountain soils,
243by nearly 18% in both regions. In the A horizons, this change was more evident in the lowland
244(by 32%) in comparison to the mountain soils (change by 8%). In the case of fulvic acids,
245their content significantly decreased more evidently in the mountain soils, by 30% in Oa and

246by 54% in A horizon, in comparison to the lowland, where they decreased by 8.5% and 24%
247respectively. In the mineral horizons of the soil on the clear-cut site, however, there was found
248a lower content of soil organic carbon in comparison to the undisturbed site. In our study,
249clear-cutting had an evident consequence in altering the quantity of soil organic matter but
250differed between the lowlands and mountain areas. Meanwhile, in the mountain soils, there
251was a more clear decrease of the fulvic fraction in the whole soil profile. In the lowlands in
252the upper soil horizons (Oa, A), it was found that the content of humic fraction content
253decreased more in the clear-cut site in comparison to the undisturbed areas (Table 1).

2543.2. *Elemental composition and UV-VIS results of humic and fulvic acids*

255 Elemental composition is one of the most important humic fraction properties. In the
256investigated soils, FA molecules contained more oxygen than HA molecules, regardless of
257management practice or altitude, but the content of carbon was varied (Table 2). Molecules of
258humic acids from the Oa horizons of the clear-cut sites, in the lowlands, were characterised by
259a higher content (although not statistically confirmed) of carbon and oxygen and lower
260content of hydrogen in comparison to the undisturbed sites. In the first mineral horizons, a
261significant decrease in the carbon content of the humic acids molecules was found in the
262lowland (from 35.39% to 35.01%) and there was a significant increase in the mountain sites
263under the influence of clear-cutting practice (from 30.54% to 32.22%). The content of
264nitrogen increased in the molecules of HA in the lowland after CC, but the change was not
265significantly confirmed in the first mineral soil horizon (from 1.94% to 2.04%). There were
266no significant differences in N content between clear cut and undisturbed soils in the
267mountain areas (Table 2). The FA from the Oa horizons, both in the lowlands and mountain
268region, did not significantly differ between clear-cut sites and those without any harvesting
269practice. On the contrary, in the A horizons of the lowland's soils, the FA molecules contained
270more carbon and significantly less oxygen than those from the undisturbed forest. In the

271investigated mountain soils from the clear-cut sites, fulvic acids contained more carbon
272(33.91%) as well and significantly less hydrogen (35.17%) compared to those from the
273undisturbed forest sites (32.95% and 36.04%, respectively). In the studied areas, the H/C ratio
274of the HA was rather high and ranged from 1.46 to 1.61 in the Oa horizons and from 1.18 to
2751.45 in the first mineral horizons (Table 2). The E4/E6 ratio of humic acids from Oa and
276analysed mineral horizons of the soils on the clear cut sites, both in the lowlands and in the
277mountain investigated region (Table 2), showed lower values (4.18 and 3.80 in the lowlands
278and 4.82 and 4.31 in the mountain) in comparison to the undisturbed forest sites (6.16 and
2794.33 in the lowlands and 6.32 and 4.89 in the mountain regions, respectively).

2803.3. EPR results

281 The studies of radical structure and concentration followed the analysis of the
282manganese, iron and copper contents in humic acids because these metal ions are known from
283their antiferromagnetic interaction with semiquinone radicals in humic acid (Jerzykiewicz et
284al., 2002). As a result, a decrease in radical concentration is observed. For the investigated
285samples, the content of manganese ions was very small (0–0.023 mg g⁻¹), and copper ions
286were not detected. Thus, these metal ions did not affect semiquinones.

287 Differently, iron ion contents were much higher and varied between samples. In humic
288acids extracted from mineral horizons, the iron concentration was higher than that of those
289that were extracted from organic ones (Table 3). It is worth mentioning that samples from the
290lowland forest areas did not exhibit the influence of clear-cutting on the metal ion
291concentrations. The increase of iron ion concentrations in HA was observed after clear-cutting
292only for samples from MF areas. Similar phenomena as described previously of radical
293quenching by metal ions were found only for the mountain soils, as presented in Table 3.

294 In the A horizons of the lowland soils, iron(III) ions did not affect radical
295concentration because metal ions are incorporated there by different functional groups unlike

296in Oa horizons. That is well presented on the EPR spectra (Fig. 1), where different lines
297attributed to different Fe(II) and Fe(III) were observed at values of g equal to 2.0, 4.25 and 6.
298This third peak (Fig. 1, MF-F AB) is especially interesting because iron is built-in here via
299nitrogen atoms, not oxygen as it is for other iron(III)-humic acid bonds. It is worth
300mentioning that elemental analysis confirms that HA characterised by a $g = 6$ peak contained
301also more nitrogen. The signal at $g = 6.0$ is commonly admitted to pertain to the trivalent
302heme-iron-like complex in which iron is at the third degree of oxidation instead of the second
303one (Krzyminiewski et al., 2011).

304 As it is presented in Table 3, g – the parameter calculated from EPR spectra for
305semiquinone radicals did not change much. Better information could be obtained from studies
306of radical concentration. The high radical concentration is commonly known as an indicator
307of high humification degree (Jerzykiewicz et al., 1999). In the presented case, a decrease in
308radical concentration is observed for HA from mountain clear-cut areas (Table 3). The
309decrease in radical concentration could be the effect of higher Fe(III) content and the
310antiferromagnetic interaction (Jeziński et al., 2002). For samples with higher Fe(III) content
311(4.796 in Oa and 5.356 mg g⁻¹ in the A horizons after clear-cutting compare to 1.562 and
3123.027 from the undisturbed forest respectively), radical content is much lower for HA from
313the clear-cutting area than from undisturbed forest.

3143.4. FTIR spectra of humic acids

315 The FT-IR spectra were typical for humic acid spectra (Figure 2). The differences,
316although small, were observed only in the signal intensities. First, lines of COOH vibrations
317at 1722 cm⁻¹ were less intense for HA samples from mineral horizons. The difference is even
318more distinguished after clear-cutting. Peaks ascribed as aliphatic C–H stretching vibrations
319(3000–2820 cm⁻¹) and anti-symmetric stretching modes of COO⁻ and/or aromatic C=C (1650
320cm⁻¹) for humic acids extracted from clear-cutting areas were characterised by higher

321intensities and were more clearly marked for HA from the soil horizons in the lowlands than
322for HA from undisturbed forest soils. The absorption band may be associated with aromatic
323C=C, C=O groups of quinones and ketones (Traversa et al., 2008).

3243.5 *¹H NMR analysis of humic acids*

325 The NMR spectra presented typical NOM spectra (Figs. 3 and 4). This information is
326valuable because of the quantification reliability (Grinhut et al., 2011; Hertkorn et al., 2007).

327 The spectra were divided into three regions. The first was with the chemical shift
328region of 0–2.33 ppm was attributed to CH₂ groups. An increase in HA from both horizons
329has been observed after clear-cutting in the lowlands and mountain regions as well (Figs. 3
330and 4). This may be linked to HA degradation after environmental disturbances such as clear-
331cutting. Demethylation during this process causes the removal of CH₂ groups, and thus the
332detection of more molecules that belonged to the consecutive CH₂ series (Grinhut et al.,
3332011). Within the chemical shift region of 2.93–4.26 (connecting with methoxy groups), there
334has been observed a decrease after clear-cutting, indicating a decrease of oxygen-containing
335functional groups (Grinhut et al., 2011). These findings are in good agreement with a decrease
336of the O/C ratio in the mineral mountain soil horizons after clear-cutting. The third region of
3375.8–8.2 ppm was assigned to aromatic protons and was able to be analysed after integral
338calculation.

339 The integration of the lines attributed to the previously mentioned three areas
340exhibited more detailed differences between samples. About 8.58% of the non-exchangeable
341protons of the HA from the undisturbed lowland forest soil (Oa horizon) were assigned to
342aromatic groups (Table 4), whereas after clear-cutting, this amount increased over three times
343(26.97%). A slighter increase in aromaticity was also detected after clear-cutting in HA
344molecules from mountain regions compared to the lowlands, but only in Oa horizons.

345 The ^1H NMR spectra exhibit not only a slight increase of aliphatic proton shifts (0–1
346ppm) but also a slight increase in aromatic proton shifts (5.8–8.2 ppm). Increase these two
347bands could be an explanation why the summary aromatic-aliphatic parameter from elemental
348analysis remains unchanged or only slightly decreases. The observed increase in the intensity
349of aliphatic and aromatic shifts is accompanied by the decrease of intensity of methoxyl shifts.
350

3514. Discussion

352 In our studies, clear-cutting enhanced transformation of soil organic matter, through an
353increase of organic matter decomposition and humification process as well. In mountain soils,
354even in natural sites and despite the soil formation process, there is a much higher content of
355TOC in mineral horizons in comparison to the lowland ones. This may be because, in the
356mountain regions, there was much higher precipitation than in the lowland, causing
357naturally higher leaching of elements, especially low molecular fractions of carbon, to the
358deeper soil horizons. According to many authors (Achat et al., 2015; Jamroz, 2009; Mayer et.
359al., 2020; Ussiri and Johnson, 2007), clear-cutting in first years enhances the decomposition
360rate of organic matter and consequently reduces the amount of soil organic matter in upper
361soil horizons. On the other hand, Falsone et al. (2012) observed that 5 years after clear-
362cutting, the content of soil organic carbon in the Oa horizon was a bit higher (186 g kg⁻¹) in
363comparison to the soil from the undisturbed area (173.7 g kg⁻¹). Thus, we can expect that
364content of SOM will recover over time but the rate will depend on type of afforestation. The
365origin of humus fractions in oligotrophic forest ecosystems is mainly plant residues rich in
366polysaccharides, lignin, aliphatic biopolymers as well as tannins (Kogel-Knabner 2002,
367Ziolkowska et al., 2020), which more often transform in such conditions into low-molecular
368humic substances, like fulvic acids, than into compounds of higher polymerisation, like humic
369acids. Coniferous and mixed coniferous forest ecosystems are characterised by the type of soil

370organic matter with the prevalence of the fulvic fraction over the humic one and in most cases
371with the mor type of humus. Fulvic fractions, apart from their low molecular weight, are
372characterised by a more aliphatic structure that favours leaching through the soil profile
373(Buurman and Jongmans, 2005; Falsone et al., 2012; Jamroz et al., 2014). Low pH and
374changes in microclimate conditions, particularly in the mountain areas with much higher
375precipitation than in the lowlands, means that removing trees alters microbial activity, and the
376weakly decomposed low-molecular fraction can be easily transported to the lower part of the
377soil profile or even leached to the groundwater. The H/C ratio of HA molecules is often used
378as an indicator of aromaticity/aliphaticity and is useful for monitoring structural changes
379(Senesi and Loffredo, 1999; Rice and MacCarthy, 1991). Results of our studies point to the
380overall more aliphatic than the aromatic character of the molecules' structures. Falsone et al.,
381(2012) found that a larger proportion of the aliphatic component could be related to a greater
382contribution of saccharide residues. Humic acids from the A horizons (in the mountain soils)
383and the Oa horizons (in the lowlands and mountain areas as well) after clear-cutting were less
384aliphatic (lower H/C ratio) than those from undisturbed forest sites. The lower H/C ratio,
385lower total acidity, higher content of carbon and oxygen in HA molecules from Oa horizons
386after clear-cutting shows that the harvesting practice altered not only the decomposition but
387also the humification process. The transformation of organic matter in the investigated sites
388after clear-cutting, particularly in Oa horizons both in the lowlands and mountain areas,
389resulted in more humified products with less aliphatic structures. The higher O/C ratio in
390fulvic acids than in humic acids in all investigated forest sites confirms the statement that FA
391are polysaccharidic in nature (Tan, 2014). This parameter significantly decreased only in the
392lowland in the first mineral soil horizon after clear-cutting. During the transformation of
393organic matter after clear-cutting, more of the aromatic substances are broken down and
394released in comparison with the polysaccharide constituents. Consequently, the humic

395 substances formed became increasingly more polysaccharidic in nature. The molecular weight
396 of humic substances is related to the E4/E6 ratio, and this ratio is considered by many authors
397 as a measure of the overall humification degree (Barancikova et al., 2018; Chen et al., 1977;
398 Zaccone et al., 2013). Results received indicate the formation of similar size molecular
399 weights of humic substances before and after clear-cutting in the coniferous mixed forest in
400 the lowlands and mountain regions as well. Barancikova et al. (2018) point out that humic
401 substances with higher molecular weights are more stable in the environment and more
402 resistant to unfavourable external factors. Analysis of the semiquinone radical structures and
403 concentrations can be useful in studies of the humification process (Klavins and Purmalis
404 2014). This type of radicals can be built into soil organic matter structures (Senesi 1990).
405 Humic substance molecules are rich in functional groups and can form, among others, metal
406 chelate complexes (Barancikova et al., 2018; Jerzykiewicz et al., 2019). EPR results indicate
407 the direct influence of the harvesting procedure on radical concentration. However, it is
408 difficult to state unambiguously what is the mechanism of the clear-cutting procedure that
409 influences the enhancement of the interaction of metal ions with unpaired electrons of HA
410 radicals. Similar results were described by Barancikova et al. (2018), who found a decrease in
411 radical content in humic acids from the forest soils after windstorms and wildfires. The free
412 radical concentration is overall in line with aromaticity, humification degree and other
413 chemical and structural properties of humic acids (Senesi et al., 2003). Other results presented
414 in this paper (H/C; O/C, E4/E6) proved the less aliphatic character of humic acid molecules
415 from the lowlands, compared to the mountain forest, as the effect of clear-cutting. Results of
416 the FTIR analysis, especially in the lowlands after clear-cutting, may reflect a higher degree
417 of organic matter decomposition (Haberhauer et al., 1998) and point to different from the
418 spruce-mountains type of litter. Spruce needles and bilberry leaves, which are often part of the
419 litter composition from mountain forest soils, contain a high amount of polyphenols, which

420slow down the decomposition processes (Albers et al., 2004, Jamroz et al., 2014) in
421comparison to pine litter in the lowlands. These results are in good agreement with those from
422elemental analysis, confirming the less aliphatic nature of humic acids from the investigated
423forest soils in the lowlands. All ¹H NMR spectra examined indicate that aliphatic structures
424are dominant components of humic acids especially in mineral soil horizons, whose contents
425even increased after clear-cutting particularly in the lowland. These results are in line with
426other authors (Bonifacio et al., 2006; Jamroz et al., 2014), who reported a higher degree of
427aliphaticity of SOM in mineral forest soil horizons.

428

4295. Concluding remarks

430 Soil organic matter in the coniferous forest ecosystems is characterised mostly by
431more aliphatic character, both in the lowlands and mountain conditions, and lower
432humification degree in comparison to other ecosystems (cropland or grassland). Forest
433management, especially clear-cutting, alters C cycling and accelerates decomposition in the
434forest floor leading to loss of humic substances with the highest C losses in the forest floor
435and the upper mineral soil. In our studies, clear-cutting enhanced the transformation of soil
436organic matter, through the increase of organic matter decomposition. Humic fraction
437decreased under the influence of clear-cutting more in the lowland areas covered by mainly
438pine forests. In the mountain forests, with Norway spruce as the main species, the more labile,
439low-molecular fulvic fraction decreased more noticeably as the effect of harvesting practice.
440Presumably, the main cause of these changes was the activation of easily soluble fractions and
441their leaching from the soil profile. Lower H/C ratio, higher content of carbon and oxygen in
442HA molecules from Oa horizons after clear-cutting shows that harvesting practice altered not
443only the decomposition but also the humification process as well. The transformation of
444organic matter in the investigated sites after clear-cutting, particularly in Oa horizons both in

445the lowlands and mountain areas, resulted in the loss of less humified organic matter
446containing humic substances of less polymerised molecules. Results presented in this paper
447have proved less aliphatic character of humic acid molecules from the lowlands, compared to
448the mountain forest as the effect of clear-cutting. Harvesting practices in mountain regions
449should be approached with particular care due to the risk of erosion of exposed surfaces and
450soils containing less humified and less stable organic matter than in the lowlands. Humic
451fractions of higher solubility, less stability and tendency to migrate through the soil profile
452may favour the leaching of nutrients and consequently cause the eutrophication of waters.
453These phenomena due to climate change are predicted to even increase, leading to the
454“brownification” of surface waters (Lurling et al., 2014).

455 Based on the literature and presented results, it could be stated that the study of humic
456fractions can help resolve scientific and practical issues in ecosystems. Practices through
457improved growth of tree seedlings, applying organic materials into deeper soil horizons and
458selection of species with nitrogen-fixing associates will prevent the reduction of C stocks, soil
459surface in the mountain regions against erosion processes as well as ensure the stability of the
460whole ecosystem.

461

462**Acknowledgments**

463This work was financed by a National Science Center, (grant no. N N305 155937) and
464statutory activity subsidy from the Polish Ministry of Science and Higher Education for the
465Faculty of Chemistry of Wrocław University.

466

467**Conflict of Interest Statement**

468The author declares that there is no conflict of interest that could be perceived as prejudicing
469the impartiality of the research reported.

471 **References**

- 472 Abbt-Braun G. and Frimmel F. H. (2002). The relevance of reference materials isolation and
473 general characterization, in: *Refractory Organic Substances in the Environment* F. H.
474 Frimmel, G. Abbt-Braun, K. G. Heumann, B. Hock, H.-D. Lüdemann, and M. Spiteller
475 (eds.) WILEY-VCH Verlag GmbH & Co. KGaA, Weinheim, 1-38,
476 <https://doi.org/10.1002/9783527611195.ch2q>
- 477 Achat D.L., Fortin M., Landmann G., Ringeval B., Augusto L. (2015). Forest soil carbon is
478 threatened by intensive biomass harvesting *Sci. Rep.* 5, 15991, <https://doi.org/10.1038/srep15991>
479 [srep15991](https://doi.org/10.1038/srep15991)
- 480 Albers, D., Migge, S., Schaefer, M., Scheu, S. (2004). Decomposition of beech leaves (*Fagus*
481 *sylvatica*) and spruce needles (*Picea abies*) in pure and mixed stands of beech and
482 spruce. *Soil Biol. Biochem.* 36, 155–164 <https://doi.org/10.1016/j.soilbio.2003.09.002>
- 483 Barancikova G., Jerzykiewicz M., Gomoryova E., Tobiasova E., Litavec T. (2018). Changes
484 in forest soil organic matter quality affected by windstorm and wildfire *J Soils*
485 *Sediments* 18, 2738–2747 <https://doi.org/10.1007/s11368-018-1942-2>
- 486 Bergholm J., Olsson B.A., Vegerfors B., Persson T. (2015). Nitrogen fluxes after clear-cutting.
487 Ground vegetation uptake and stump/root immobilisation reduce N leaching after
488 experimental liming, acidification and N fertilisation *For. Ecol. Manag.* 342, 64-75,
489 <https://doi.org/10.1016/j.foreco.2015.01.009>
- 490 Bonifacio, E., Santoni, S., Celi, L., Zanini, E. (2006). Spodosol-Histosol evolution in the
491 Krkonose National Park (CZ). *Geoderma* 131, 237–250
492 <https://doi.org/10.1016/j.geoderma.2005.03.023>
- 493 Borelli P., Panagos P., Märker M., Modugno S., Schütt B. (2017). Assessment of the impacts
494 of clear-cutting on soil loss by water erosion in Italian forests: First comprehensive

495 monitoring and modelling approach *Catena*, 149 (3), 770-781,
496 <https://doi.org/10.1016/j.catena.2016.02.017>

497Buurman, P., Jongmans, A.G. (2005). Podzolisation and soil organic matter dynamics
498 *Geoderma* 125, 71–83 <https://doi.org/10.1016/j.geoderma.2004.07.006>

499Cerli C., Celi L., Kaiser K., Guggenberger G., Johansson M.-B., Cignetti A., Zanini E. (2008).
500 Changes in humic substances along an age sequence of spruce stands planted on former
501 agricultural land *Organic Geochemistry* 39, 1269-1280
502 <https://doi.org/10.1016/j.orggeochem.2008.06.001>

503Chen, Y., N. Senesi, and M. Schnitzer. (1977). Information provided on humic sub-
504 stances by E₄/E₆ ratios. *Soil Sci. Soc. Am. J.* 41, 352–358
505 <https://doi.org/10.2136/sssaj1977.03615995004100020037x>

506 Conte P., Spaccini R., Piccolo A. (2004). State of the art of CPMAS ¹³C-NMR spectroscopy
507 applied to natural organic matter. *Prog. Nucl. Mag. Res. Sp.* 44, 215–223
508 <https://doi.org/10.1002/chin.200443280>

509De Nobili M., Bravo C., Chen Y. (2020). The spontaneous secondary synthesis of soil organic
510 matter components: A critical examination of the soil continuum model theory *Applied*
511 *Soil Ecology*, 154, 103655 <https://doi.org/10.1016/j.apsoil.2020.103655>

512Falsone G., Celi L., Caimi A., Simonov G., Bonifacio E. (2012). The effect of clear cutting on
513 podzolisation and soil carbon dynamics in boreal forests (Middle Taiga zone, Russia)
514 *Geoderma*, 177-178, 27-38 <https://doi.org/10.1016/j.geoderma.2012.01.036>

515FAO (2015). Word Reference Base for Soil Resources. Word Soil Resources Report No. 106,
516 FAO, Rome [http://www.fao.org/soils-portal/data-hub/soil-classification/world-](http://www.fao.org/soils-portal/data-hub/soil-classification/world-reference-base/en/)
517 [reference-base/en/](http://www.fao.org/soils-portal/data-hub/soil-classification/world-reference-base/en/)

518FinP (2019). Forests in Poland in: Milewski W. (ed.). The State Forests Information Centre,
519 Warsaw [http://www.lasy.gov.pl/pl/informacje/publikacje/in-english/forests-in-poland/
520 \[lasy-w-polsce-2017-en.pdf\]\(http://www.lasy.gov.pl/pl/informacje/publikacje/in-english/forests-in-poland/lasy-w-polsce-2017-en.pdf\)](http://www.lasy.gov.pl/pl/informacje/publikacje/in-english/forests-in-poland/lasy-w-polsce-2017-en.pdf)

521Fukuzawa K., Shibata H., Takagi K., Nomura M., Kurima N., Fukazawa T., Satoh F., Sasa K.
522 (2006). Effects of clear-cutting on nitrogen leaching and fine root dynamics in a cool-
523 temperate forested watershed in northern Japan *For. Ecol. Manag.* 225, 257-261
524 <https://doi.org/10.1016/j.foreco.2006.01.001>

525Grinhut T., Hertkorn N., Schnitt-Kopplin P., Hadar Y., Chen Y. (2011). Mechanisms of humic
526 acids degradation by white rot fungi explored using ¹H NMR spectroscopy and
527 FTICR mass spectroscopy *Environ. Sci. Technol.* 45, 2748-2754
528 <https://doi.org/10.1021/es1036139>

529Haberhauer, G., Rafferty, B., Strebl, F., Gerzabek, M.H., (1998). Comparison of the
530 composition of forest soil litter derived from three different sites at various
531 decompositional stages using FTIR spectroscopy *Geoderma* 83, 331–342
532 [https://doi.org/10.1016/S0016-7061\(98\)00008-1](https://doi.org/10.1016/S0016-7061(98)00008-1)

533Hedde M., Aubert M., Decaens T., Bureau F. (2008). Dynamics of soil carbon in a beechwood
534 chronosequence forest *Forest Ecology and Management*, 255, 193-202
535 <https://doi.org/10.1016/j.foreco.2007.09.004>

536Hertkorn N., Ruecker C., Meringer M., Gugisch R., Frommberger M., Perdue E.M., Witt M.,
537 Schnitt-Kopplin P. (2007). High-precision frequency measurements: indispensable
538 tools at the core of the molecular-level analysis of complex systems *Anal Bioanal*
539 *Chem* 389, 1311-1327 <https://doi.org/10.1007/s00216-007-1577-4>

540Ishikawa H., Osono T., Takeda H. (2007). Effects of clear-cutting on decomposition processes
541 in leaf litter and the nitrogen and lignin dynamics in a temperate secondary forest *J.*
542 *For. Res.*, 12, 247-254 <https://doi.org/10.1007/s10310-007-0013-0>

543Jamroz E. (2009). Effect of clear-cutting on the properties of humic compounds in Podzols in
544 the Śnieżnik Massif region (in Polish with English Abstract) *Soil Science Annual LX*
545 (4), 35-41.

546Jamroz E. (2012). [Properties of soil organic matter in the forest soils under mountain dwarf
547 pine in the Śnieżnik Klodzki Reserve *Sylwan*, 156 \(11\), 825-832
548 <https://doi.org/10.26202/sylwan.2011133>](https://doi.org/10.26202/sylwan.2011133)

549Jamroz E., Kocowicz A., Bekier J., Weber J. (2014). Properties of soil organic matter in
550 Podzols under mountain dwarf pine (*Pinus mugo* Turra.) and Norway spruce (*Picea*
551 *abies* (L.) Karst.) in various stages of dieback in the East Sudety Mountains, Poland
552 *For. Ecol. Manag.* 330, 261-270 <https://doi.org/10.1016/j.foreco.2014.07.020>

553Jandl R., Lindner M., Vesterdal L., Bauwens B., Baritz R., Hagedorn F., Johnson D.W.,
554 Minkkinen K., Byrne K.A. (2007). How strongly can forest management influence soil
555 carbon sequestration? *Geoderma*, 137, 253-268,
556 <https://doi.org/10.1016/j.geoderma.2006.09.003>

557Jerzykiewicz M., Drozd J., Jezierski A. (1999). Organic radicals and paramagnetic metal
558 complexes in municipal solid waste composts. An EPR and chemical study
559 *Chemosphere*, 39, 253-268 [https://doi.org/10.1016/S0045-6535\(99\)00107-1](https://doi.org/10.1016/S0045-6535(99)00107-1)

560Jerzykiewicz M., Jezierski A., Czechowski F., Drozd J. (2002). Influence of metal ions
561 binding on free radical concentration in humic acids. A quantitative Electron
562 Paramagnetic Resonance study *Organic Geochemistry* 33, 265 –268
563 [https://doi.org/10.1016/S0146-6380\(01\)00158-9](https://doi.org/10.1016/S0146-6380(01)00158-9)

564Jerzykiewicz M., Barancikova G., Jamroz E., Kałuza-Haladyn A. (2019). Application of EPR
565 Spectroscopy in Studies of Soils from Destroyed Forests *Appl Magn Reson* 50, 753–
566 760 <https://doi.org/10.1007/s00723-018-1055-5>

567Jeziński A., Czechowski F., Jerzykiewicz M., Golonka I., Drozd J., Bylińska E., Chen Y.
568 Seaward M.R.D. (2002). Quantitative EPR study on free radicals in the natural
569 polyphenols interacting with metal ions and other environmental pollutants
570 *Spectrochim. Acta Part A Mol Biomol. Spectrosc.* 58, 1293-1300, DOI: [10.1016/s1386-](https://doi.org/10.1016/s1386-1425(01)00718-1)
571 [1425\(01\)00718-1](https://doi.org/10.1016/s1386-1425(01)00718-1)

572Jussy J.-H., Ranger J., Bienaime S., Dambrine E. (2004). Effects of clear-cut on the situ
573 nitrogen mineralization and the nitrogen cycle in a 67-year-old Douglas-fir
574 (*Pseudotsuga menziesii* (Mirb.) Franco) plantation *Ann.For. Sci.* 61, 397-408,
575 <https://doi.org/10.1051/forest.2004033>

576Klavins, M., Purmalis, O. (2014). Characterization of humic acids from raised bog peat
577 *Latvian Journal of Chemistry*, 52 (1-2), 83–97 DOI: [10.7250/msac.2013.026](https://doi.org/10.7250/msac.2013.026)

578Kögel-Knabner, I. (1997). ¹³C and ¹⁵N NMR spectroscopy as a tool in soil organic matter
579 studies. *Geoderma* 80, 243–270 [https://doi.org/10.1016/S0016-7061\(97\)00055-4](https://doi.org/10.1016/S0016-7061(97)00055-4)

580Kögel-Knabner I. (2002). The macromolecular organic composition of plant and microbial
581 residues as inputs to soil organic matter *Soil Biology & Biochemistry*, 34, 139-162
582 [https://doi.org/10.1016/S0038-0717\(01\)00158-4](https://doi.org/10.1016/S0038-0717(01)00158-4)

583Krzyminiewski R., Kruczynski Z., Dobosz B., Zajac A., Mackiewicz A., Leporowska E.,
584 Folwaczna S. (2011). EPR Study of Iron Ion Complexes in Human Blood *Appl Magn*
585 *Reson* 40, 321–330 <https://doi.org/10.1007/s00723-011-0219-3>

586Lal R. (2005). Forest soils and carbon sequestration *Forest Ecology and Management*, 220,
587 242-258 <https://doi.org/10.1016/j.foreco.2005.08.015>

588Lan G., Wu Z., Yang C., Sun R., Chen B., Zhang X. (2020). Forest conversion alters the
589 structure and functional processes of tropical forest soil microbial communities. *Land*
590 *Degradation and Development*, <https://doi.org/10.1002/ldr.3757>

591 Loffredo E. and Senesi N. (2006). The role of humic substances in the fate of anthropogenic
592 organic pollutants in soil with emphasis on endocrine disruptor compounds in: *Soil and*
593 *Water Pollution Monitoring, Protection and Remediation*, Twardowska I. et al. (eds.)
594 Springer, 3-23 https://doi.org/10.1007/978-1-4020-4728-2_4

595 Lüring M., Waajen G., van Oosterhout F. (2014). Humic substances interfere with phosphate
596 removal by lanthanum modified clay in controlling eutrophication *Water Res.* 54, 78-
597 88, <https://doi.org/10.1016/j.watres.2014.01.059>

598 Mayer M., Prescott C.E., Abakerd W. E.A, Augustoe L., Cécillon L., Ferreirah G.W.D.,
599 Jamesi J., Jandl R., Katzensteiner K., Laclau J.P., Laganièrem J., Nouvellonk Y., Parém
600 D., Stanturf J.A., Vanguelovao E.I., Vesterdalp L. Tamm (2020). Influence of forest
601 management activities on soil organic carbon stocks: A knowledge synthesis. *For. Ecol.*
602 *Manag.*, 466, 118127, <https://doi.org/10.1016/j.foreco.2020.118127>

603 Olk D.C., Bloom P.R., Perdue E.M., McKnight D.M., Chen Y., Farenhorst A., Senesi N., Chin
604 Y.-P., Schmitt-Kopplin P., Hertkorn N., Harir M. (2019). Environmental and
605 agricultural relevance of humic fractions extracted by alkali from soils and natural
606 waters *J. Environ. Qual.* 48, 217-232, <https://doi.org/10.2134/jeq2019.02.0041>

607 Ojeda G., Ortiz O., Medina C.R., Perera I., Alcaniz J.M. (2015). Carbon sequestration in a
608 limestone quarry mine soil amended with sewage sludge *Soil Use and Manage*, 31 (2),
609 270-278, <https://doi.org/10.1111/sum.12179>

610 Piirainen S., Finer L., Mannerkoski H., Starr M. (2002). Effects of forest clear-cutting on the
611 carbon and nitrogen fluxes through podzolic soil horizons *Plant and Soil*, 239, 301-
612 311, <https://doi.org/10.1023/A:1015031718162>

613 Prescott C.E., Blevins L.L., Staley C.L. (2000). Effects of clear-cutting on decomposition
614 rates of litter and forest floor in forests of British Columbia *Can. J. For. Res.* 30,
615 1751–1757 <https://doi.org/10.1139/x00-102>

616 Prescott C.E. (2005). Do rates of litter decomposition tell us anything we really need to
617 know? *For. Ecol. Manag.*, 220, 66-74 <https://doi.org/10.1016/j.foreco.2005.08.005>

618 Rice, J.A., MacCarthy, P. (1991). Statistical evaluation of the elemental composition of humic
619 substances. *Org. Geochem.* 17 (5), 635–648 [https://doi.org/10.1016/0146-](https://doi.org/10.1016/0146-6380(91)90006-6)
620 [6380\(91\)90006-6](https://doi.org/10.1016/0146-6380(91)90006-6)

621 Rocha J.H.T., Marques E.R.G., Goncalves J.L.d.M., Hübner A., Brandani C.B., Ferraz A.d.V.,
622 Moreira R.M. (2016). Decomposition rates of forest residues and soil fertility after
623 clear-cutting of *Eucalyptus grandis* stands in response to site management and
624 fertilizer application *Soil Use Manag.* 32 (3), 289-302,
625 <https://doi.org/10.1111/sum.12283>

626 Schnitzer M. (1994). A chemical structure for humic acid. Chemical, ¹³C NMR, colloid
627 chemical, and electron microscopic evidence. In: *Humic Substances in the Global*
628 *Environment and Implications on Human Health*, N. Senesi, and T. M. Miano (eds.).
629 Proc. 6th Intern. Meeting of the Intern. Humic Substances Soc., Monopoli, Bari,
630 Italy, September 20–25, 1992. Elsevier, Amsterdam, 57–69.

631 Senesi N. (1990). Application of Electron Spin Resonance (ESR) spectroscopy in soil
632 chemistry. *Advances in Soil Science*, 14, 77-130

633 Senesi N., Loffredo E. (1999). The chemistry of soil organic matter In: Sparks D.L. (ed) *Soil*
634 *Physical Chemistry*, second ed. CRC Press, Boca Raton, 239-370

635 Senesi, N., Loffredo, E. (2001). Soil humic substances. In: Steinbuchel, A. (Ed.),
636 *Biopolymers*, Wiley-VCH, 1, 247 – 299

637 Senesi N., D’Orazio V., Ricca G. (2003). Humic acids in the first generation of EUROSOLS
638 *Geoderma*, 116, 325-344 [https://doi.org/10.1016/S0016-7061\(03\)00107-1](https://doi.org/10.1016/S0016-7061(03)00107-1)

639 Senesi N., Schnitzer M. (1977). Effect of pH, Reaction Time, Chemical Reduction and
640 Irradiation on ESR Spectra of Fulvic Acid. *Soil Science*, Vol. 123, 224-234

641 https://journals.lww.com/soilsci/Abstract/1977/04000/EFFECTS_OF_pH,_REACTI
642 [ON_TIME,_CHEMICAL_REDUCTION.3.aspx](https://journals.lww.com/soilsci/Abstract/1977/04000/EFFECTS_OF_pH,_REACTI)

643 Smolander A., Törmänen T., Kitunen V., Lindroos A.-J. (2019). Dynamics of soil nitrogen
644 cycling and losses under Norway spruce logging residues on a clear-cut *For. Ecol.*
645 *Manag.*, 449, 117444, <https://doi.org/10.1016/j.foreco.2019.06.041>

646 Stevenson F.J. (1994). *Humus Chemistry: Genesis, Composition, Reactions*, 2nd ed. Wiley,
647 New York, 496 pp.

648 Swift, R.S., (1996). Organic matter characterization. In: *Methods of soil analysis. Part 3.*
649 *Chemical methods – SSSA Book Series no.5* Soil Science Society of America and
650 American Society of Agronomy, pp. 1011–1068
651 <https://doi.org/10.2136/sssabookser5.3.c35>

652 Tan, K.H. (2014). *Humic Matter in Soil and the Environment: Principles and Controversies*,
653 2nd edn, CRC Press, 439 pp. <https://doi.org/10.1201/b17037>

654 Traversa, A., D’Orazio, V., Senesi, N. (2008). Properties of dissolved organic matter in forest
655 soils: influence of different plant covering. *For. Ecol. Manag.* 256, 2018– 2028
656 <https://doi.org/10.1016/j.foreco.2008.07.038>

657 Ussiri D.A.N. & Johnson C.E. (2007). Organic matter composition and dynamics in a
658 northern hardwood forest ecosystem 15 years after clear-cutting *For. Ecol. Manag.*, 240,
659 131-142 <https://doi.org/10.1016/j.foreco.2006.12.017>

660 Valenzuela E. I., Cervantes F.J. (2021). The role of humic substances in mitigating greenhouse
661 gases emissions: Current knowledge and research gaps *Sci. Total Environ.*, 750, 141677,
662 <https://doi.org/10.1016/j.scitotenv.2020.141677>

663 Weber, J., Chen, Y., Jamroz, E., Miano, T. (2018). Preface: Humic substances in the
664 environment *J. Soils Sediments*, 18, 2665-2667, <https://doi.org/10.1007/s11368-018->
665 [2052-x](https://doi.org/10.1007/s11368-018-2052-x)

666Zaccone C., Pabst S., Senesi G.S., Shotyk W., Miano T.M. (2013). Comparative evaluation of
667 the mineralogical composition of Sphagnum peat and their corresponding humic acids,
668 and implications for understanding past dust depositions *Quaternary International* 306,
669 80-87, <https://doi.org/10.1016/j.quaint.2013.04.017>

670Ziolkowska A., Debska B. & Banach-Szott M. (2020). Transformations of phenolic
671 compounds in meadow soils. *Sci. Rep.* 10, 19330, [https://doi.org/10.1038/s41598-020-](https://doi.org/10.1038/s41598-020-76316-7)
672 [76316-7](https://doi.org/10.1038/s41598-020-76316-7)

673

674

675

676

677

678

679

680