

**Analysis and Optimal Design of Batch and Two-Column Continuous
Chromatographic Frontal Processes for Monoclonal Antibody
Purification**

**Ce Shi^a, Sebastian Vogg^b, Dong-Qiang Lin^a, Mattia Sponchioni^c,
Massimo Morbidelli^{d*}**

a. Key Laboratory of Biomass Chemical Engineering of Ministry of Education,
College of Chemical and Biological Engineering, Zhejiang University, Hangzhou
310027, China

b. YMC ChromaCon, Technoparkstrasse 1, 8005 Zürich, Switzerland

c. Department of Chemistry, Materials and Chemical Engineering “Giulio Natta”,
Politecnico di Milano, Via Mancinelli 7, 20131 Milano, Italy

d. Institute for Chemical and Bioengineering, Department of Chemistry and Applied
Biosciences, ETH 11 Zurich, Vladimir-Prelog-Weg 1-5/10, 8093 Zürich, Switzerland

1. Introduction

The increasing market share of monoclonal antibodies (mAbs), with 10% annual growth and more than 70 formulations approved in the last decade, makes mAbs one of the most important classes of biopharmaceuticals today^[1-5]. The increasing number of therapeutic indications and the emerging of biosimilars contribute to strengthen the

demand for safe, efficient and cost effective manufacturing processes^[6,7]. Continuous integrated bioprocessing indeed goes in this direction^[8-10] and it is therefore encouraged by regulatory authorities ^[11,12].

Frontal chromatography has seen increased interest for protein purification, in particular as a polishing step in downstream processes for therapeutic proteins production, as for example in the purification of monoclonal antibodies (mAbs) from high molecular weight impurities, e.g., aggregates, using cation exchange resins^[13-17]. The schematic diagram of frontal chromatography operated on a single (batch) column is shown in Figure 1a. Here, the eluate from the capture step (feed), containing both the monomer and the aggregates, is first loaded into the column. The impurity (aggregates) binds stronger to the cation exchange resin and displaces the weaker binding product (monomer), which thus elutes first and is collected in the product pool. Loading is continued until reaching, in general, sufficiently high purity in the pool and relatively high recovery (yield). Next, a washing step is applied to recover the residual monomer still present in the column into the product pool, so as to further increase the yield, but being careful not to elute also the aggregates, which would spoil the purity below specifications. Finally, regeneration and re-equilibration (RR) are carried out to elute all the impurities (and remaining product) into the waste and prepare the column for the next loading step.

In such a batch operation, the loading time and linear velocity have to be properly selected so as to process the largest amount of material, without letting too much aggregate into the product pool, so as to preserve purity. In the following washing step,

the linear velocity and duration should be carefully chosen to elute as much as possible of the remaining product, thus maximizing the yield, while not desorbing the impurity beyond the purity specification ^[18]. Therefore, batch frontal chromatography suffers from an intrinsic purity-yield tradeoff. High loadings and harsh washing conditions enable the recovery of much of the product, and thus an improved monomer yield, but an overall low purity. On the other hand, low loadings and too mild washing conditions enable high purity of the pool but prevent full recovery of the product, at the expense of the process yield.

In order to alleviate this purity-yield tradeoff, a novel cyclic two column continuous chromatographic process (referred to as Flow2) has been developed ^[18]. The schematic diagram is illustrated in Figure 1b. This is a periodic process, where each switch is constituted of three steps, after which the two columns exchange their role. In the first step, the two columns are fed while being interconnected, so that the breakthrough of the aggregates from the first column is captured by the second column. In the next interconnected washing step, with inline dilution between the two columns, both the product and the unbound impurity left in the first column are eluted into the second column. With the proper inline dilution, both of them are bound in the second column. Finally, in the third step, the two columns are disconnected. The first one undergoes the RR process while the second one is loaded with the fresh feed.

With two columns interconnected during loading and the following washing with inline dilution, aggregates are prevented from eluting from the second column into the product pool and more monomer can be eluted from the first column, thus improving

the tradeoff between purity and yield. In addition, the washing conditions (buffer and duration) do not need to be carefully designed as in the case of batch operation, thus increasing the process robustness. However, the design of this cyclic process needs to account for the increased number of operational parameters and complexity of the process dynamics, which can be best achieved using model-based approaches [19-24].

In this work, the fundamental aspects of batch and continuous frontal chromatography are analyzed. A model-based optimal design procedure, valid for both batch and Flow2 processes, is developed with reference to three process performance parameters: yield (recovery), purity and productivity. In particular, Pareto Fronts of productivity and yield, with purity fixed at given specification values (P_{spec}), computed based on reliable chromatographic models of the two processes, are discussed. Next, the two processes, each operated at its own optimal conditions, are compared. This analysis is conducted with reference to the polishing of a mAb of industrial relevance and accounts, in addition to yield, purity and productivity [25], also for the important concept of process robustness, which is a major concern particularly in biopharmaceutical applications [26,27]. This has been quantified using a rigorous model-based sensitivity analysis of the process performance with respect to its operating process parameters.

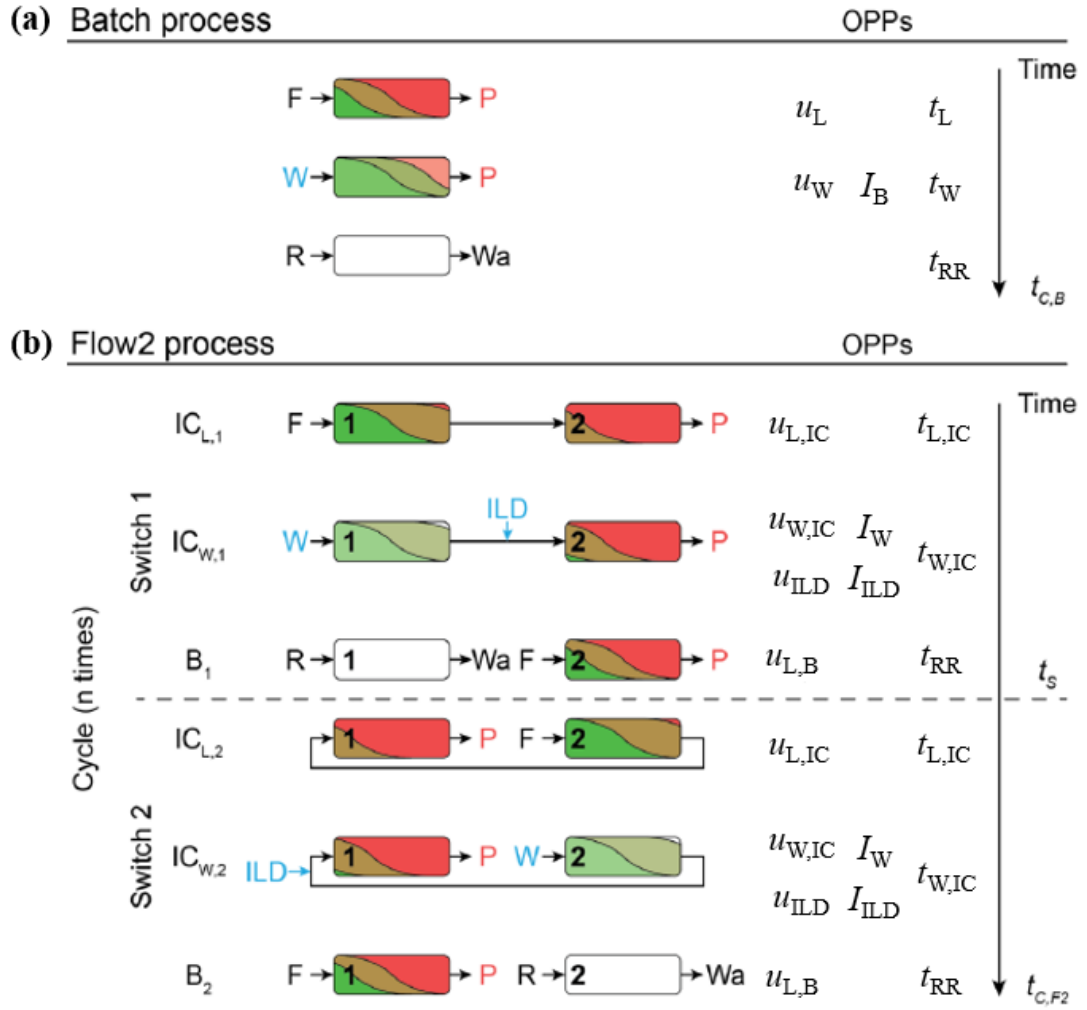


Figure 1. Schematic diagram of batch frontal chromatography (a) and continuous Flow2 processes (b)^[18].

2. Model-Based Optimal Design Methods

2.1 The Chromatographic Model

The chromatographic lumped kinetic model with linear driving force approximation has been used in all simulations ^[16,28,29]:

$$\frac{\partial c_i}{\partial t} = -\frac{u_{sf}}{\varepsilon_{t,i}} \frac{\partial c_i}{\partial x} + \frac{u_{sf}}{\varepsilon_{t,i}} d_{ax,i} \frac{\partial^2 c_i}{\partial x^2} - \frac{1-\varepsilon_{t,i}}{\varepsilon_{t,i}} \frac{\partial q_i}{\partial t} \quad (1)$$

$$\frac{\partial q_i}{\partial t} = k_{m,i} (q_i^{eq} - q_i) \quad (2)$$

with $i = M, A$ and the boundary conditions:

$$c_i(x, t = 0) = 0$$

$$c_I(x, t = 0) = I_0$$

$$c_{pH}(x, t = 0) = pH_0$$

$$c_i(x, t = 0) = c_{i,in}(t) + d_{ax,i} \left. \frac{\partial c_i}{\partial x} \right|_{x=0}$$

$$\left. \frac{\partial c_i}{\partial x} \right|_{x=L_{col}} = 0$$

The mixing node in the continuous process has been simulated as follows:

$$c_{i,in,col2} = \frac{C_{i,in,col1} u_{W,IC} + C_{i,ILD} u_{ILD}}{u_{W,IC} + u_{ILD}} \quad (3)$$

The adsorption equilibrium has been described through a surrogate model, which mimics the behavior of the DLVO-derived model, based on the competitive Langmuir isotherm and appropriate empirical correlations ^[18,30] as follows:

$$q_i^{eq} = \frac{H_i c_i}{1 + \sum_j \frac{H_j c_j}{q_j^{sat}}} \quad (4)$$

$$q_i^{sat} = a_i^{sat} pH + b_i^{sat} \quad (5)$$

$$H_i = \alpha_i I^{-\beta_i} \quad (6)$$

$$\log_{10} \alpha_i = a_i^\alpha pH + b_i^\alpha \quad (7)$$

$$\beta_i = a_i^\beta pH + b_i^\beta \quad (8)$$

The meaning of all variables and parameters is explained in the notation section, while the parameter values adopted in the simulations are listed in Table 1.

Table 1. Values of the chromatographic model parameters ^[18]

	Parameter	Unit	Salt	Monomer	HMW
d_{col}	Column diameter	cm	-----0.5-----		
L_{col}	Bed height	cm	-----5-----		
ε_b	Bed porosity	-	-----0.39-----		
$\varepsilon_{t,i}$	(Accessible) total porosity	-	0.95	0.65	0.65
$\varepsilon_{p,i}$	(Accessible) particle porosity	-	0.93	0.43	0.43
a_i^{Sat}	Slope of q_i^{Sat} vs. pH	-	n/a	0	0
b_i^{Sat}	Intercept of q_i^{Sat} vs. pH	-	n/a	212	106
a_i^{α}	Slope of $\log_{10}\alpha$ vs. pH	-	n/a	-1.270	-3.090
b_i^{α}	Intercept of $\log_{10}\alpha$ vs. pH	-	n/a	31.22	46.90
a_i^{β}	Slope of β vs. pH	-	n/a	1.128	0.676
b_i^{β}	Intercept of β vs. pH	-	n/a	5.227	9.870
$d_{ax,i}$	Axial dispersion coefficient	cm	0.034	37	98
$k_{m,i}$	Mass transfer coefficient	1/min	170	1.3	0.53

2.2 Process performance parameters

As mentioned above, the process performance is quantified in the following using three process performance parameters: the productivity, Pr of the target species, i.e., mAb, defined as:

$$Pr = \frac{m}{n_{col} V_{col} t_C} \quad (9)$$

where m is the mass of target recovered in the product pool in one cycle of duration t_C

using n_{col} columns, each with volume V_{col} , the yield, Y of the target protein defined as:

$$Y_i = \frac{m}{m_{\text{load}}} \quad (10)$$

where m_{load} is the amount of target protein loaded on the column in one cycle, and the purity, P defined as:

$$P = \frac{m}{\sum_i m_i} \quad (11)$$

where the sum at the denominator is extended to the mass m_i of all proteins present in the product pool.

2.3 Optimization methods

The optimal design of frontal chromatographic processes can be complex. For this, a specific procedure has been devised, based on a series of suitable steps, and referred to as the *design procedure* (DP) in the following. This allows to facilitate and guarantee the identification of the optimal operating conditions, for a given set of performance parameters. The obtained results are compared with corresponding results obtained by a multi-objective optimization procedure based on Genetic Algorithms ^[31].

The general concept of the design procedure is based on the evaluation of the loading and washing times (see Figure 1), which describe the Pareto Front of productivity and yield at a given purity equal to the specification value, P_{spec} . The regeneration and re-equilibration time, t_{RR} (Figure 1) is expected to derive from a specifically designed experimental study conducted on a single column and is therefore considered as a given value in this work. The basic idea is to first compute, all the other parameters and operating conditions being fixed, the loading times leading to purity

values at the end of the first loading step, P_1 ranging from 100% to P_{spec} . In the following washing step, the process purity can only decrease and the corresponding duration is computed such that, for each one of the previous P_1 values, it leads to the required purity P_{spec} . For each pair of loading and washing time values, we then compute the corresponding productivity and yield values, being in all cases the purity equal to P_{spec} . At this point, we can order all the obtained values in a productivity versus yield plot and obtain the desired Pareto Front at fixed purity. This procedure can be repeated by changing any other of the relevant parameters, like the buffer compositions or the column or particle size, and compute the new Pareto Front. This approach is useful not only to find the optimal operating conditions of the process, but also to quantify the robustness of a given process design, as well as the economic impact of the different operating conditions.

Alternatively, the optimization problem can be approached as a whole, without attempting any problem decomposition, using a fully general multi-objective optimization algorithm. The function ‘Gamultiobj’ of the MATLAB library ^[32], based on a Genetic Algorithm, has been used in this work and the obtained results are compared with those of the *ad-hoc* design procedure.

In the next section, we describe in detail the algorithm to compute the optimal loading and washing times described above for both the batch and the Flow2 processes.

3. Optimal Design of Frontal Chromatography Processes

3.1. The Batch Process

The Pareto Front of productivity and yield with constant fixed P_{spec} , computed

with both methods, are compared first in the case of a batch process. All parameters, unless otherwise specified, are kept constant at the values summarized in Table 2.

Table 2. Batch Operating Conditions

Parameters	Unit	Value
$c_{L, mono}$	g/L	4.5
$c_{L, agg}$	g/L	0.5
I_L	mM	80
I_W	mM	120
I_{RR}	mM	1000
pH_L	-	5
pH_W	-	5
pH_{RR}	-	5
u_L	cm/hr	300
u_W	cm/hr	300
u_{RR}	cm/hr	300
t_{RR}	min	15

With illustrative purposes, the requested purity the value was set as $P_{spec} = 99\%$. The procedure develops according to the following three steps:

Step 1: Compute the purity values at the column outlet, P_1 during the loading phase (Figure 1a) for increasing loading time values, t_L .

As t_L increases, first the product breaks through from the outlet of the column into

the product pool, followed by the impurity at longer times, thus reducing the purity in the product pool, P_1 , as shown in Figure 2a. As the loading proceeds the purity decreases from 100% until $P_1 = P_{\text{spec}}$, which indicates the maximum acceptable value of t_L . In this conditions, in fact, the washing time in the next step should be set to zero to avoid any further decrease in purity below specification, but of course this would not allow for any improvement in the process yield. Accordingly, at optimal operation, the value of t_L should be shorter than 72 min, which corresponds to P_1 values ranging from 100% to the $P_{\text{spec}} = 99\%$ value.

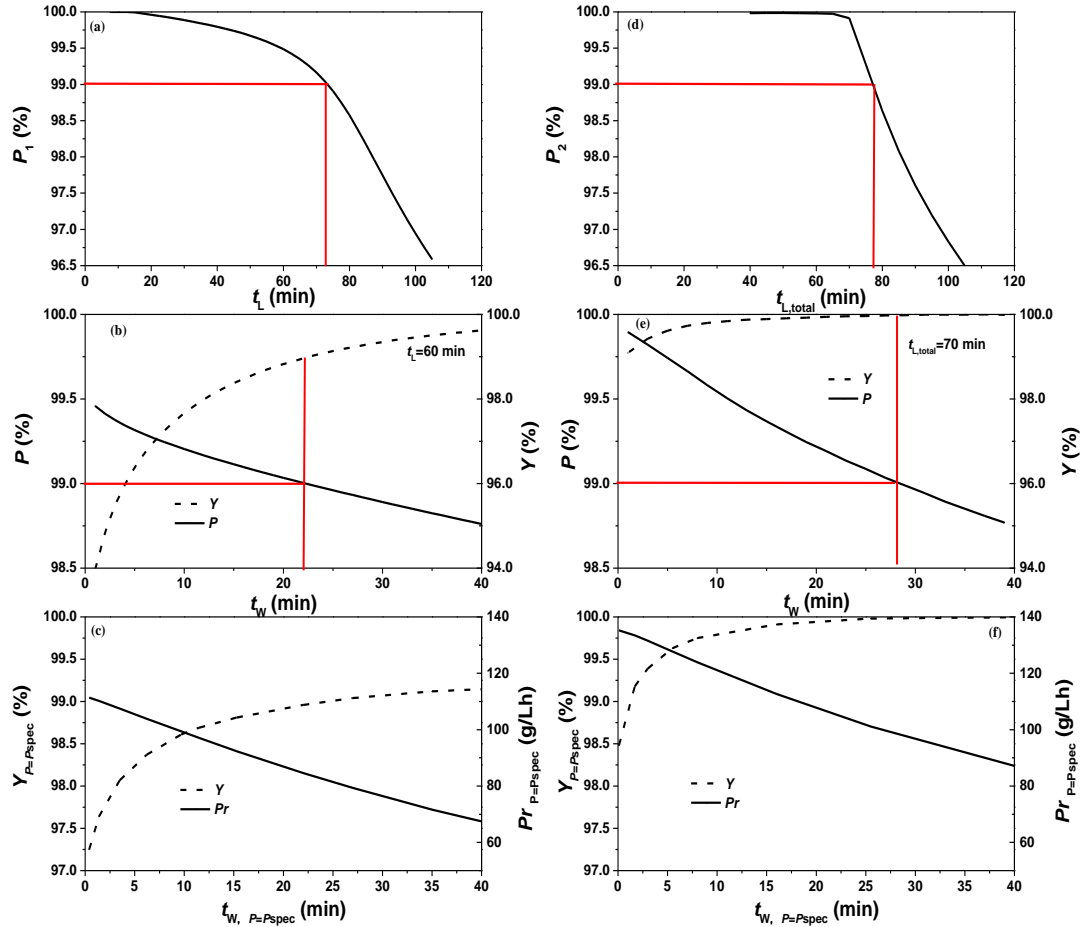
Step 2: For each of these pairs of values t_L and P_1 , compute the corresponding washing time, t_w , which, starting from P_1 , leads to a final purity, P equal to P_{spec} .

The obtained process yield and purity values are shown in Figure 2b as a function of the washing time, t_w , as an example for the case of $t_L=60$ min and $P_1=99.5\%$. It is seen that the process purity, P decreases while Y increases for increasing washing times. The decrease in the process purity is because more eluate, including both product and impurity, is washed out into the product pool. On the other hand, the yield increases when increasing t_w since more product is recovered. By selecting the purity value equal to P_{spec} , the corresponding values of the yield, $Y_{P=P_{\text{spec}}}$ and the washing time, $t_{W,P=P_{\text{spec}}}$ are found.

Step 3: Repeat the procedure above to calculate productivity, Pr and yield, Y for each of the pairs of values t_L and P_1 in Figure 2a and derive the corresponding Pareto Front at fixed purity equal to P_{spec} .

Using the t_L and P_1 values in Figure 2a, the curve shown in Figure 2c is obtained,

199 representing the yield as a function of the washing time leading to process purity, $P =$
 200 99%. On the other hand, it is seen that the corresponding productivity values, computed
 201 through equation (19), decrease with t_{wash} , leading to the trade-off between Pr and Y
 202 illustrated by the Pareto Front, with fixed purity P_{spec} , in Figure 3.



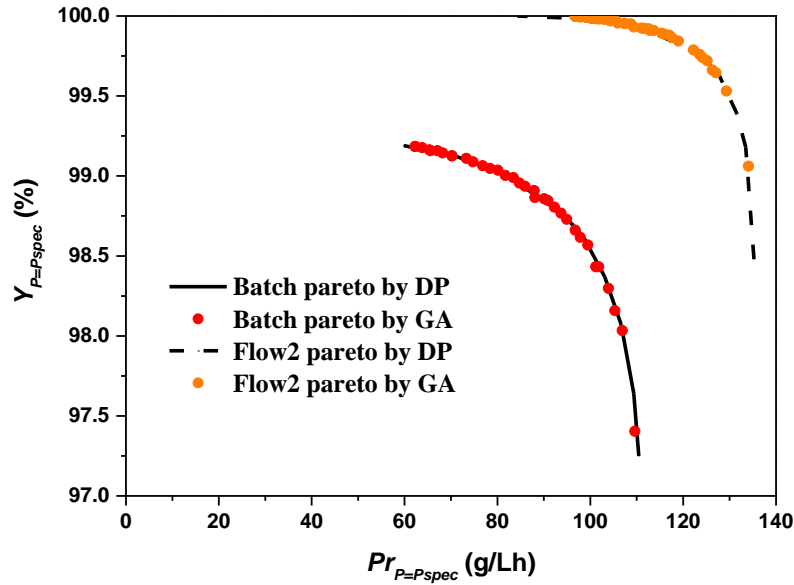
203
 204 Figure 2. The design procedure for the batch and Flow2 processes. (a) Batch P_1 as a
 205 function of the loading time; the bar indicates P_{spec} (b) Batch P and Y as a function of
 206 the washing time at $t_L = 60$ min and $P_1 = 99.5\%$ (c) The relationship between $Y_{P=P_{\text{spec}}}$
 207 and $Pr_{P=P_{\text{spec}}}$ as a function of $t_{W, P=P_{\text{spec}}}$ for a batch process. (d) Flow2 P_2 as a function
 208 of the loading time, $t_{L,\text{total}}$; the bar indicates P_{spec} (e) Flow2 P and Y as a function of the
 209 washing time at $t_{L,\text{total}} = 70$ min and $P_2 = 99.8\%$ (f) The relationship between $Y_{P=P_{\text{spec}}}$

and $Pr_{P=P_{spec}}$ as a function of $t_{W,P=P_{spec}}$ for a Flow2 process.

The results of the *ad-hoc developed design procedure* (DP) are compared with those of a general multiobject Genetic Algorithm (GA), using 50 individuals per generation and a maximum of 200 generations. The purity constraint has been introduced by imposing heavy penalties on purity values smaller than P_{spec} . At each iteration, the 35 fittest individuals are selected to plot the Pareto Front and produce the next generation. The GA optimization is terminated when the 200th generation is reached or when the average relative change in the best fitness function value, with respect to the previous generation, is less than or equal to the function tolerance (10^{-4}). The performance values corresponding to the best 35 individuals in the last generation are compared with the Pareto Front computed through the design procedure in Figure 3. It is seen that all the points corresponding to the GA optimization are on or slightly below the curve generated by the design procedure, indicating that the two methods provide essentially the same result. In addition, it is found that all the points obtained through the GA method exhibit a purity value equal to the constrain value of 99% with a maximum 1 % relative deviation.

It is worth noticing that, although the two methods give equivalent results, the associated computational effort is quite different. In the *design procedure*, only 35*10 objective functions are calculated: 35 for getting the Pareto Front, and 10 each for determining the t_W values corresponding to 99% purity. On the other hand, in the multi-objects GA optimization, 50*200 functions are calculated: one each for the 50

232 individuals constituting each of the 200 generations.



233

234 *Figure 3. Pareto Front of Pr and Y computed with the design procedure (DP)*
 235 *compared with the results of the global optimization based on genetic algorithm (GA),*
 236 *for $P_{spec} = 99\%$ and other parameter values as in Tables 1, 2 and 3.*

237

238 3.2 The Continuous Flow2 Process

239 The two optimization methods considered above have been developed and
 240 compared also for the Flow2 process. In particular, we optimize the interconnected
 241 loading and washing times, $t_{L,IC}$ and $t_{W,IC}$ (Figure 1b) in order to obtain the Pareto Front
 242 of productivity, Pr and Y with fixed purity equal to P_{spec} . Note that in this process, the
 243 batch loading time is fixed and equal to the regeneration and re-equilibration time, t_{RR} ,
 244 which is considered as a given constant in this analysis (Figure 1b). Accordingly, the
 245 total loading time given by $t_{L,total} = t_{RR} + t_{L,IC}$, can actually be changed only through the
 246 interconnected loading time, $t_{L,IC}$. All remaining parameters are kept constant and equal

to the values summarized in Tables 1 and 3. It is to be noted that all the simulation results reported for the continuous Flow2 process refer to steady state conditions. These are obtained by simulating the entire dynamics of the process and considering the transient behavior completed when the mass flow of each protein entering and leaving differ for no more than 1.0 % , which means that no protein is accumulated or lost in the system during every switch.

Table 3. Flow2 Operating Conditions

Parameters	Unit	Value
$c_{L, mono}$	g/L	4.5
$c_{L, agg}$	g/L	0.5
I_{IC}	mM	80
I_W	mM	120
I_{ILD}	mM	10
I_{RR}	mM	1000
$pH_{L,IC}$	-	5
$pH_{W,IC}$	-	5
pH_{ILD}	-	5
pH_{RR}	-	5
$u_{L,IC}$	cm/hr	300
u_W	cm/hr	300
u_{ILD}	cm/hr	300
u_{RR}	cm/hr	300

As mentioned above, differently from the batch process, the Flow2 process has two loading phases within one switch: interconnected and batch loading. In the following, we consider these two steps together with a total duration, $t_{L,total} = t_{RR} + t_{L,IC}$, with a purity value P_2 , which is analogous to the purity P_1 in the batch process. After the washing step, the product purity is indicated as P which represents the final product purity at the end of the switch as indicated in Figure 1b.

The design procedure in this case is organized as follows:

Step 1: Simulate the process for changing total loading times, $t_{L,total}$ so as to obtain purity values, P_2 ranging from 100% to the requested $P_{spec}=99\%$.

As shown in Figure 2d, the obtained P_2 values decrease as a function of the loading time, $t_{L,total}$: starting from 100% to values below the imposed process purity specification, P_{spec} , which is reached at $t_{L,total} = 78$ min.

Step 2: For each pairs of values $t_{L,total}$ and P_2 in Figure 2d, with $P_2 > P_{spec}$, compute the interconnected washing time ($t_{W,IC}$ in Figure 1b) leading to a product purity, P at the end of the switch equal to the requested P_{spec} .

As an example, Figure 2e shows the results obtained for the pair of values $t_{L,total} = 70$ min and $P_2 = 99.8\%$. As expected, it is seen that as the washing time increases, the purity decreases while the yield increases. The values $Y_{P=P_{spec}}$ and $t_{W,P=P_{spec}}$, corresponding to the requested purity value P_{spec} , can be obtained.

Step 3: Repeat the previous step for each of the pairs of values P_2 and $t_{L,total}$ in Figure 2d and with the corresponding productivity, Pr and yield, Y values derive the

corresponding Pareto Front, at fixed purity equal to P_{spec} .

Repeating the step above for all the pairs of values P_2 and $t_{L,total}$ in Figure 2d, the maximum yield values for each washing time, t_w , compatible with the fixed purity, P_{spec} can be computed as shown in Figure 2f. From these, and the corresponding productivity values, Pr computed through Equation 9, the Pareto Front for productivity and yield at fixed purity equal to P_{spec} in Figure 3 is obtained.

The latter is compared in the same figure with the corresponding Pareto Front for the batch frontal chromatography process, obtained as discussed above. The higher efficiency of the Flow2 process, which will be discussed later in more detail, is clearly indicated by the movement of the Pareto Front to the upper right corner of the figure.

As mentioned above, the multi-object global optimization of the Flow 2 process has also been performed based on a genetic algorithm, using 50 individuals per generation and a maximum of 300 generations. The obtained results, corresponding to the 35 best individuals in the last generation, are shown by the points shown in Figure 3 and compared with the results of the ad-hoc design procedure. It is seen that the two methods provide equivalent results, although the GA approach requires a significantly larger computational effort, as already discussed in the context of the batch process.

4. Analysis and Comparison of Optimized Batch and Flow 2 Process Performance

A general difficulty when comparing different processes, and not only in chromatography, is to select “fair” conditions for the comparison, that is operating conditions that do not favor one or the other. In this work, we consider each process

under independently optimized operating conditions. In particular, we are going to analyze the behavior of batch and Flow 2 processes as a function of operating parameters which are known to be most relevant in chromatography, such as linear velocities in the columns and composition of the loading and washing buffers. In all cases, we consider for each process the optimal Pareto yield-productivity (with fixed purity, P_{spec}) obtained by optimizing the durations of both the loading and washing steps. In the following, the design procedure developed in this work will be used, since we have shown that this provides the same results as a general multi-objective optimization, but with lower computational effort. The objective is to analyze and compare the behavior of each of such processes with respect to both process performance and robustness.

4.1 The relevant performance parameters

Among the various chromatographic steps involved in the production of mAbs, frontal chromatography is a quite promising candidate for the polishing step after protein capture, where high purity, productivity and robustness are requested. For example, in the case of aggregate removal, mAb purity values well above 95% are typically needed. In Figure 4, the tradeoff between productivity and yield for optimal batch process operations at specification purities, P_{spec} equal to 95%, 98% and 99% is shown. It can be seen that more stringent purity constraints move the Pareto Front to the lower left corner with correspondingly lower values for both productivity and yield. It can also be seen that the yield is always higher than 98%, indicating that this is not the discriminating performance parameter in designing frontal chromatography

processes. Accordingly, in the following we will base the process comparison on the tradeoff between yield and productivity as well as on the operation robustness, with a stringent purity specification of $P_{\text{spec}} = 99\%$.

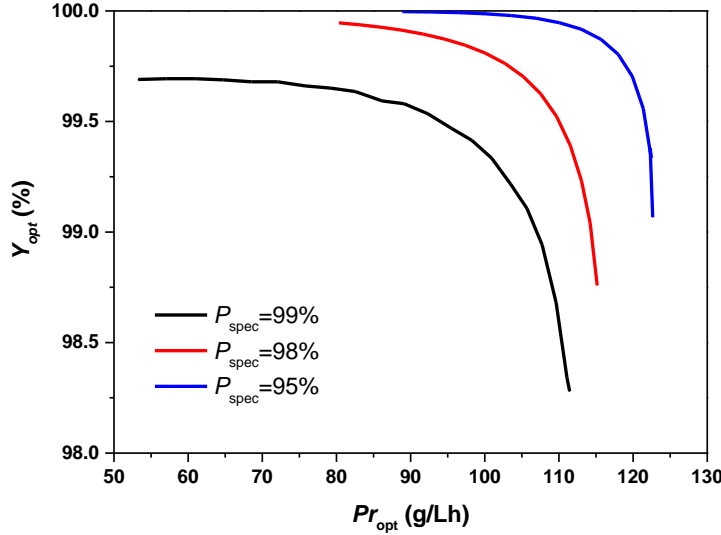


Figure 4. Tradeoff between yield and productivity (Pareto Front) for the batch process at various P_{spec} values. Operating conditions as in Tables 1 and 2.

4.2 Role of linear velocities in loading and washing

In this section, the effect of the liquid velocity during loading and washing is analyzed in both Batch and Flow2 process. The values considered range from 300 cm/hr to the value compatible with the maximum pressure drop tolerated by the stationary phase as computed through the Blake-Kozeny equation ^[33]

$$\frac{\Delta P}{L_{\text{col}}} = \frac{150\mu}{d_p^2} \frac{(1-\varepsilon_b)^2}{\varepsilon_b^3} u_{\text{sf}} \quad (12)$$

Considering the maximum pressure drop equal to 0.15 MPa^[34], the maximum linear velocity of 3000 cm/hr is obtained for the 5 cm bed height and 50 μm particle radius

considered in this work. The values of the remaining operating parameters used in all simulations considered in the following are listed in Tables 1 to 3.

4.2.1 Batch process

The effect of loading and washing velocities is analyzed first with u_L changing from 300 to 3000 cm/hr with fixed $u_W = 300$ cm/hr, and then with u_W changing from 1 to 3000 cm/hr with fixed $u_L = 300$ cm/hr. The Pareto fronts obtained for various u_L values are shown in Figure 5a. Note that, in general, the Pareto fronts do not necessarily cover the entire range of possible yield and productivity values, since some portions of them may not be achievable with the considered operating conditions. In particular, it is seen that as the loading velocity increases, the process performance first improves, with the Pareto moving to the right of the plot, thus enabling higher productivity values for a given recovery value. This is because at higher loading velocity the loading time is shortened to the benefit of productivity. However, when further increasing the loading velocity, intraparticle mass transfer becomes more and more limiting [35,36], leading eventually to the earlier breakthrough of the impurities and then to lower purity values. This explains the worsening of the Pareto front at 3000 cm/hr with respect to lower loading velocities in Figure 5a. The effect of the washing velocity is showed in Figure 5b. It is seen that as the washing velocity increases from 300 to 900 cm/hr, the Pareto front extends to higher productivities, but it decreases again for further increasing u_W values. For the maximum allowable washing linear velocity of 3000 cm/hr, the process performance is in fact inferior to all the other washing velocities in almost the entire range of operating conditions. Limiting intraparticle mass transfer prevails at such

washing linear velocity, so that more target protein remains in the column, and both yield and productivity decrease.

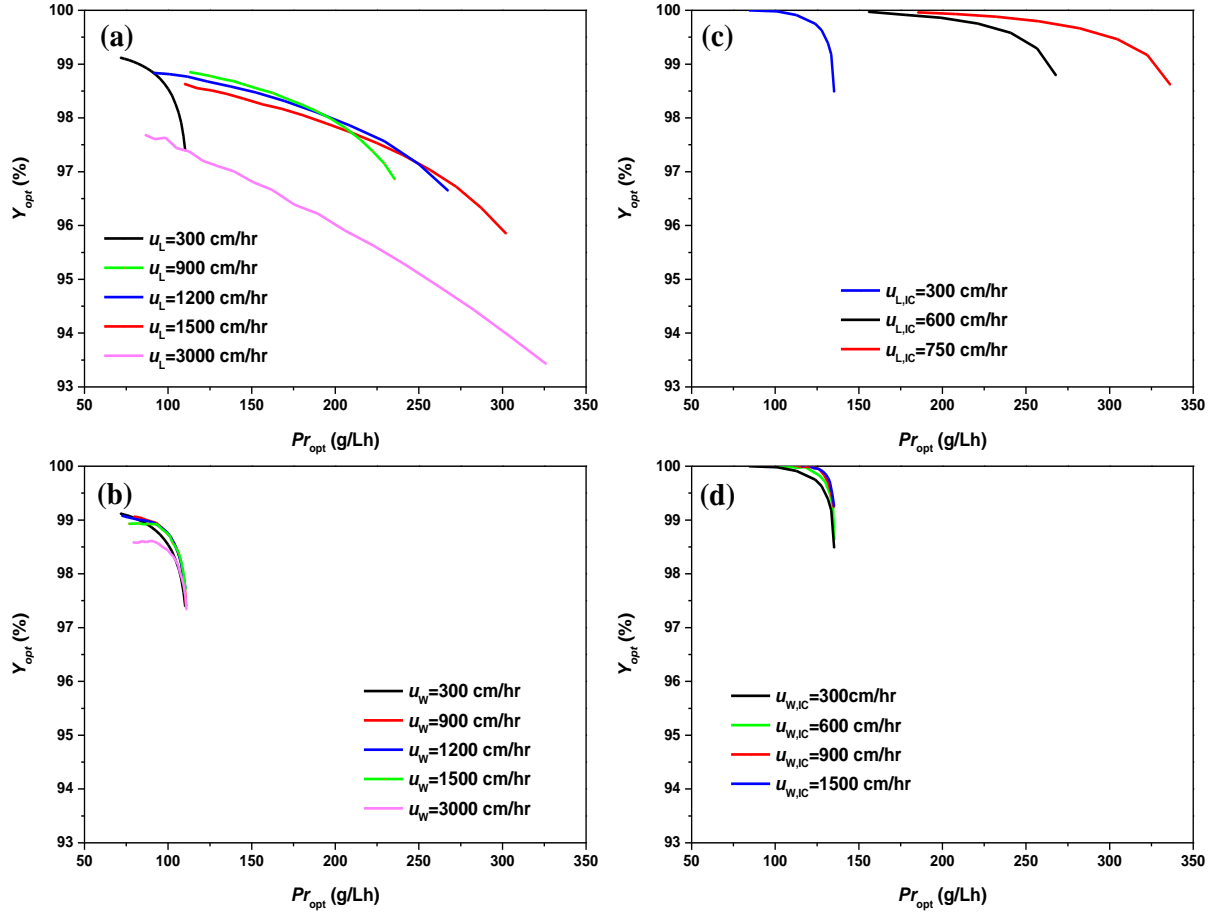


Figure 5. Tradeoff (Pareto front) between yield and productivity of (a) batch process under various loading velocities with $u_W = 300$ cm/hr (b) batch process under various washing velocities with $u_L = 300$ cm/hr (c) Flow2 process under various interconnected loading velocities with $u_{W,IC} = 300$ cm/hr (d) Flow2 process under various interconnected washing velocities with $u_{L,IC} = 300$ cm/hr

Thus summarizing, since the Pareto fronts in both Figures 5a and 5b tend to cross each other, the best velocity values have to be defined depending on the specific operating conditions. In particular, the idea of using the maximum possible velocity in

either loading or washing, corresponding in this case to 3000 cm/hr, is not recommended.

4.2.2 Continuous Flow2 process

The analysis of the role of liquid velocities in this case is more complex than for batch processes because we now deal with four velocities: the interconnected loading velocity, $u_{L,IC}$, the interconnected washing velocity, $u_{W,IC}$, the inline dilution velocity, u_{ILD} and the batch loading velocity, $u_{L,B}$. It appears reasonable to take equal the batch and interconnected loading velocities, i.e. $u_{L,IC} = u_{L,B}$, so as to apply in both the same residence time. In addition, the inline dilution velocity is taken equal to the interconnected washing velocity, i.e. $u_{W,IC} = u_{ILD}$ as the resulting buffer composition for the downstream column remains constant independent of the linear velocity applied, thereby reducing the number of varied operating variables. Accordingly, in the following we analyze the effect of two velocities: $u_{L,IC}$ and $u_{W,IC}$. These will be changed independently, one at a time, in the range 300 cm/hr to 1500 cm/hr. The latter corresponds to the largest velocity compatible with the considered maximum pressure drop of 0.15 MPa, and it is half of the one considered in the case of the batch process because two columns are operated in series in the interconnected step. The simulation results are shown in Figure 5c and 5d.

In Figure 5c it is seen that larger values of the loading velocity, $u_{L,IC}$ improve significantly the tradeoff between yield and productivity of the Flow2 process. However, no Pareto Front could be calculated for loading velocities larger than 750 cm/hr, since it was not possible to achieve the requested purity of 99%. For such values of $u_{L,IC}$ (and

of $u_{L,B}$), together with the fact that the batch loading time must be fixed to match the R-R time, too many aggregates breakthrough into the product pool. Nevertheless, it is found that, although the loading velocity is constrained by time scheduling, the Flow2 process can offer better performances than the batch operation. In particular, it can approach productivities larger than 300 g/L/h while keeping the yield in the order of 99%, which for the batch process (Figure 5a) can only be achieved by decreasing the productivity by at least three times in the order of 100 g/L/h.

Also in the case of the washing step, the process performance improves for increasing linear velocity values, $u_{W,IC}$ as shown in Figure 5d. This is because in the Flow2 process, the inline dilution allows all proteins washed out from the first column to be captured by the second column, regardless of the more or less harsh washing conditions in the first column. Better performance can be achieved compared to the batch process in terms of both yield and productivity.

Thus concluding, for the Flow2 process we can recommend the higher loading and washing velocities compatible with the pressure drop constraint, provided that the P_{spec} is obtained (750 cm/hr and 1500 cm/hr, respectively in this work). This is the main reason for the continuous process to exhibit better productivities than the corresponding batch operation, thus over compensating for the increased column volume utilized.

4.3 Role of buffer composition in loading and washing

In this section, the effect of the ionic strength of the loading and washing buffers is investigated. Also in this case, we analyze the effect of each one separately, by keeping the other fixed. In particular, we consider the ionic strengths of the loading

buffer, I_L ranging from 40 mM to 120 mM, and for the washing buffer higher values, with I_W ranging from 80 mM to 200 mM. The values of the remaining operating parameters are summarized in Tables 1 to 3, and in particular the ionic strength for inline dilution buffer is kept constant at $I_{LD} = 10$ mM.

4.3.1 Batch process

The Pareto fronts computed for various values of the ionic strength of the loading buffer, I_L and at fixed ionic strength of the washing buffer, I_W equal to 120 mM, are shown in Figure 6a. It is seen that the process performance improves as I_L decreases. Lower ionic strength values, in fact, facilitate the displacement of the monomer by the aggregates by enlarging the difference of adsorptivity between aggregates and monomer. Using equations 6, 7 and 8, it can be seen that the ratio of the Henry coefficients of aggregates and monomer (H_{agg}/H_{mono}) increases from about 40 to 600, as I_L decreases from 120 to 40 mM. Therefore, lower ionic strengths of the loading buffer can increase the monomer content and decrease the aggregate content in the product pool of the loading step. This comes of course in addition to the fact that using lower I_L typically implies also higher feed dilution and therefore lower protein concentration in the feed.

With respect to the washing conditions, a non-monotonous behavior is observed in Figure 6b: the process performance initially improves with increasing the wash buffer ionic strength, but then deteriorates at higher values. In particular, at $I_W = 140$ mM the Pareto becomes minute, indicating that the specified purity value can be achieved only for a restricted range of yield and productivity values. The reason is that in the batch process, the washing buffer has to be carefully tuned in order to elute the

remaining monomer inside the column without eluting also the aggregates. Too low I_w lead to too much monomer left in the column, thus decreasing yield and productivity, while too high I_w increase the aggregate leakage, thus leading to lower product purities.

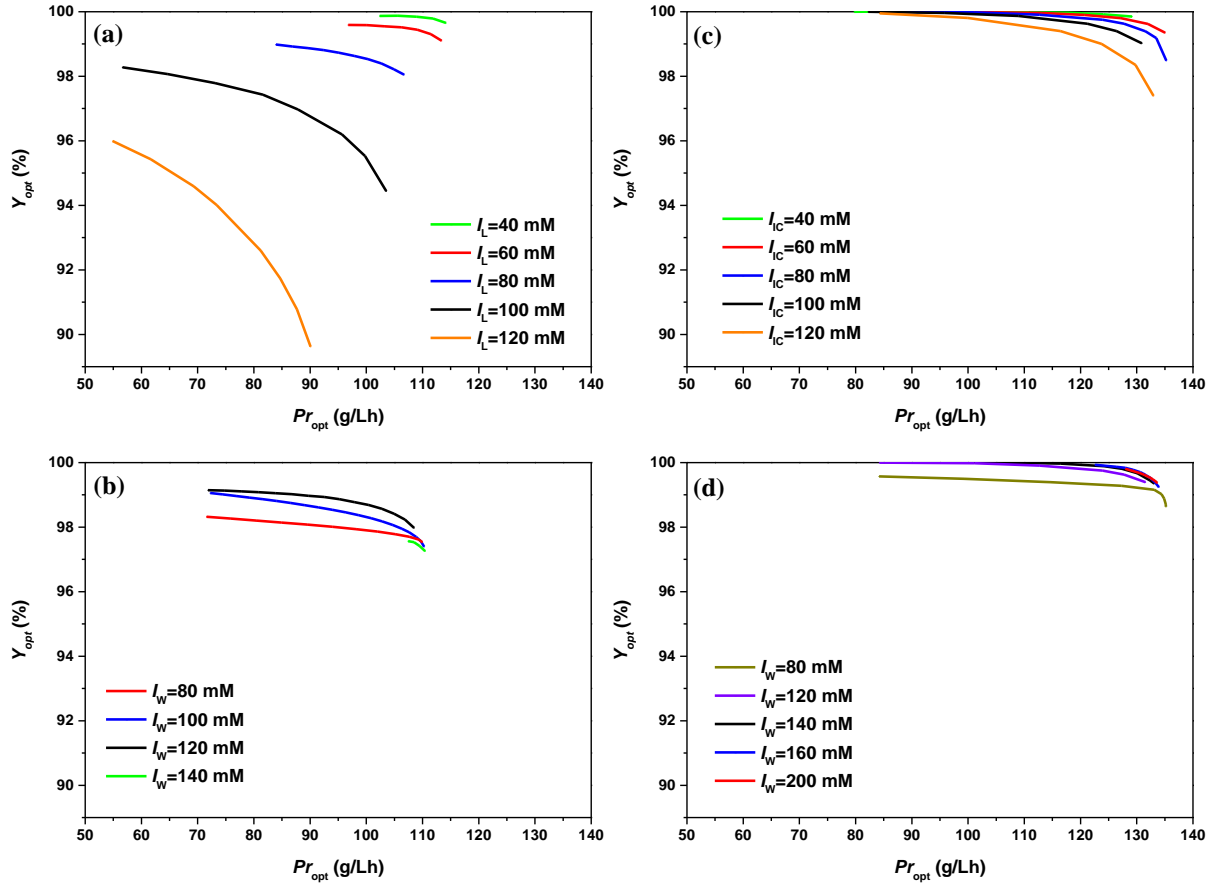


Figure 6. Tradeoff (Pareto Front) between yield and productivity of (a) Batch process for various ionic strengths of the interconnected loading buffer, I_L , with $I_w = 120$ mM. (b) Batch process for various ionic strengths of the interconnected washing buffer, I_w , with $I_L = 80$ mM. (c) Flow2 process for various ionic strengths of the interconnected loading buffer, I_{LC} , with $I_w = 120$ mM. (d) Flow2 process for various ionic strengths of the interconnected washing buffer, I_w , with $I_{LC} = 80$ mM.

The conclusion of this analysis is that, for batch processes, while for the loading

buffer lower ionic strength values (40 mM in this work) are recommended, for the washing buffer a detailed analysis is needed to identify the proper ionic strength (120 mM in this work). The latter constitutes a delicate design problem, which obviously reflects at the manufacturing level in a control problem, which may have a strong influence on the performance of the entire process. This aspect will be discussed later in terms of process robustness.

4.3.2 Continuous Flow2 process

In the case of the Flow2 process we have four buffers involved and therefore four variables to be designed and optimized: the interconnected loading buffer ionic strength, $I_{L,IC}$, the batch loading buffer ionic strength, $I_{L,B}$, the interconnected washing buffer ionic strength, $I_{W,IC}$, and the inline dilution washing buffer ionic strength, I_{ILD} . In the following, I_B is set to be equal to I_{IC} , since, for practical reasons, it is convenient to use in both steps the same feedstock, coming from the protein A eluate and corrected to this ionic strength value. In addition, the I_{ILD} is set to be fixed at 10 mM, so that the Pareto optimization can be limited only to the interconnected loading buffer ionic strength, $I_{L,IC}$ and the interconnected washing buffer ionic strength, $I_{W,IC}$. Similarly as for the batch process, we investigate the process performance first for I_{IC} ranging from 40 to 120 mM at fixed $I_W = 120$ mM, and then for I_W changing from 80 to 200 mM at fixed $I_{IC} = 80$ mM. The obtained results are shown in Figures 6c and 6d, respectively.

In Figure 6c it can be seen that, similarly to the batch process, also for the Flow2 process the performance improves, that is the Pareto Front moves to the upper right corner of the plot, as I_{IC} decreases, although to a smaller extent. On the other hand, for

I_w , the data in Figure 6d show that the process performance first improves as the washing buffer ionic strength increases, but then reaches a kind of plateau for I_w values in the order of 140 to 200 mM where the process performance remains substantially unchanged. Besides providing in general better performances than the batch process, it is remarkable that this behavior of the Flow2 process is qualitatively different from that of the batch process, which indicates a superior robustness to changes in the washing buffer. In Figure 6b it can be seen, in fact, that the batch process for the largest values of I_w , from around 100 to 140 mM, undergoes tremendous changes in its performance. This indicates that the Flow2 process can tolerate larger changes in the washing buffer ionic strength without compromising the process performance, which instead drops substantially for the batch process. This higher robustness of the Flow2 process clearly originates from the higher process flexibility, and in particular by the inline dilution, which allows for the re-adsorption in the downstream column of the impurities desorbed in the upstream column, therefore preventing product pool contamination even at strong washing conditions.

To conclude, for the Flow2 process, lower ionic strength for the loading buffer, I_c (40 mM in this work) and higher ionic strength for the washing buffer, I_w (higher than 160 mM in this work) are generally recommended to obtain best process performance in terms of both yield and productivity.

5 Process Robustness and Sensitivity Analysis

Robustness is an important factor when evaluating the performance of a process.

This is relevant at the process development stage, since higher robustness makes it easier to identify operating conditions leading to optimal process performance, but it matters also at the manufacturing scale, by facilitating the design of the controller needed to reject possible disturbances and keep the process within specifications.

In this section, we address this issue through the process sensitivity analysis^[37]. In particular, we define the normalized sensitivity, $S(\psi;\varphi)$ of the generic output process parameter, ψ to small changes of the generic input process parameter, φ as follows:

$$S(\psi;\varphi) = \frac{\partial \psi}{\partial \varphi} \left(\frac{\varphi}{\psi} \right) \quad (13)$$

The partial derivative quantifies the local change of ψ with respect to a small change of φ , *while keeping all remaining parameters constant*, and it is therefore appropriate to estimate the sensitivity of the output, ψ with respect to the input, φ . The ratio φ/ψ is introduced to make the sensitivity dimensionless, that is independent of the units used in the evaluation of the input and output process parameters. However, it should be noted that the normalized sensitivity values alone do not represent the output variability due to a given input parameter, since this is also affected directly by the intrinsic variability of the input parameter.

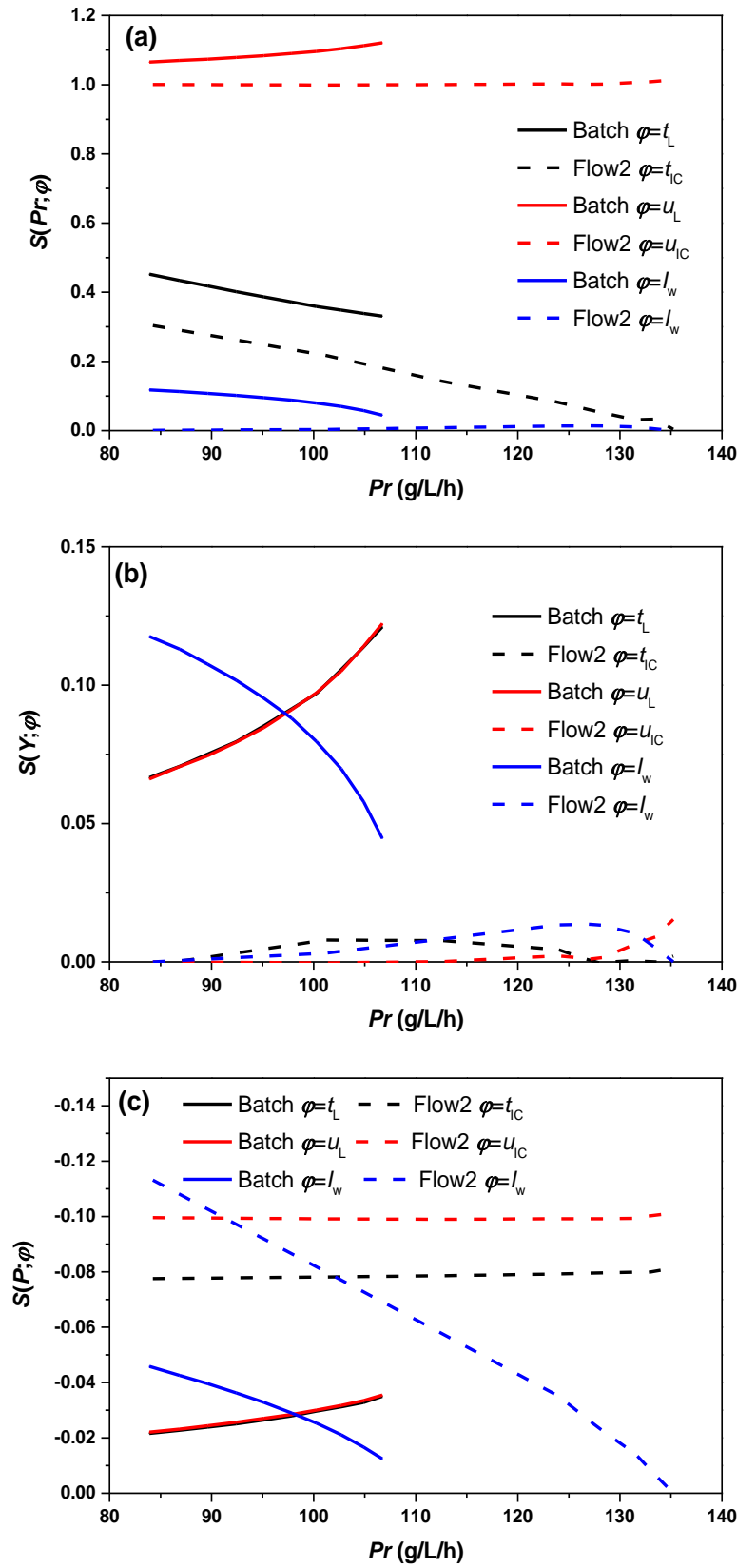
In the following, the sensitivity value is approximated through the incremental ratio $((\psi(\varphi + \Delta \varphi) - \psi(\varphi))/\Delta \varphi)$, where the numerator represents the change in the output parameter corresponding to an imposed change of the input parameter, equal to +1% of the reference conditions considered in the sensitivity analysis. In particular, we consider, as output process parameters, the three parameters determining the process performance: productivity, yield and purity, that is $\psi = Pr, Y$ and P , respectively. As

process input parameters we consider the loading time, the loading velocity and the washing buffer ionic strength, that is $\varphi = t_L, u_L$ and I_w , respectively.

As reference conditions for the sensitivity analysis we select the optimised operating conditions along the Pareto fronts with $P_{\text{spec}} = 99\%$ shown in Figure 3 for the batch and Flow2 process, respectively. Accordingly, for the various operating conditions along the Pareto Front, we compute, for each given change in the input parameter, the corresponding changes in the three performance parameters above, and from these the corresponding sensitivity values through equation (13). The obtained results, shown in Figure 7a to 7c, indicate that productivity and yield exhibit a *positive* sensitivity value, while the the purity sensitivity is *negative*. This is consistent with the fact that each of the considered operating conditions belong to the Pareto front at a fixed purity value equal to 99%. The perturbed operating parameters, in fact, although leading to improvements in productivity and yield (positive sensitivity value), lead to a lower, not acceptable, purity value (negative sensitivity value) and therefore cannot be part of the Pareto Front. In the same Figure 7 it is also seen that the Flow 2 process, which can achieve larger productivity values because of its higher efficiency, also exhibits a generally better sensitivity behavior. More specifically, the very high sensitivity of the yield with respect to the washing buffer ionic strength, I_w exhibited by the batch process is largely removed in the Flow2 process.

In particular, Figure 7a shows that productivity is the most sensitive of all considered output parameters, and the corresponding sensitivity values are substantially equivalent for the two processes. Significantly lower sensitivity of the Flow2 process

is observed only for the washing buffer ionic strength, I_w and for the larger productivity values. The strong reduction of the yield sensitivity with respect to I_w , mentioned above, is illustrated in Figure 7b: The sensitivity values for the Flow2 process are less than 0.015, compared to values about ten times larger, in the order of 0.1, for the batch process. This is a consequence of the stabilizing effect of the inline dilution process discussed above in the context of Figures 6b and 6d. It is worth noting that, in Figure 7b, the yield sensitivities with respect to t_L and u_L for the batch process are almost coincident. This is because these two parameters have the same effect on the loading amount, which is in fact defined by their product, $(t_L * u_L)$, which in turns leads to the same change in the yield value. The same observation applies to the purity sensitivities, as it can be seen in Figure 7c. On the other hand, in Figure 7c, the batch process shows lower purity sensitivities than the Flow2 process, with respect to any of the input process parameters considered. However, the sensitivities with respect to the loading time and velocity are less critical in the context of process control (switching times and flow rates are typically well controlled), and the observed differences are lower than in the case above. On the other hand, the most critical sensitivity, with respect to I_w , can be substantially decreased when operating at high productivity values.



550

551 *Figure 7. Normalized sensitivity of productivity (a), yield (b) and purity (b) with respect*

to t_L , u_L and I_W for the batch process and to t_{IC} , u_{IC} and I_W for the Flow2 process, along the corresponding Pareto fronts shown in Figure 3.

6 Conclusion

Frontal chromatography has been analyzed with respect to two alternative implementations: a single column, batch process and a two-column, continuous process, referred to as Flow2. In particular, its application in the polishing step of the processes for mAb manufacturing, where the immunogenic high molecular weight species are removed from the target protein, is investigated with reference to a case of industrial relevance. An ad-hoc procedure for process optimization has been developed and validated through a general multiobjective optimization procedure, in terms of the Pareto Front of yield and productivity at a given purity, for both processes. The derived procedure allowed a thorough comparison of the two processes at their corresponding optimal operating conditions and not only in terms of the performance parameters, such as yield, purity and productivity, but also with respect to process robustness through a proper local sensitivity analysis.

The obtained results indicate that, although column interconnection puts constraints on the linear velocities of the Flow2 process, this in general achieves better performance in terms of both yield and productivity at a given purity, compared to the batch process. An important component of this improvement is the higher flexibility provided by the inline dilution process, which allows the Flow2 to better tolerate higher, and therefore more advantageous, ionic strength values in the washing step. The

comparison of the normalized sensitivities computed for the two processes shows that the Flow2 process exhibits much smaller yield sensitivities than the batch one. Even more importantly, it is shown that for the Flow2 process, optimal operating conditions can be found, where the process performance is not very sensitive to the washing conditions and in particular to the buffer ionic strength. This is a very important result, since this high sensitivity represents one of the largest drawbacks of the frontal chromatography batch process, which is the reason for its lack of robustness and reflects in difficulties in determining washing conditions that can guarantee reliable operation particularly at the large scale.

The discussed robustness analysis is based on the study of the *local* sensitivity values defined by equation (13). These local values are relevant, for example, for determining the characteristics of the algorithm used to control a specific input variable, e.g. determine the largest deviations that can be allowed to the input variable. It is interesting to note that the above reached conclusion is in good agreement with the analysis reported by Vogg et al. ^[18] based on a *global* definition of robustness. In particular, they compared the robustness of the two processes by considering the amplitude of the range of operating conditions, which allow respecting specific requirements in terms of process performance parameters. While their results indicate that the Flow2 process is characterized by larger robust design spaces than the batch process, our results augment this behavior by showing decreased local sensitivity of the Flow2 process towards process parameter variations at given sets of process parameters.

596 **Notations**

597	d_p	Particle diameter (m)
598	$d_{ax,i}$	Axial dispersion coefficient of i component (m ² /s)
599	H_i	Henry coefficient (atm*m ² /mg)
600	I_B	Ionic strength of Batch loading buffer of Flow2 process (mM)
601	I_C	Ionic strength of Batch and Flow2 interconnected loading
602	buffer (mM)	
603	I_L	Ionic strength of Batch and Flow2 loading buffer (mM)
604	I_W	Ionic strength of Batch and Flow2 washing buffer (mM)
605	I_{LD}	Ionic strength of Flow2 inline dilution buffer (mM)
606	$k_{m,i}$	Mass transfer coefficient of i component (m ² /s)
607	L_{col}	Column length (m)
608	Pr	Process Productivity (g/Lh)
609	P_i	Purity for i component
610	P_{spec}	Specification value for purity
611	u_L	Batch loading linear velocity (mL/min)
612	u_w	Batch washing linear velocity (mL/min)
613	$u_{L,IC}$	Flow2 interconnected loading linear velocity (cm/hr)
614	$u_{W,IC}$	Flow2 interconnected washing linear velocity (cm/hr)
615	u_{ILD}	Flow2 inline dilution linear velocity (cm/hr)
616	$u_{L,B}$	Flow2 disconnected loading linear velocity (cm/hr)
617	q_i	Concentration of i component in solid phase (mg/mL)

618	q_i^{eq}	Equilibrium concentration of i component in solid phase
619	(mg/mL)	
620	q_i^{sat}	Saturation binding capacity of i component in solid phase
621	(mg/mL)	
622	t_L	Batch loading time (min)
623	t_w	Batch washing time (min)
624	t_{RR}	R-R time for batch and Flow2 process (min)
625	$t_{L,IC}$	Flow2 interconnected loading time (min)
626	u_{sf}	Superficial velocity (m/s)
627	Y_i	Yield for i component
628	$\varepsilon_{t,i}$	Total accessible porosity of I component
629	μ	Mobile phase dynamic viscosity (1.0 mPas)

630

631 **References:**

- 632 [1] H. L. Levine, J. E. Lilja, R. Stock, H. Hummel, S. D. Jones, *BioProcess Int* **2012**, 10, 20.
- 633 [2] D. M. Ecker, S. D. Jones, H. L. Levine, *MAbs*, **2015**, 9.
- 634 [3] S. Singh, N. K. Tank, P. Dwiwedi, J. Charan, R. Kaur, P. Sidhu, V. K. Chugh, *Current clinical*
- 635 *pharmacology* **2018**, 13, 85.
- 636 [4] H. Kaplon, J. M. Reichert, *MAbs*, Taylor & Francis **2019**, 219.
- 637 [5] A. L. Grilo, A. Mantalaris, *Trends Biotechnol.* **2019**, 37, 9.
- 638 [6] S. S. Farid, J. Washbrook, N. J. Titchener Hooker, *Biotechnol. Progr.* **2005**, 21, 486.
- 639 [7] C. A. Challener, *Biopharm Int.* **2014**, 27, 20.
- 640 [8] S. Vogg, T. Muller-Spath, M. Morbidelli, *Current opinion in chemical engineering* **2018**, 22, 138
- 641 [9] F. Feidl, S. Vogg, M. Wolf, M. Podobnik, C. Ruggeri, N. Ulmer, R. Wälchli, J. Souquet, H. Broly,
- 642 A. Butté, *Biotechnol. Bioeng.* 2020, 117, 1367.
- 643 [10] D. J. Karst, F. Steinebach, M. Morbidelli, *Curr. Opin. Biotech.* 2018, 53, 76.
- 644 [11] FDA, Modernizing the way drugs are made: a transition to continuous manufacturing , **2017**.
- 645 [12] FDA, Quality considerations for continuous manufacturing-guidance for industry **2017**.
- 646 [13] G. Carta, A. Jungbauer, *Protein chromatography: process development and scale-up*, John Wiley
- 647 & Sons, **2020**.
- 648 [14] T. Ichihara, T. Ito, Y. Kurisu, K. Galipeau, C. Gillespie, *MAbs*, Taylor & Francis **2018**, 325.

649 [15] H. F. Liu, B. McCooey, T. Duarte, D. E. Myers, T. Hudson, A. Amanullah, R. van Reis, B. D. Kelley,
650 *J. Chromatogr. A* **2011**, 1218, 6943.

651 [16] D. Pfister, L. Nicoud, M. Morbidelli, *Continuous Biopharmaceutical Processes: Chromatography,*
652 *Bioconjugation, and Protein Stability*, Cambridge University Press **2018**.

653 [17] M. T. Stone, K. A. Cotoni, J. L. Stoner, *J. Chromatogr. A* **2019**, 1599, 152.

654 [18] S. Vogg, T. Müller-Späth, M. Morbidelli, *J. Chromatogr. A* **2020**, 460943.

655 [19] N. J. Titchener Hooker, P. Dunnill, M. Hoare, *Biotechnol. Bioeng.* **2008**, 100, 473.

656 [20] S. L. C. Ferreira, R. E. Bruns, E. G. P. Da Silva, W. N. L. Dos Santos, C. M. Quintella, J. M. David,
657 J. B. de Andrade, M. C. Breitzkreitz, I. C. S. F. Jardim, B. B. Neto, *J. Chromatogr. A* **2007**, 1158, 2.

658 [21] C. Shi, Z. Gao, Q. Zhang, S. Yao, N. K. Slater, D. Lin, *J. Chromatogr. A* **2020**, 460936.

659 [22] N. Andersson, H. Knutson, M. Max-Hansen, N. Borg, B. Nilsson, *Ind. Eng. Chem. Res.* **2014**, 53,
660 16485.

661 [23] T. Müller-Späth, G. Ströhlein, L. Aumann, H. Kornmann, P. Valax, L. Delegrange, E. Charbaut, G.
662 Baer, A. Lamproye, M. Jöhnck, *J. Chromatogr. A* **2011**, 1218, 5195.

663 [24] D. Baur, M. Angarita, T. Müller Späth, M. Morbidelli, *Biotechnol. J.* **2016**, 11, 135.

664 [25] S. M. Pirrung, M. Ottens, *Preparative Chromatography for Separation of Proteins* **2017**, 269.

665 [26] T. Müller Späth, M. Krättli, L. Aumann, G. Ströhlein, M. Morbidelli, *Biotechnol. Bioeng.* **2010**, 107,
666 652.

667 [27] J. Siitonen, M. Mänttari, A. Seidel-Morgenstern, T. Sainio, *J. Chromatogr. A* **2015**, 1391, 31.

668 [28] G. Guiochon, A. Felinger, D. G. Shirazi, *Fundamentals of preparative and nonlinear*
669 *chromatography*, Elsevier, **2006**.

670 [29] G. Carta, A. Jungbauer, Wiley-VCH Verlag GmbH, **2010**.

671 [30] B. Guélat, R. Khalaf, M. Lattuada, M. Costioli, M. Morbidelli, *J. Chromatogr. A* **2016**, 1447, 82.

672 [31] G. Zames, N. M. Ajlouni, N. M. Ajlouni, N. M. Ajlouni, J. H. Holland, W. D. Hills, D. E. Goldberg,
673 *Information Technology Journal* **1981**, 3, 301.

674 [32] <https://it.mathworks.com/help/gads/gamultiobj.html>, **2020**

675 [33] J. P. Abulencia, L. Theodore, *Fluid flow for the practicing chemical engineer*, Wiley Online Library,
676 **2009**.

677 [34] [https://www.merckmillipore.com/Web-CH-site/de_DE/-/CHF/ShowDocument-](https://www.merckmillipore.com/Web-CH-site/de_DE/-/CHF/ShowDocument-Pronet?id=201806.078)
678 [Pronet?id=201806.078](https://www.merckmillipore.com/Web-CH-site/de_DE/-/CHF/ShowDocument-Pronet?id=201806.078), **2018**

679 [35] D. Farnan, D. D. Frey, C. Horváth, *Biotechnol. Progr.* **1997**, 13, 429.

680 [36] G. Gotmar, T. Fornstedt, G. Guiochon, *J. Chromatogr. A* **1999**, 831, 17.

681 [37] A. Varma, M. Morbidelli, *Parametric sensitivity in chemical systems*, Cambridge University Press,
682 **2005**.

683 [38] D. J. Downing, R. H. Gardner, F. O. Hoffman, *Technometrics* **1985**, 27, 151.

684 [39] T. J. Krieger, C. Durston, D. C. Albright, *Transactions of the American Nuclear Society* **1978**, 28.