

Title: Senescence: Still an Unsolved Problem of Biology

A manuscript for consideration as a *Reviews and Syntheses piece* in *Ecology Letters*.

Authors: Mark Roper*, Pol Capdevila & Roberto Salguero-Gómez

Author Affiliations:

Department of Zoology, University of Oxford, 11a Mansfield Road, Oxford, OX1 3SZ, UK.

Mark Roper, Pol Capdevila & Roberto Salguero-Gómez

Centre for Biodiversity and Conservation Science, University of Queensland, St Lucia 4071 QLD, Australia.

Roberto Salguero-Gómez

Evolutionary Demography laboratory, Max Plank Institute for Demographic Research, Rostock 18057, Germany.

Roberto Salguero-Gómez

Contributions

M.R. and R.S.G. conceived the project. M.R. and P.C. conducted the analyses with input from R.S.-G. and produced all visualisations. M.R. drafted the first version and, together with P.C. and R.S.-G., revised and edited the manuscript.

Competing interests

The authors declare no competing interests.

Corresponding Author:

Mark Roper

Department of Zoology, University of Oxford, 11a Mansfield Road, Oxford, OX1 3SZ, UK.

07715877987|mark.roper@keble.ox.ac.uk

Keywords: ageing, comparative demography, life history, mortality, reproduction

Manuscript contents:

Abstract (150 words), Main text (3957 words), 44 references, 4 figures, 1 text box. Supplementary Info (separate file) contains R code, 3 figures, and 3 tables.

Data accessibility statement

Data are available from the COMPADRE Plant Matrix Database and COMADRE Animal Matrix Database (www.compadre-db.com). Code used for analysis is available in the supplementary information. Should the manuscript be accepted, the data supporting the results will be archived in an appropriate public repository (Dryad, Figshare or Hal) and the data DOI will be included at the end of the article.

Abstract

Despite *ca.* seven decades of theoretical elaboration since Peter Medawar's foundational 'An Unsolved Problem of Biology', the fundamental problem of the evolution of senescence, *i.e.* the increasing risk of mortality and decline in reproduction with age after maturity, remains unsolved. Theories of senescence predict the inescapability of senescence, or its universality among species with a clear germ-soma barrier. Here, using demographic information for 475 multicellular species, we exemplify the discrepancy between these theoretical predictions and empirical data. We derive age-based trajectories of mortality and reproduction whose form cannot be satisfactorily explained by the expectation of universal senescence, and show that species' may often display senescence for one fitness component but not the other. We propose that theories of senescence must be extended beyond merely individual chronological age; size, the species' ecological context, and kin selection may all play currently hidden, yet integral roles in shaping patterns of senescence.

Main text

Introduction

The evolution of senescence, the increasing risk of mortality and decline in reproduction with age after maturity, has long been explained by a collation of theories defining the ‘classical evolutionary framework of ageing’. The central logic common to these theories argues that the force of natural selection weakens with age (Medawar 1952; Williams 1957; Hamilton 1966; Kirkwood 1977). Selection becomes too weak to oppose the accumulation of genes that negatively affect older age classes (Medawar 1952), or favours these genes if they also have beneficial effects at earlier ages in life (Williams 1957), when the contribution individuals make to future populations, *i.e.* reproductive value (Fisher 1930), is assumed to be greater. Selection should therefore favour resource investment into earlier reproduction rather than late-life maintenance (Kirkwood 1977). Ultimately, these theories predict, directly (Hamilton 1966) or indirectly (Medawar 1952; Williams 1957), that senescence is inescapable (Hamilton 1966), or at least inevitable in organisms with a clear germline-soma separation (Williams 1957; Kirkwood 1977).

Emerging empirical data has thrown a challenge to the classical evolutionary framework of ageing (see e.g. Baudisch *et al.* 2013; Jones *et al.* 2014). A recent comparative depiction of demographic ageing patterns across 46 species of animals, plants, and algae (Jones *et al.* 2014) has contradicted the expectations of the classical evolutionary framework. Many of the examined species display negligible (Finch 1994) or even negative (Vaupel *et al.* 2004) senescence, where the risk of mortality remains constant or decreases with age, and reproduction remains constant or increases with age. This mismatch between expectations and observations limits the predictive power of the classical evolutionary framework to explain the diversity of senescence across the tree of life. We now need to understand why some species succumb to senescence, and what allows others to escape its forces. What are the mechanisms

behind such variation (Baudisch & Vaupel 2012; Jones & Vaupel 2017), and how prevalent are such “exceptions” are to the assumed rule of universal senescence?

With ever growing amounts of readily available longitudinal demographic datasets (e.g. Salguero-Gómez *et al.* 2015; Salguero-Gómez *et al.* 2016), comparative demography offers a tool to begin to unlock the key answers to this question. Here, we utilise high-resolution demographic information for wild populations (Box 1) of 80 animal and 395 plant species worldwide (See Materials and methods) to (i) provide a quantitative evaluation of the rates of actuarial senescence – the change in mortality risk with age after maturation – across multicellular organisms, (ii) test whether the classical evolutionary framework explains the examined diversity of senescence rates, with special attention to predictions from germ-soma separation, and (iii) propose how to widen the classical evolutionary framework of ageing to better encompass the study of senescence across the tree of life.

Briefly, we first derived life tables (Chiang 1984) from a selection of species’ matrix population models (Caswell 2001), each of which summarise the population dynamics of the studied species under natural conditions (See Box 1 and Materials and methods). We only considered the adult part of the life table in our studies of senescence, *i.e.* from the age of maturity onwards, as this is when senescence is predicted to start (Williams 1957; Hamilton 1966). We then quantified the rate of actuarial senescence on the survivorship trajectory of each species’ life table using a ‘shape’ metric of senescence (Baudisch & Stott 2019). Our analysis uses a ‘pace-shape’ framework of ageing (Keyfitz 1977; Baudisch 2011), where the pace of ageing quantifies the speed of life via mean life expectancy (Baudisch 2011). The shape of ageing (*i.e.* senescence) quantifies the spread and timing of mortality events, normalised by mean life expectancy, which facilitates cross-species comparison. The shape metric, S , is bound between -0.5 and 0.5 (See Materials and Methods), where $S > 0$ indicates that most mortality events occur at advanced ages (*i.e.* actuarial senescence), while $S < 0$ indicates low

mortality late in life, *i.e.* escape from actuarial senescence. We determined a bound around zero using a root mean square distance measure (See Materials and Methods), with values of S that fall within the bound deemed to be indifferent from zero. We therefore describe species with such values as displaying *negligible* actuarial senescence.

Previous studies have suggested that phylogenetic relatedness may play a role in determining whether a given species displays positive, negligible or negative actuarial senescence (Jones *et al.* 2009; Jones *et al.* 2014). Here, we quantify the role of evolutionary history on actuarial senescence across our 475 species by estimating its phylogenetic signal (Pagel 1999) using phylogenies for animals (Sánchez-Reyes & O'Meara 2019) and plants (Jin and Qian 2019) respectively. Finally, the central assumption of the classical evolutionary framework of ageing, that the force of natural selection weakens with age, rests on the assumption that older individuals contribute less to future populations. This is both because the theories assume fewer individuals survive to later age classes (Medawar 1952), and that individuals are expected to favour reproduction at young rather than old ages (Williams 1957; Hamilton 1966; Kirkwood 1977). To observe how different age classes contribute to future populations in our study species, we use the derived life tables (Chiang 1984) to quantify age-specific reproduction rates ($m(x)$) to see if they match the pattern of actuarial senescence already quantified (See Material and methods).

Material and Methods

Data

We used the COMADRE Animal Matrix Database (v. 3.0.0) (Salguero-Gómez *et al.* 2016) and COMPADRE Plant Matrix Database (v. 5.0.0) (Salguero-Gómez *et al.* 2015) to obtain age trajectories of survival and reproduction. These open-access data repositories consist of a collection matrix population models (MPMs) (Caswell 2001) incorporating high-resolution

demographic information on the survival and reproduction patterns of over 1,000 animal and plant species worldwide and associated metadata (Salguero-Gómez *et al.* 2015; Salguero-Gómez *et al.* 2016). Both databases include information on species for which the data have been digitised and thoroughly error-checked. We imposed a series of selection criteria to restrict our analyses to data of the highest quality possible.

- (i) MPMs were parameterised with field data from non-disturbed, unmanipulated populations (*i.e.* natural populations) to best describe the species' age trajectories.
- (ii) MPMs had dimension $\geq 3 \times 3$ (*i.e.* rows \times columns). Generally, low dimensions MPMs lack quality for the estimation of life history traits (Salguero-Gómez & Plotkin 2010). This selection criterion also helps avoid problems with quick convergence to stationary equilibrium, at which point the estimates of life history trait values and rates of senescence become unreliable (Jones *et al.* 2014; Horvitz & Tuljapurkar 2009).
- (iii) MPMs were only used when the entire life cycle was explicitly modelled including recordings of survival, development, and reproduction for all life cycle stages.
- (iv) When multiple studies existed for the same species, we considered only the study of greater duration to ensure the highest temporal variation in the population dynamics was captured.
- (v) Studies of annual plant species modelled using seasonal projection matrices were not included; we chose only species using an annual time step. This is due to the difficulties of converting their population dynamics to an annual basis to compare with all other species' models.
- (vi) Included MPMs have stage-specific survival values ≤ 1 . In a small number of published models, the stage-specific survival values can exceed 1 due to clonality

being hidden in the matrix, rounding errors, or other mistakes in the original model
(Salguero-Gómez *et al.* 2015; Salguero-Gómez *et al.* 2016).

- (vii) MPMs were from species of which phylogenetic data was available, to ensure we
were able to account for phylogenetic relatedness on our models.

The result of these criteria was a subset of 475 species of animals and plants from the initial
databases, which we used for our analysis. Of these, 80 were animals, with 15 invertebrates
and 65 vertebrates. The remaining 395 species were plants, with 25 gymnosperms and 370
angiosperms. We provide a list of all the species used, their categorisation of senescence
including a value of S , and their relevant source study in the supplementary information (Table
S1).

Quantifying actuarial senescence

MPMs are a summary of the population dynamics of a given species, from which we can
calculate several life history traits. To do so, we first must decompose an MPM (A) into its
sub-components (Caswell 2001):

U – containing the stage-specific survival rates

F – containing the stage-specific per-capita reproduction rates

C – containing stage-specific per-capita clonality rates

$$A = U + F + C \quad \text{equation 1}$$

This decomposition facilitates the estimation of key life history traits, including a rate
of senescence (S) (Baudisch & Stott 2019). Calculating S requires first obtaining the age-
specific survivorship curve $l(x)$ from U . To obtain $l(x)$ we first have to define age, and the
definition of age requires a choice of a stage that corresponds to “birth”. Following Jones *et al.*
(2014), we defined the stage corresponding to birth as the first established non-propagule
stage (e.g., not seeds or seed bank in the case of plants, nor larvae or propagules in animals)

due to the estimate uncertainty of parameters involved in those stages. The calculation of $l(x)$ was then implemented according to Caswell (p. 118-21) (2001).

$$l(x) = e^t U^x e^j \quad x = 0, 1, \dots \quad \text{equation 2}$$

Where e is a vector of ones, and we start with a single individual in the stage j defined to correspond to birth.

We considered survivorship trajectories beginning at the age of maturity (α - calculated following 5.47–5.54 in Caswell (2001)) and ending at the age at which 5% survivorship from maturity occurs (ω). This is because a cohort modelled by iteration of the U matrix eventually decays exponentially at a rate given by the dominant eigenvalue of U , and converges to a quasi-stationary distribution given by the corresponding right eigenvector w . Once this convergence has happened, mortality remains constant with age, and so to prevent our conclusions being overly influenced by this assumption, we calculated the age at which the cohort had converged to within a specified percentage (5%) of the quasi-stationary distribution (Jones *et al.* 2014, Horvitz & Tuljapurkar 2009).

Following Baudisch & Stott (2019), the function $H(x)$ defines the cumulative hazard of mortality up to age x as

$$H(x) = \int_{\alpha}^x \mu(t) dt \quad \text{equation 3}$$

Where $\mu(x)$ denotes the age-specific mortality function capturing the average hazard of death of an individual at age x , and $H(x)$ corresponds to the logarithmic transformation of the survivorship trajectory ($H(x) = -\ln(l(x))$).

S (Baudisch & Stott 2019) is quantified as the difference in areas under the age-specific survivorship curves of a standardised survivorship curve that assumes constant mortality, and therefore has a value of 0.5, and the survivorship curve in question:

$$S = 0.5 - \int_{\alpha}^{\omega} H(x) \quad \text{equation 4}$$

Theoretically, the maximum and minimum values of the second term in equation 4 are 1 and 0 respectively. The value of S is therefore bound between -0.5 and 0.5. If most mortality occurs later in life, $S > 0$, individuals in the population display actuarial senescence. On the contrary, if $S < 0$, the risk of mortality declines with age and the individuals in the population escape actuarial senescence. Values of $S \sim 0$ indicate negligible senescence, where risk of mortality remains relatively constant with age. We determined a bound around zero to infer which values of S should be considered as negative, negligible, or positive senescence respectively for the species in our dataset. For both animals and plants separately, we assumed that the root mean squared difference between a species' value of S and zero is less than or equal to some value, ϵ , such that:

$$\sqrt{\frac{\sum (S(i) - 0)^2}{n}} \leq \epsilon$$

Where $S(i)$ is the value of s for species i , and n is the total number of species in our dataset which are animals (80) or plants (395), respectively. We quantified bounds of $-0.109 \leq S \leq 0.109$ for animals and $-0.129 \leq S \leq 0.129$ for plants. For each taxonomic kingdom, values of S that fall within the bound are considered not different from zero and therefore categorised as negligible senescence. The inequality assumes no statistical distribution of the values of S across species.

The metric of actuarial senescence, S measures the spread of mortality throughout the life course of a cohort, but does not distinguish between extrinsic and intrinsic causes. If, however, mortality is biased towards the latter life stages ($S > 0$), *i.e.* older age, then this is indicative of older ages classes succumbing to mortality at a greater rate relative to their younger counterparts. Whether the ultimate cause of death is interal or external is irrelevant, S merely describes which age classes are more vulnerable to such mortality. If older ages classes are more vulnerable to mortality then this is indicative of demographic actuarial senescence.

Phylogenetic analyses for actuarial senescence

After quantifying each species' rate of actuarial senescence, we accounted for the phylogenetic relatedness of the species studied to determine the influence of a species' evolutionary history on its value of S . To explore the effects of phylogenetic relationships between the species included in this study, we obtained animal and plant phylogenies from different sources. The plant phylogeny was obtained using the *V.PhyloMaker* R package (Jin and Qian 2019). *V.PhyloMaker* allows to build a rooted and time-calibrated phylogeny using a species list, based on already built plant phylogenies (Smith & Brown 2018; Zanne *et al.* 2014). The animal phylogeny was produced using the *datelife* R package (Sánchez-Reyes & O'Meara 2019), a service that uses publically accessible phylogenetic source data to build a chronogram – rooted and time-calibrated tree - given an input phylogeny that we sourced from the Open Tree of Life (Hinchliff *et al.* 2015). In some cases, for both plant and animal phylogenies, we detected polytomies (*i.e.* >2 species with the same ancestor), which can interfere in our phylogenetic signal analyses (see Revell 2012). Polytomies were resolved using the function “multi2di” from *ape* package (Paradis, Claude & Strimmer 2004), which transforms polytomies into a series of random dichotomies with one or several branches of length zero. Trees were visualised using the *ggtree* R package (Yu *et al.* 2017).

To evaluate the role of phylogenetic relatedness in determining the patterns of variation of actuarial senescence we estimated Pagel's λ (Pagel 1999). This metric is an index bounded between zero and one, where values ~ 0 indicate that the evolutionary history of the species explains little about the variation of the trait measured, and values ~ 1 suggest that the evolutionary history of species fully explains the observed variation of their traits. To estimate Pagel's λ we used the R package *phytools* (Revell 2012). A full summary of the phylogenetic signals obtained for each of the four monophyletic groups can be found in the Supplementary Information (Table S3).

Age-specific reproduction analysis

We calculated reproductive age-trajectories for the species in our analysis to investigate whether reproductive trajectories matched patterns of actuarial senescence. Age-specific reproduction ($m(x)$) was calculated following Caswell (p. 118-21) (2001). Briefly, the proportional structure of the cohort at age x is given by

$$\mathbf{p}(x) = \frac{U^x \mathbf{e}_j}{\mathbf{e}^T U^x \mathbf{e}_j} \quad x = 0, 1, \dots \quad \text{equation 4}$$

The total sexual reproductive output per individual at age x is given by

$$m(x) = \mathbf{e}^T \mathbf{F} \mathbf{p}(x) \quad \text{equation 5}$$

For the remaining 463 species that are not displayed in Figure 2, the $l(x)$ and $m(x)$ trajectories are found in the Supplementary information (Fig.S3).

Results

Actuarial senescence is not the rule

The majority of animal species in our study (59/80) display no change in their risk of mortality with age. In particular, *increases* in the risk of mortality with age are especially scarce across invertebrates in our data, with the water flea (*Daphnia pulex* – Fig.1) as the sole example of positive actuarial senescence. The remaining 14 invertebrate species display negligible actuarial senescence, as in the case of the long-wristed hermit crab (*Pagurus longicarpus* – Fig. 1), or even negative actuarial senescence, for example the sea whip (*Leptogorgia virgulata* – Fig 1), actuarial senescence. Across vertebrates, 72% of species, including the guppy (*Poecilia reticulata* – Fig 1), display no change in risk of mortality with age (Fig. 1; Table S1). Positively senescent species, however, are more common in vertebrates (18%;12/65) than invertebrates (6%;1/15); these species are primarily mammals (75%; Table S1) such as the moose (*Alces alces* – Fig.1). Further species such as the eastern mud turtle (*Kinosternum subrubrum*) and two birds: the white-tailed eagle (*Haliaeetus albicilla*) and Heermann's gull (*Larus heermanni*) also display positive actuarial senescence. The six negatively senescent vertebrate species span across mammals (3), ray-finned fish (1), and reptiles (2) (e.g. the South American river turtle *Podocnemis expansa* – Fig.1; Table S1).

The majority of examined plant species also display negligible senescence. Indeed, only 2% of 375 plant species exhibit positive senescence, including the scots pine (*Pinus sylvestris*) and the great laurel (*Rhododendron maximum*; Fig. 1). Approximately 23% of angiosperms show a decreasing risk of mortality with age (e.g. *Opuntia rastrera* – Fig. 1), compared to 40% of gymnosperm species (e.g. *Pinus lambertiana* – Fig 1). Overall, 98% of our studied plant species do not undergo actuarial senescence.

Patterns of senescence are driven by phylogenetic relatedness in plants, but not animals.

Estimates of phylogenetic signal on actuarial senescence were not significant across the pool of examined animals (Fig S1; Table S3). Specifically, Pagel's λ (17) was not significantly different from zero for the both the full phylogenetic analysis across animals ($\lambda = 0.22$, $p = 0.18$), and also when considering vertebrates and invertebrates separately (Table S3). These results indicate that the patterns of senescence across animals cannot be explained by phylogenetic relatedness, under a brownian model of evolution. On the other hand, phylogenetic relatedness plays some role in senescence patterns across plants (Fig. S2; Table S3). A full analysis including both angiosperms and gymnosperms showed a Pagel's λ of 0.31 ($p < 0.001$), most likely due to the significant phylogenetic signal in angiosperms ($\lambda = 0.27$, $p = 0.001$). Independent phylogenetic analysis of actuarial senescence across gymnosperms raised a non-significant signal ($\lambda = 0.27$, $p = 0.08$), likely due to the small sample size of gymnosperms ($n = 25$ species).

Patterns of reproduction and actuarial senescence are somewhat independent across animals and plants.

Patterns of $m(x)$ are diverse and not always determined by whether the examined species display or escape actuarial senescence (Fig.2; Fig.S3). In plants, for example, both the scots pine and the great laurel display actuarial senescence (Fig. 1), but their reproductive outputs do not decline with age (Fig. 2). This pattern is contrasting to both examples of animals displaying positive senescence, where the moose (*Alces alces*) and water flea (*Daphnia pulex*) also display reproductive decline with age (Fig. 2).

The patterns of actuarial senescence and reproductive output do not always align in species that display negligible or negative senescence. The flatweed provides an example of

where both components of senescence align with both species exhibiting negligible senescence and a relatively constant $m(x)$ trajectory. The long-wristed hermit crab and the sugar pine, however, also display negligible senescence but have increasing $m(x)$ trajectories. It appears from our study species that both components of senescence can sometimes follow variable, independent, trajectories.

Discussion

The emerging landscape of our study of 475 species indicates that (i) senescence is not inescapable across the Tree of Life, (ii) senescence is not inevitable in species with a germ-soma barrier, and (iii) senescence is prevalent in some species without a clear germ-soma barrier. These findings are in direct contradiction with the predictions of universal senescence, or universal senescence in species that separate germ line and soma (Hamilton 1966; Kirkwood 1977). Our comparative ageing analyses, the largest to date, provides a clear view of the discrepancy between senescence theory and data. Importantly, our results do not provide evidence against any evolutionary mechanism of senescence, mutation accumulation (Medawar 1952) or antagonistic pleiotropy (Williams 1957) for example. Rather, our results display that not all populations succumb to a weakening of the force of natural selection with age. We now need to understand the mechanisms behind this variation of age-trajectories of mortality and reproduction, and why some species succumb to senescence whilst others appear to escape its forces (Baudisch & Vaupel 2012; Jones & Vaupel 2017).

Considering first the analysis of actuarial senescence, most of our study species display no significant change in the risk of mortality with age (Fig. 1; Table S1; Table S2). Generally, this finding supports the original conundrum that the presence of senescence is inherently paradoxical. If natural selection is a fitness-maximising agent (Hamilton 1964), then one would *a priori* not expect the evolution of a phenomenon so seemingly detrimental. Perhaps a

determination to label senescence as a universal force is born out of its obvious effects in humans when, in reality, it is mostly absent from nature (Fig. 1). Some authors have suggested that this may be due to organisms not living long enough in the wild, but see Box 1 for why this is not the case. In addition, while our analyses include species that display both positive and negative actuarial senescence (Fig.1; Table S1; Table S2), not all of these can be explained under the classical evolutionary framework. For example, although a small proportion, seven angiosperms – species with no clear germ-soma separation – display positive actuarial senescence (Table S2). On the other hand, three mammals, species with a clear germ-soma barrier, display negative actuarial senescence (Table S1).

Our results also show that age-trajectories of mortality and reproduction are often independent (Fig. 2; Fig. S3). For each species in our study, we only consider a single studied population, and so this decoupling is not be an artefact of intra-specific variation across different populations. It follows that species may display actuarial senescence, but not reproductive senescence, and *vice versa*. Thus, we urge future work to consider that senescence is a two-component phenomenon of which, as displayed here, both are not destined to the same fate. To fully divulge the senescence profile of a species, one must consider both mortality and reproduction.

Studies on reproductive senescence are sparser than their actuarial senescence counterparts. Some important longitudinal investigations into reproductive senescence have been conducted suggest that rates of reproduction, like mortality hazards, can also both increase or decrease with age (Jones *et al.* 2009; Jones *et al.* 2014; Lemaître & Gaillard 2017; Barneche *et al.* 2018). Our results support observations that reproductive patterns are variable across species (Fig. 2; Fig. S3). Recently, Baudisch & Stott (2019) have developed a methodology to quantify reproductive senescence patterns using a metric parallel to the one we use here, S , for

actuarial senescence. It would now be interesting to see to what extent patterns of actuarial and reproductive senescence co-vary, both within and between species.

In general, our results display the discrepancy between the predictions of the classical evolutionary framework of ageing and empirical data. We suggest that the theory needs to be widened to better encompass the biology of a more diverse range of taxa. For example, the models of the classical evolutionary framework are purely age-structured, yet, in some species, demographic patterns of survival and reproduction may be influenced equally or even more by factors besides age (Caswell 2001), such as size for example. Indeed, it has been shown that the force of selection does not always decline with age for species where size is a better predictor (Caswell, H. & Salguero-Gómez 2013), and empirical examples can be found in sessile, modular species (Baudisch *et al.* 2013; Hughes 1984), or species with indeterminate growth forms (Vaupel *et al.* 2004). Perhaps not by coincidence, in our analyses, 98% of studied plants and all of our studied corals show no increase in risk of mortality with age (e.g. *Paramuricea clavata*; Fig. 1; Table S1; Table S2).

Many of the predictions made explicit from the classical framework of ageing have, until recently, long stood the test of time. Higher rates of extrinsic mortality, *i.e.* deaths due to the background environment, are expected to accelerate rates of senescence, whereas juvenile mortality is predicted not to play a role in the evolution of senescence (Williams 1957). Theoretical advancements, however, have shown that, for extrinsic mortality to have a significant effect on the evolution of senescence, it must be age-dependent (Caswell 2007). Also, by biasing the stable age distribution of a population towards younger ages, high birth rates can also reduce the strength of selection with age (Wensink, Caswell & Baudisch 2017). The strength of selection at a given age is dependent on both the abundance of individuals in a given age class *and* the respective reproductive value of that age class (Wensink, Caswell & Baudisch 2017). Following this logic, some species that display senescence yet retain high

reproduction at old ages (e.g. *Pinus sylvestris*; Fig. 2) may have a stable age distribution biased towards younger individuals. This outcome would render selection too weak to promote an escape from senescence. Ultimately, how the environment shapes patterns of birth and deaths will dictate both the reproductive value of age classes and the stable age-distribution of the classes (Wensink, Caswell & Baudisch 2017). In turn, the resulting dynamics of these pressures will affect the relative strengths of age-specific selection gradients (Lande 1982) for mortality and reproduction, and therefore, the patterns of senescence.

Finally, we have only considered patterns of survival and reproduction with respect to effects on the focal individual. If, however, an individual's survival and/or reproduction affects the fitness of others and the interacting individuals are relatives, selection on the demographic age trajectories will also be weighted by these effects (Bourke 2007). In our study, the killer whale (*Orcinus orca*) experiences negligible actuarial senescence (Table S1; Fig. S3). Killer whales are an exemplar where post-reproductive survival is hypothesised to have evolved due to the positive effects individuals can have on the survival and reproduction of grand-offspring, *i.e.* the 'grandmother hypothesis' (Hawkes *et al.* 1998; Johnstone & Cant 2010; Natrasset *et al.* 2019). Although, on the other hand, post-reproductive survival is also suggested to have evolved because of similar benefits in Elephants, yet the Asian elephant (*Elaphus maximus*) population in our study displays positive actuarial senescence (Table S1; Fig. S3). Our study is not suited to provide a detailed account of the effects of sociality on the evolution of senescence. Further evidence, however, is beginning to accrue elsewhere that it may play an important role beyond the remits of 'grandmothering' (Berger *et al.* 2018; Hammers *et al.* 2019; Natrass *et al.* 2019).

In summary, the emerging picture of senescence across multicellular organisms is at odds with the widely cited predictions of universal senescence from the classical evolutionary framework (Medawar 1952; Williams 1957; Hamilton 1966; Kirkwood 1977). We propose that

the field would benefit significantly from shiften attention towards the underlying mechanisms allowing species to *escape* from senescence. We expect the greatest progress to be made by researchers honing their focus to widening the classic evolutionary theories to a framework not solely focused on age, but instead inclusive of the aforementioned factors and with a special focus on actuarial and reproductive senescence as potentially differing trajectories. Most ageing research likely stems from human desire to increase human health and life span (Jones & Salguero- Gómez 2017). This desire requires understanding the variation in patterns of senescence across the tree of life. For now, senescence remains an unsolved problem of biology.

Acknowledgements

We thank P. Barks, I. Stott & M. Bonsall for key input in the analyses of demographic entropy. M.R was supported by funding from the Biotechnology and Biological Sciences Research Council([grant number BB/M011224/1]). P.C. was supported by a Ramón Areces Foundation Postdoctoral Scholarship. This research emerged through funding by NERC NE/M018458/1 to R.S-G. We thank the hundreds of population ecologists who have contributed with data to COMPADRE & COMADRE.

References

- Medawar, P. B. *An unsolved problem of biology* (H. K. Lewis, 1952).
- Hamilton, W. D. The moulding of senescence by natural selection. *J. Theor. Biol.* **12**, 12–45 (1966).
- Williams, G. Pleiotropy, natural selection, and the evolution of senescence. *Evolution* **11**, 398–411 (1957).
- Kirkwood, T. B. L. Evolution of ageing. *Nature* **270**, 301–304 (1977).
- Baudisch, A. *et al.* The pace and shape of senescence in angiosperms. *J. Ecol.* **101**, 596–606 (2013).
- Jones, O.R. *et al.* Diversity of ageing across the tree of life. *Nature* **505**, 169–174 (2014).
- Finch, C. E. *Longevity, Senescence and the Genome* (Univ. Chicago Press, 1994).
- Vaupel, J. W., Baudisch, A., Doëlling, M., Roach, D. A. & Gampe, J. The case for negative senescence. *Theor. Popul. Biol.* **65**, 339–351 (2004).
- Jones, O.R., & Vaupel J.W. Senescence is not inevitable. *Biogerontology* **18**, 965–971 (2017).
- Baudisch, A. & Vaupel, J.W. Getting to the root of aging. *Science* **338**, 618–619 (2012).
- Salguero-Gómez, R. *et al.* COMADRE: a global database of animal demography. *J. Anim. Ecol.* **85**, 371–385 (2016).
- Salguero-Gómez, R. *et al.* The COMPADRE plant matrix database: an open online repository for plant demography. *J Ecol.* **103**, 202–218 (2015).
- Chiang, C. L. *The life table and its applications* (Krieger Publishing, 1984).
- Caswell, H. *Matrix Population Models* (Sinauer Associates, 2001).

447 Baudisch, A. & Stott, I. A pace and shape perspective on fertility. *Methods in Ecology and*
 448 *Evolution* **10(11)**, 1941-1951 (2019).
 449 Keyfitz, N. *Applied Mathematical Demography* (John Wiley and Sons, 1977).
 450 Baudisch, A. The pace and shape of ageing. *Methods in Ecology and Evolution* **2(4)**, 375-382
 451 (2011).
 452 Jones, O.R. *et al.* Senescence rates are determined by ranking on the fast-slow life-history
 453 continuum. *Ecology Letters*. **11(7)**, 664-673 (2009).
 454 Pagel, M. Inferring the historical patterns of biological evolution. *Nature*. **401**, 877-884 (1999).
 455 Sánchez-Reyes, L.L. & O'Meara, B.C. DateLife: Leveraging databases and analytical tools to
 456 reveal the dated Tree of Life. *BioRxiv*. (2019).
 457 Jin, Y. and Qian, H. V. PhyloMaker: an R package that can generate very large phylogenies for
 458 vascular plants. *Ecography*. **42**, 1353-1359 (2019).
 459 Hamilton, W.D. The genetical evolution of social behaviour. *J. Theor. Biol.* **7**, 1-16 (1964).
 460 Lemaître, J-F. & Gaillard, J-M. Reproductive senescence: new perspectives in the wild. *Biol.*
 461 *Rev.* **92(4)**, 2182-2199 (2017).
 462 Barneche, D.R., Robertson, D.R., White, C.R. & Marshall, D.J. Fish reproductive-energy
 463 output increases disproportionately with body size. *Science*. **360**, 642-645 (2018).
 464 Caswell, H. & Salguero-Gómez, R. Age, stage and senescence in plants. *J. Ecol.* **101**, 585–
 465 595 (2013).
 466 Hughes, T. P. Population dynamics based on individual size rather than age: a general model
 467 with a reef coral example. *Am. Nat.* **123**, 778-795 (1984).

468 Caswell, H. Extrinsic mortality and the evolution of senescence. *Trends in Ecology and*
469 *Evolution*. **22**, 173–174 (2007).

470 Wensink, M.J., Caswell, H. & Baudisch, A. The Rarity of Survival to Old Age Does Not Drive
471 the Evolution of Senescence. *Evol. Biol.* **44**, 5-10 (2017).

472 Lande, L. A quantitative genetic theory of life history evolution. *Ecology*. **63**, 607–615. (1982).

473 Bourke, A.F.G. Kin Selection and the Evolutionary Theory of Aging. *Ann. Rev. Ecol. Evo.*
474 *Syst.* **38**, 103-128 (2007).

475 Hawkes, K., O’Connell, J. F., Blurton-Jones, N. G., Alvarez, H. & Charnov, E. L.
476 Grandmothering, menopause, and the evolution of human life histories. *Proc. Natl Acad. Sci.*
477 *USA*. **95**, 1336–1339 (1998).

478 Johnstone, R.A. & Cant, M.A. The evolution of menopause in cetaceans and humans: the role
479 of demography. *Proc. R. Soc. B*. **277** 3765-3771 (2010).

480 Natrass, S. *et al.* Postreproductive killer whale grandmothers improve the survival of their
481 grandoffspring. *Proc. Natl Acad. Sci. USA* (December 2019).

482 Berger, V., Lemaître, J-F., Allainé, D., Gaillard, J-M. & Cohas, A. Early and Adult Social
483 Environments Shape Sex-Specific Actuarial Senescence Patterns in a Cooperative Breeder.
484 *Am. Nat.* **192:4**, 525-536 (2018).

485 Hammers, M. *et al.* Breeders that receive help age more slowly in a cooperatively breeding
486 bird. *Nature Communications*. **10:1301** (2019).

487 Jones, O.R. & Salguero- Gómez, R. Life History Trade-Offs Modulate the Speed of
488 Senescence. *The Evolution of Senescence in the Tree of Life*. Eds. Shefferson, R.P., Jones, O.R.
489 & Salguero-Gómez, R. (Cambridge University Press, 2017).

490 Salguero-Gómez, R. & Plotkin, JB. Matrix dimensions bias demographic inferences:
 491 implications for comparative plant demography. *Am Nat.* **176**(6), 710-72. (2010).

492 Horvitz C.C. & Tuljapurkar, S. Stage dynamics, period survival, and mortality plateaus. *Am*
 493 *Nat.* **172**, 203–215. (2009).

494 Smith, S. A., & Brown, J. W. Constructing a broadly inclusive seed plant phylogeny. *American*
 495 *Journal of Botany.* **105**(3), 302-314 (2018).

496 Zanne, A.E. et al. Three keys to the radiation of angiosperms into freezing environments.
 497 *Nature.* **506**, 89 (2014).

498 Hinchliff, C. E. et al. Synthesis of phylogeny and taxonomy into a comprehensive tree of life.
 499 *Proc. Natl. Acad. Sci.* **112**, 201423041 (2015).

500 Revell, L. J. Phytools: an R package for phylogenetic comparative biology (and other things).
 501 *Methods in Ecology and Evolution.* **3**, 217-223 (2012).

502 Paradis, E., Claude, J. & Strimmer, K. APE: Analyses of phylogenetics and evolution in R
 503 language. *Bioinformatics.* **20**, 289–290. (2004).

504 Yu, G., Smith, D., Zhu, H., Guan, Y. & Lam, T.T-Y. ggtree: an R package for visualization
 505 and annotation of phylogenetic trees with their covariates and other associated data. *Methods*
 506 *in Ecology and Evolution.* **8**(1), 28-36 (2017).

507 Comfort, A. (1979) *The Biology of Senescence*. Churchill Livingstone, London.

508 Rose, M.R. (1991) *Evolutionary Biology of Aging*. Oxford University Press, New York.

509 Hayflick, L. (2000) The future of ageing. *Nature.* **408**, 267–269.

510 Monaghan, P., Charmantier, A., Nussey, D. H., & Ricklefs, R. E. (2008). The evolutionary
 511 ecology of senescence. *Functional Ecology.* **22**(3), 371–378.

512 Nussey, D. H., Coulson, T., Festa-Bianchet, M., & Gaillard, J.-M. (2008). Measuring
 513 senescence in wild animal populations: Towards a longitudinal approach. *Functional*
 514 *Ecology*. **22**(3), 393–406.

515 Nussey, D. H., Froy, H., Lemaître, J.-F., Gaillard, J.-M., & Austad, S. N. (2013). Senescence
 516 in natural populations of animals: Widespread evidence and its implications for bio-
 517 gerontology. *Ageing Research Reviews*. **12**(1), 214–225.

518 Gaillard, J.-M. & Lemaître, J.-F. (2020). An integrative view of senescence in nature. *Functional*
 519 *Ecology*. **34**(1), 4–16.

520 Martínez A, D.E. (1998) Mortality Patterns Suggest Lack of Senescence in *Hydra*.
 521 *Experimental Gerontology*. **33**, 217–225.

522 Schaible, R., Scheuerlein, A., Dańko, N.J., Gampe, J., Martínez, D.E. & Vaupel, J.W. (2015)
 523 Constant mortality and fertility over age in *Hydra*. *Proceeding of the National Academy of*
 524 *Sciences of the United States of America*. **112**, 15701–15706.

525 Dańko, M.J., Kozłowski, J. & Schaible, R. (2015) Unraveling the non-senescence phenomenon
 526 in *Hydra*. *Journal of Theoretical Biology*. **382**, 137–149.

527 Koons, D.N. *et al.* (2014) Methods for studying cause-specific senescence in the wild. *Methods*
 528 *in Ecology and Evolution*. **5**, 924–933.

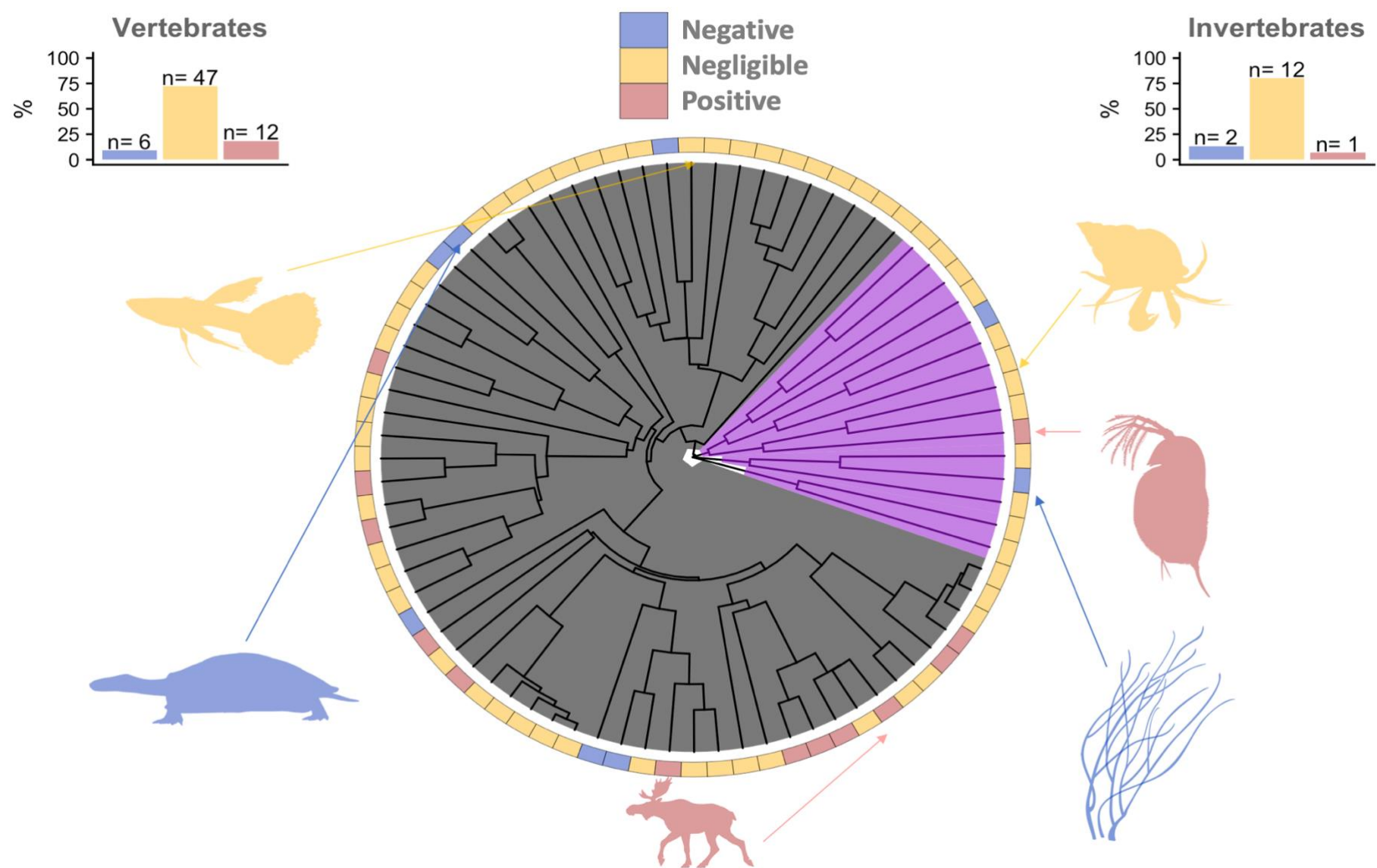
529 Moorad, J., Promislow, D. & Silvertown, J. (2019). Evolutionary Ecology of Senescence and
 530 a Reassessment of Williams’ ‘Extrinsic Mortality’ Hypothesis. *Trends in Ecology and*
 531 *Evolution*. **34**, 519–530.

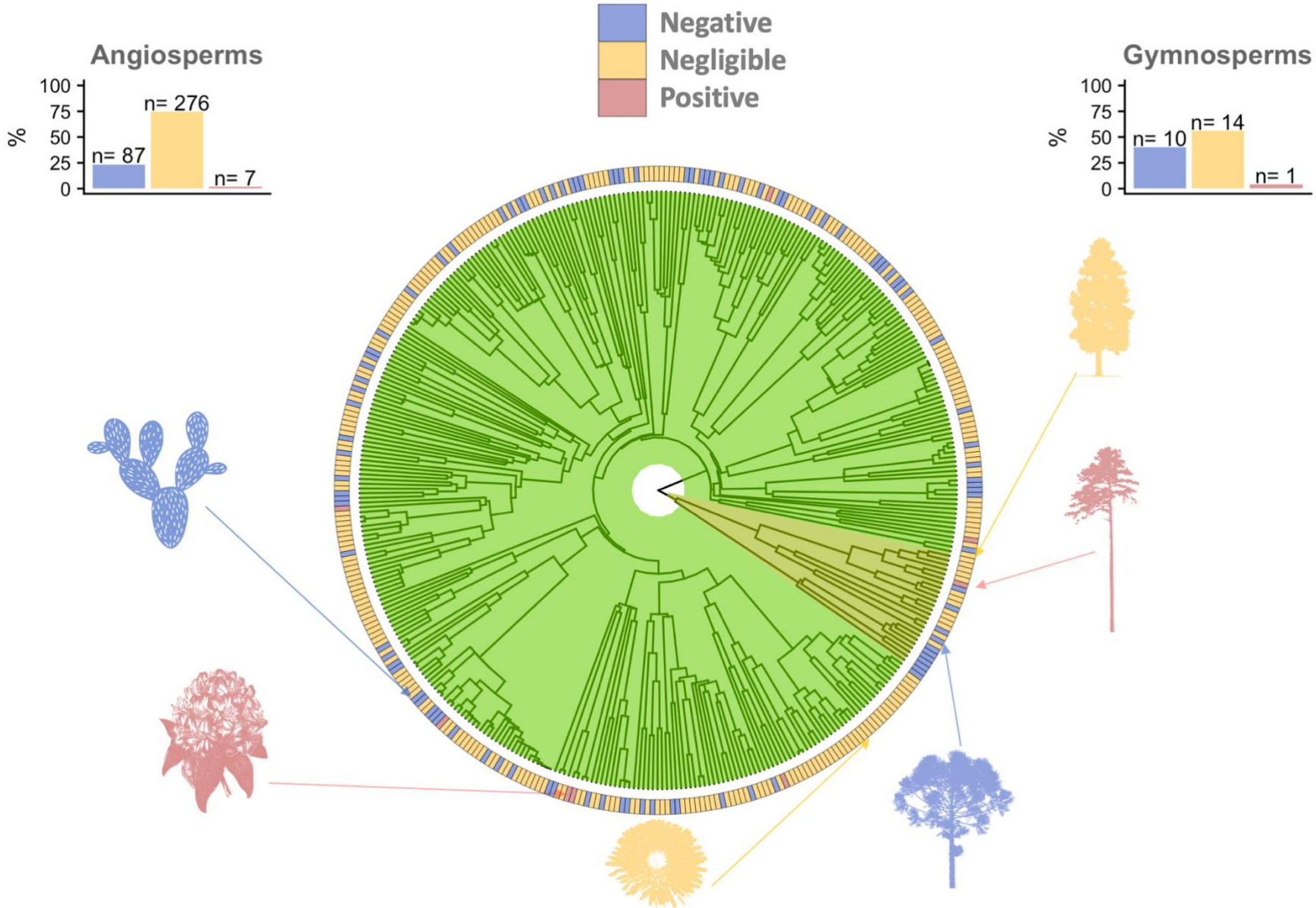
532
 533

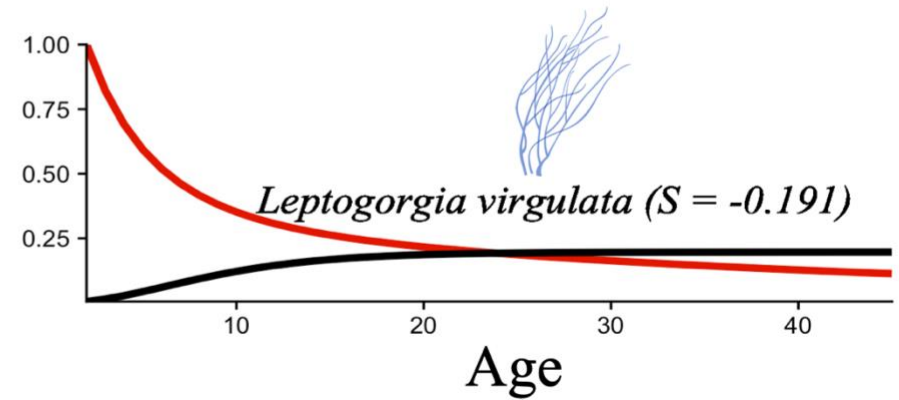
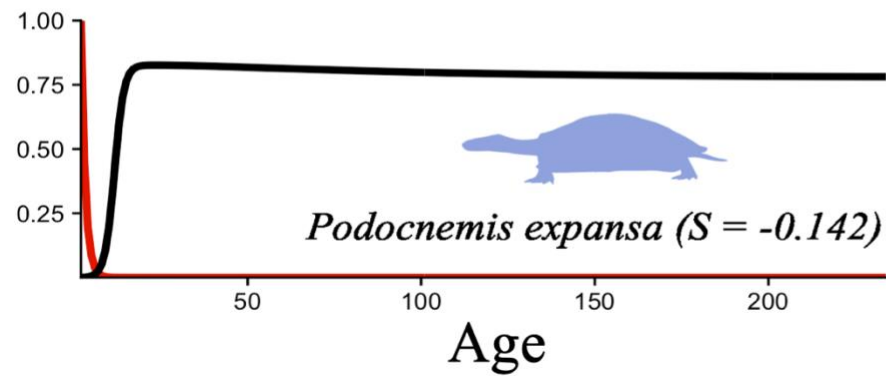
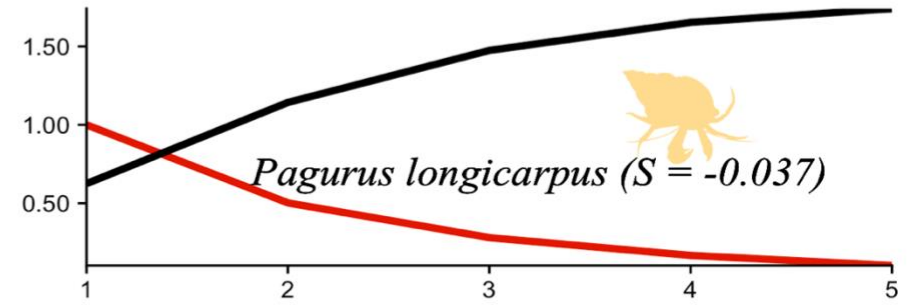
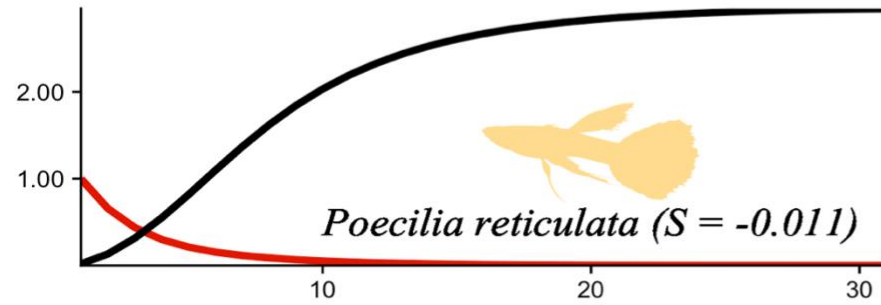
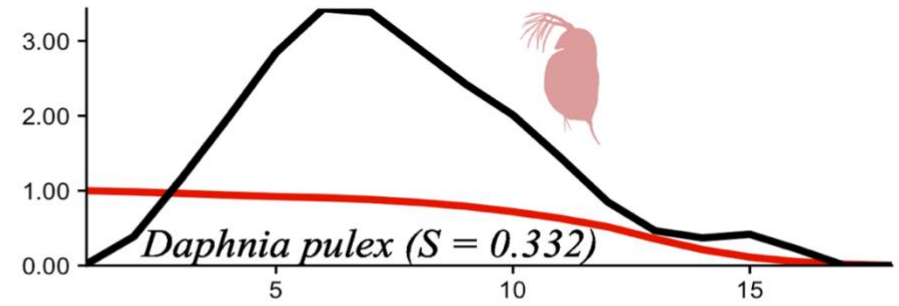
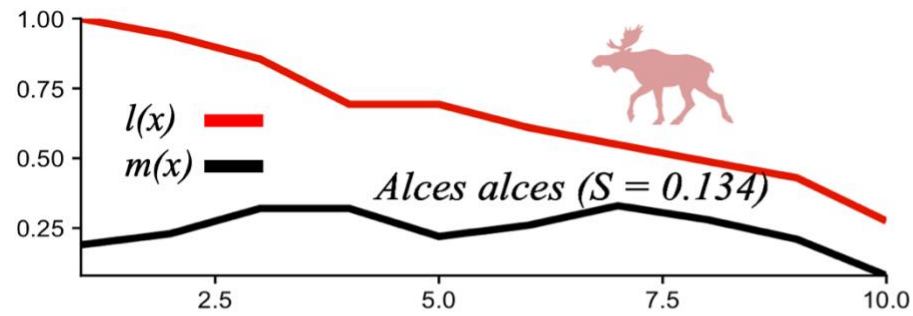
Figure Legends

Figure 1 The evolution of and escape from senescence across multicellular life. The classical evolutionary framework of ageing does not explain the evolution of actuarial senescence across our study species. Positive, negligible and negative patterns of senescence are dispersed throughout the four examined clades, with the percentages of each pattern within each clade shown in the bar charts of each of the figures. **a)** Actuarial senescence across animals. Depicted around the phylogeny are six representative species, displaying positive (red), negligible (yellow), and negative (blue) senescence from each clade. Clockwise, representing invertebrates, these species are *Pagurus longicarpus*, *Daphnia pulex* and *Leptogorgia virgulata*. For vertebrates, again clockwise, these species are *Alces alces*, *Poecilia reticulata*, and *Podocnemis expansa*. **b)** Actuarial senescence across plants. Depicted around the phylogeny are six representative species, displaying positive (red), negligible (yellow), and negative (blue) senescence from each clade. For gymnosperms, these species are *Pinus lambertiana*, *Pinus sylvestris*, and *Taxus floridana*. For angiosperms, these species are *Hypochaeris radicata*, *Rhododendron maximum*, and *Opuntia rastrera*.

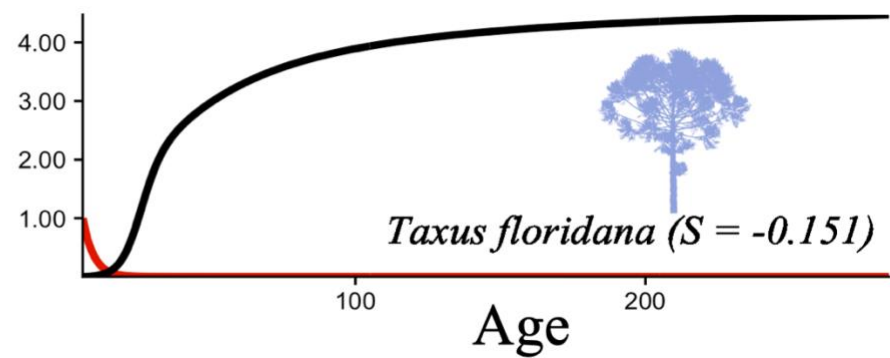
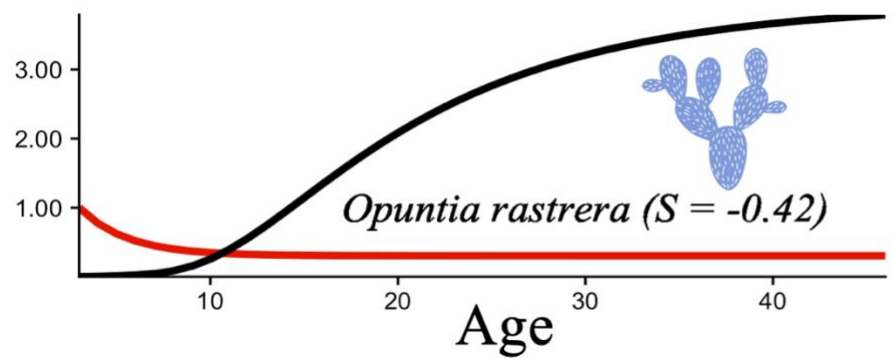
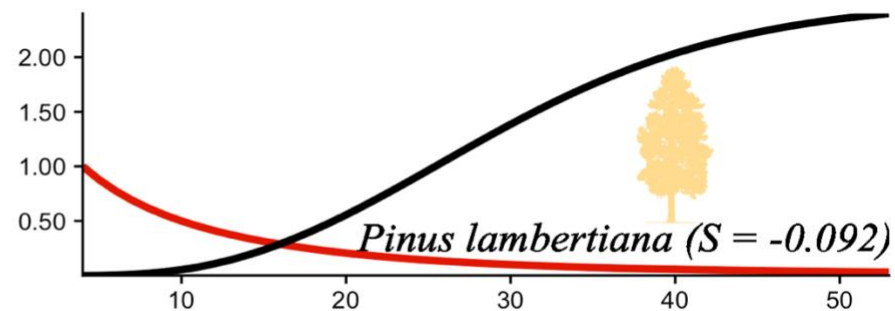
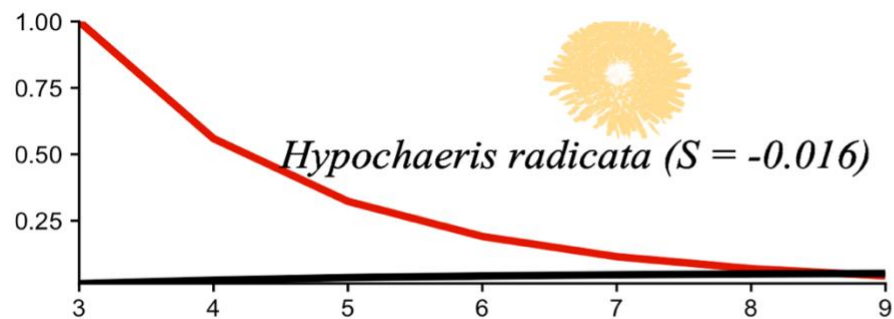
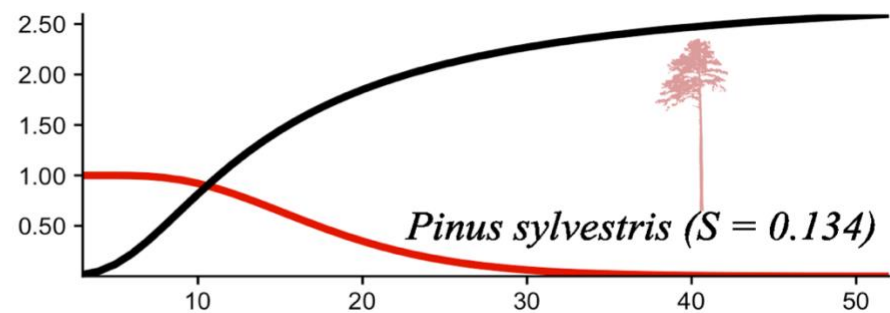
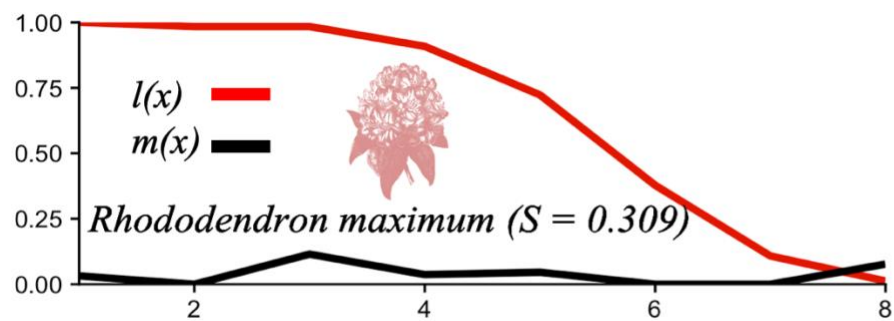
Figure 2 Age-based patterns of survivorship ($l(x)$ - red) and reproduction ($m(x)$ - black) are often decoupled, as shown for a selected subset of the examined species in Figure 1. **a)** $l(x)$ and $m(x)$ trajectories for the six selected animal species from Figure 1 and **b)** $l(x)$ and $m(x)$ trajectories for the six selected plant species from Figure 1. Species are representative of vertebrates, invertebrates, gymnosperms, and angiosperms. Trajectories are conditional upon reaching the age of maturity, at which the mature cohort is defined to have entered adulthood with a survivorship of 1. The trajectories of $l(x)$ and $m(x)$ run from age at maturity to the age at which 5% of the mature cohort is still alive.







561 2b)



Box 1: The use of wild populations in studies of senescence.

Individuals in wild populations typically face various sources of ‘extrinsic’ mortality (Medawar 1952; Williams 1957), that is, mortality due to environmental factors such as predation or disease, for example. Authors have subsequently suggested that, because extrinsic mortality is inevitably present in the wild, individuals will never attain ages at which intrinsic decline becomes apparent, and thus senescence may only be observed if organisms are maintained in ‘optimal’ conditions *i.e.* negligible extrinsic mortality (Comfort 1979; Rose 1991; Hayflick 2000). As longitudinal demographic studies have accrued, however, evidence for senescence in the wild is now widespread (Reviewed in Monaghan *et al.* 2008; Nussey *et al.* 2008; Nussey *et al.* 2013; Gaillard & Lemaître 2020), rendering the suggestion that individuals in wild populations do not attain ages at which senescence can be observed as incorrect (Nussey *et al.* 2008). Additionally, supplying an extra layer of complexity to the matter, some species, like *Hydra*, have been experimentally maintained in captive, optimal conditions and yet displayed no apparent decline in mortality or fertility (Martínez 1998; Schaible *et al.*, 2015; Dańko *et al.*, 2015).

The effect of captivity on the rate of senescence can vary significantly. Williams (1957) predicted that senescence should be greater in populations exposed to higher levels of mortality. Following this logic, captive populations should have a reduced rate of senescence relative to their wild population counterparts. However, only increased *age-dependent* mortality has the ability to alter the selection gradients on age-specific mortality and fertility (Caswell 2007; Wensink, Baudisch and Caswell 2017). Age-independent mortality, by definition of being age-independent, has no effect on the selection gradients (Caswell 2007; Koons *et al.* 2014; Wensink, Baudisch and Caswell 2017; Moorad, Promislow & Silvertown 2019) and so it cannot alone shape senescence outcomes. Therefore, William’s prediction should be amended to expecting a reduced rate of senescence in captive populations if the relieved pressure of mortality is expressed in an age-dependent manner, whereby older individuals are less likely to die. With ever-growing data sources of both wild and captive populations of the same species, a future comparative test of this prediction between wild and captive populations may prove fruitful.