

Allergy – Letter to the Editor

Enzymatic activity of ACE2 regulates type 2 airway inflammation in mice

To the Editor,

Coronavirus disease 2019 (COVID-19), caused by the novel severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2), has led to a global pandemic. SARS-CoV-2 spike protein binds to the angiotensin-converting enzyme 2 (ACE2), a transmembrane endopeptidase on the surface of the airway epithelium, on host cells for invasion and infection; therefore, most current COVID-19 research has focused on ACE2. Patients with chronic obstructive pulmonary disease and idiopathic pulmonary fibrosis are reportedly at high risk of COVID-19 morbidity and mortality,^{1,2} whereas those with asthma are not.³ Asthma is a heterogeneous disease triggered by environmental factors such as house dust mites (HDM) and viruses that is typically characterized by chronic airway inflammation.⁴ Exposure to these factors promotes epithelial cell damage, leading to the release of cytokines that will provoke a type 2 inflammatory response.⁵ Kimura et al.⁶ reported that interleukin (IL)-13 exposure reduces ACE2 expression in airway epithelium of patients with asthma, whereas interferons enhance ACE2 expression.² Altogether, ACE2 low expression in epithelial cells might protect asthma patients from COVID-19. However, the relationship between asthma-related allergic inflammation and ACE2 expression in the airway has not yet been assessed *in vivo*.

To explore these associations, we used an HDM-induced asthma mouse model. C57BL/6J mice were intratracheally exposed or not to HDM (4–15 mice/group) at days 0, 7, and 14. Bronchoalveolar lavage fluid (BALF) and lung tissue samples were harvested at day 3, 10, and 17 to investigate eosinophil numbers and *Ace2* expression (Figure 1A).

The eosinophil counts in BALF samples increased from day 3 to day 17 concomitantly with repeated HDM exposure in a step-wise fashion (Figure 1B; control vs day 17 [$p < 0.0001$], day 10 vs day 17 [$p = 0.0028$]), whereas lung tissue *Ace2* expression decreased (Figure 1C; control vs day 17 [$p = 0.0002$], control vs day 10 [$p = 0.0033$], day 3 vs day 10 [$p = 0.0066$]). These findings agree with Jackson et al. data⁷, showing that *ACE2* was significantly reduced in human bronchial epithelial brush samples after a segmental bronchial allergen challenge. *ACE2* activity was also significantly suppressed by HDM injection (Figure 1D, $p < 0.0001$). These results indicate that *ACE2* expression and/or function is associated with the pathogenesis of allergic airway inflammation.

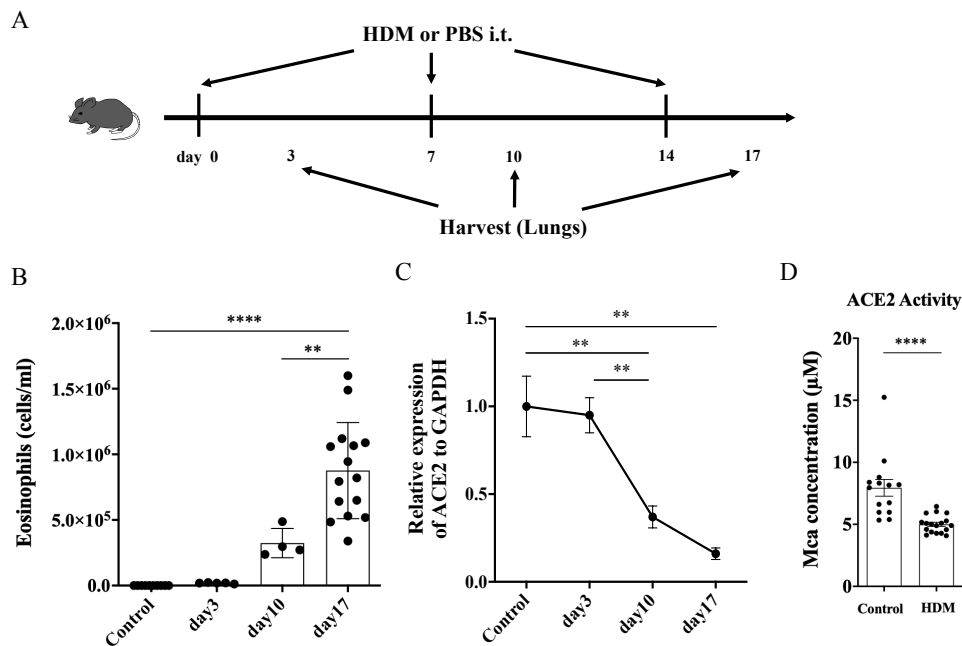


FIGURE 1. Eosinophils counts in the bronchoalveolar fluid (BALF) and *Ace2* expression in lung tissues from house dust mites (HDM)-exposed mice. (A) Experimental scheme. (B) Number of eosinophils in bronchoalveolar lavage fluid (BALF) samples. (C) Relative *Ace2* expression in lung tissues analyzed by quantitative real-time polymerase chain reaction. (D) *ACE2* enzymatic activity in BALF was measured on day 14 after 6 h (14–18 mice/group from three independent experiments). Results are shown as mean ± SD (B) or SEM (C). Significance was determined by one-way ANOVA and Tukey's multiple comparisons test. ** $p < 0.01$ and **** $p < 0.0001$.

Additionally to its important role as a receptor for SARS-CoV-2, *ACE2* is the main

enzyme for catalyzing angiotensin II into angiotensin¹⁻⁷ and has an anti-inflammatory effect in cardiovascular diseases.⁸ Therefore, we hypothesized that ACE2 activity could modulate the type 2 inflammation of asthma. To test this hypothesis, we injected HDM-induced asthma mice with diminazene aceturate (DIZE), an activator of ACE2,⁹ and examined its impact on the airway inflammatory process (Figure 2A).

DIZE treatment had no impact on *Ace2* expression (Figure 2B), whereas its activity was significantly elevated (Figure 2C, $p = 0.0148$). DIZE also significantly prevented the eosinophilia induced by HDM exposure (Figure 2D, $p = 0.0003$). Hematoxylin and eosin (HE), and periodic acid-Schiff (PAS) staining showed that DIZE markedly inhibited the HDM-induced infiltration of inflammatory cells and goblet cell hyperplasia (Figure 2E; HE [$p = 0.0002$], PAS [$p < 0.0001$]). DIZE treatment also suppressed the airway hyperreactivity in response to methacholine stimulation (Figure 2F, $p = 0.0483$). Furthermore, DIZE treatment resulted in significantly lower levels of IL-5 and IL-13 in the BALF and of IL-33 in the lungs, whereas IL-4 levels remained unaltered in the BALF (Figure 2G; IL-5 [$p = 0.0108$], IL-13 [$p = 0.0462$], IL-33 [$p < 0.0001$]). Overall, these results suggest that ACE2 activity, but not its expression, could regulate type 2 allergic airway inflammation in asthma. Interestingly, reduced IL-13 levels in BALF had no impact on *Ace2* expression in the lungs of this asthma model.

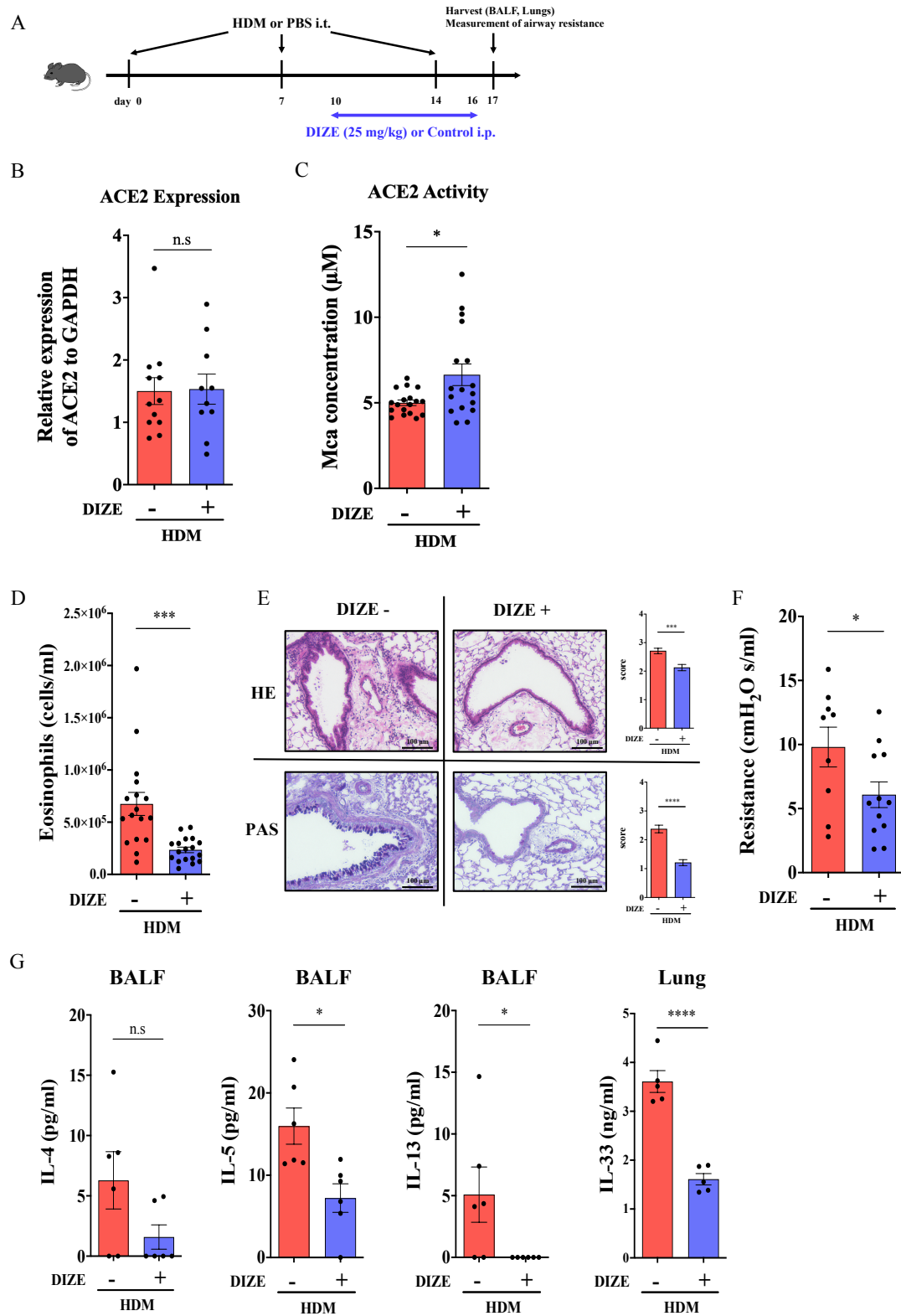


FIGURE 2. ACE2 activation by diminazene aceturate (DIZE) reduces type 2 inflammation in house dust mites (HDM)-exposed mice. (A) Experimental scheme. (B), *Ace2* expression in the lung determined by quantitative reverse-transcription polymerase chain reaction (10–12 mice/group from two independent experiments). (C) ACE2 enzymatic activity in bronchoalveolar lavage fluid (BALF) was measured on day 14 after 6 h (14–18 mice/group from three independent experiments). (D) Eosinophils count in BALF (5–6 mice/group from three independent

experiments). (E) Hematoxylin and eosin, and periodic acid-Schiff staining in the lung were blindly scored by five individuals on a scale of 0–3 for peribronchial inflammation (6 mice/group). (F) Bronchial hyperresponsiveness to 50 mg/mL methacholine (4–6 mice/group from two independent experiments). (G) IL-4, IL-5, and IL-13 protein levels in BALF and IL-33 in lung tissue was measured by enzyme-linked immunosorbent assay (5–6 mice/group). Results are shown as mean \pm SEM (B–G). Significance was determined by One-way ANOVA and Bonferroni (B,C) and Student's *t*-test (D–G). **p*<0.05, ***p*<0.01, ****p*<0.001, and *****p*<0.0001.

IL-33 is released by the lung epithelia and evokes type 2 cytokine production by group 2 innate lymphoid cells (ILC2).⁵ Since DIZE treatment significantly reduced the IL-33 concentration in the lung, DIZE may inhibit IL-33 production by lung epithelial cells, leading to the suppression of ILC2-mediated cytokine release.

It has been speculated that suppression of IL-13 by asthma treatment may cancel out any resistance against SARS-CoV-2 infection due to the reduction of ACE2 levels. Our findings suggest that potentiation of ACE2 activity can prevent asthma-associated type 2 allergic airway inflammation without affecting ACE2 expression in the lungs. This effect offers new possibilities to treat asthma patients infected by SARS-CoV-2. Although tissues from type 2 cytokine-high allergic patients reportedly showed significantly lower ACE2 expression, which were inversely correlated with the Th2 cytokine levels and signature molecule expression,⁶ further investigations are required to clarify whether reduction of IL-13 in the asthmatic airway after treatment of asthma directly affects ACE2 expression in the airway epithelia.

References

1. George PM, Wells AU, Jenkins RG. Pulmonary fibrosis and COVID-19: the potential role for antifibrotic therapy. *Lancet Respir Med* 2020;8:807-815. doi:10.1016/S2213-2600(20)30225-3
2. Hoffmann M, Kleine-Weber H, Schroeder S, et al. SARS-CoV-2 cell entry depends on ACE2 and TMPRSS2 and is blocked by a clinically proven protease inhibitor. *Cell* 2020;181:271-280.e8. doi:10.1016/j.cell.2020.02.052
3. Matsumoto K, Saito H. Does asthma affect morbidity or severity of Covid-19? *J Allergy Clin*

Immunol 2020;146:55-57. doi:10.1016/j.jaci.2020.05.017

4. Pavord ID, Beasley R, Agusti A, et al. After asthma: redefining airways diseases. *Lancet* 2018;391:350-400. doi:10.1016/S0140-6736(17)30879-6
5. Akdis CA, Arkwright PD, Brüggemann MC, et al. Type 2 immunity in the skin and lungs. *Allergy* 2020;75:1582-1605. doi:10.1111/all.14318
6. Kimura H, Francisco D, Conway M, et al. Type 2 inflammation modulates ACE2 and TMPRSS2 in airway epithelial cells. *J Allergy Clin Immunol* 2020;146:80-88.e8. doi:10.1016/j.jaci.2020.05.004
7. Jackson DJ, Busse WW, Bacharier LB, et al. Association of respiratory allergy, asthma, and expression of the SARS-CoV-2 receptor ACE2. *J Allergy Clin Immunol* 2020;146:203-206.e3. doi:10.1016/j.jaci.2020.04.009
8. Santos RAS, Oudit GY, Verano-Braga T, Canta G, Steckelings UM, Bader M. The renin-angiotensin system: going beyond the classical paradigms. *Am J Physiol Heart Circ Physiol* 2019;316:H958-970. doi:10.1152/ajpheart.00723.2018
9. Rajapaksha IG, Mak KY, Huang P, Burrell LM, Angus PW, Herath CB. The small molecule drug diminazene aceturate inhibits liver injury and biliary fibrosis in mice. *Sci Rep* 2018;8:10175 doi:10.1038/s41598-018-28490-y

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